

Evidence Synthesis

Number 142

Screening for Latent Tuberculosis Infection in Adults: An Evidence Review for the U.S. Preventive Services Task Force

Prepared for:

Agency for Healthcare Research and Quality
U.S. Department of Health and Human Services
5600 Fishers Lane
Rockville, MD 20857
www.ahrq.gov

Contract No. HHSA-290-2012-00015-I, Task Order No. 4

Prepared by:

RTI International–University of North Carolina Evidence-based Practice Center
Research Triangle Park, NC

Investigators:

Leila C. Kahwati, MD, MPH
Cynthia Feltner, MD, MPH
Michael Halpern, MD, PhD, MPH
Carol L. Woodell, BSPH
Erin Boland, BA
Halle R. Amick, MSPH
Rachel Palmieri Weber, PhD
Daniel E. Jonas, MD, MPH

**AHRQ Publication No. 14-05212-EF-1
March 2016**

This report is based on research conducted by the RTI International–University of North Carolina Evidence-based Practice Center (EPC) under contract to the Agency for Healthcare Research and Quality (AHRQ), Rockville, MD (Contract No. HHS-2012-00015-I, Task Order No. 4). The findings and conclusions in this document are those of the authors, who are responsible for its contents, and do not necessarily represent the views of AHRQ. Therefore, no statement in this report should be construed as an official position of AHRQ or of the U.S. Department of Health and Human Services.

The information in this report is intended to help health care decisionmakers—patients and clinicians, health system leaders, and policymakers, among others—make well-informed decisions and thereby improve the quality of health care services. This report is not intended to be a substitute for the application of clinical judgment. Anyone who makes decisions concerning the provision of clinical care should consider this report in the same way as any medical reference and in conjunction with all other pertinent information (i.e., in the context of available resources and circumstances presented by individual patients).

The final report may be used, in whole or in part, as the basis for development of clinical practice guidelines and other quality enhancement tools, or as a basis for reimbursement and coverage policies. AHRQ or U.S. Department of Health and Human Services endorsement of such derivative products may not be stated or implied.

Acknowledgments

The authors acknowledge the following individuals for their contributions to this project: Tina Fan, MD, MPH, AHRQ Medical Officer; Tracy Wolff, MD, MPH, AHRQ Associate Scientific Director; current and former members of the U.S. Preventive Services Task Force who contributed to topic deliberations; Philip LoBue, MD, FACP, FCCP, and Christine Ho, MD, Centers for Disease Control and Prevention (CDC); expert reviewers John Bernardo, MD, Boston University School of Medicine, Dick Menzies, MD, MSc, McGill University, Neil Schluger, MD, Columbia University Medical Center, and Steven Teutsch, MD, MPH, Robert Wood Johnson Foundation; federal partner reviewers Suzanne Marks, MPH, Gerald H. Mazurek, MD, and Andrew Vernon, MD, MHS, CDC; Evelyn Whitlock, MD, MPH, Director, Kaiser Permanente Research Affiliates EPC; and RTI International–University of North Carolina EPC staff Meera Viswanathan, PhD; Russell Harris, MD, MPH; Molly Howard, PharmD, BCPS, CPP; Makda Majerette, BA; Roberta C. Wines, MPH, Christiane Voisin, MSLS; Rachael Posey, MSLS; Janice Handler, BA; Jennifer Drolet, MA; and Loraine Monroe.

Structured Abstract

Purpose: To assess the benefits and harms of targeted screening and treatment for latent tuberculosis infection (LTBI) in higher-risk adults in primary care settings.

Data Sources: MEDLINE, the Cochrane Library, and trial registries through August 3, 2015; bibliographies from retrieved articles, outside experts, and reviewers.

Study Selection: Two investigators independently selected studies using a priori inclusion and exclusion criteria. We selected studies that evaluated tuberculin skin test (TST) using the Mantoux method, or tests evaluating commercial interferon gamma release assays (IGRA). We selected trials of treatment that evaluated current Centers for Disease Control and Prevention (CDC)-recommended pharmacotherapy regimens for LTBI for synthesis of benefits and harms. We excluded studies of people with underlying immunosuppression and for whom LTBI screening and treatment would be part of standard disease management by specialty care providers (e.g., individuals with HIV, history of or planned organ transplant, or planned or active use of tumor necrosis factor-alpha [TNF- α] inhibitors). We excluded poor-quality studies, studies assessing specificity in high tuberculosis (TB)-burden countries, and studies assessing harms and benefits in developing countries.

Data Extraction: One investigator extracted data and a second checked accuracy. Two reviewers independently rated quality for all included studies, using predefined criteria.

Data Synthesis: We did not identify any studies that compared screening with no screening. We included 71 studies of fair to good quality; 66 assessed test accuracy or reliability and 5 assessed benefits and harms of treatment. Pooled estimates for sensitivity of TST at both the 5-mm and 10-mm induration thresholds for positivity were 0.79; the pooled estimate at the 15-mm threshold was 0.52. Pooled estimates for sensitivity of IGRA tests ranged from 0.77 to 0.90. Estimates for specificity of TST at 5 mm varied considerably by TB burden of the study setting (0.94 to 0.97 in low TB-burden countries, 0.30 in intermediate TB-burden country). Pooled estimates for specificity at 10-mm and 15-mm thresholds were 0.97 and 0.99, respectively. Pooled estimates for specificity of IGRA tests ranged from 0.95 to 0.98. We found evidence for at least moderate interrater reliability for both TST and IGRA tests.

The best evidence on effectiveness for treatment for LTBI was from the International Union Against Tuberculosis (IUAT) trial, a large (N=27,830) good-quality randomized controlled trial (RCT) that evaluated multiple treatment durations for daily isoniazid. It found a relative risk (RR) for progression to active TB at 5 years of 0.35 (95% CI, 0.24 to 0.52) for 24 weeks of isoniazid compared with placebo (N=13,955, number needed to treat [NNT], 112). Our sensitivity analyses adding 4 RCTs that did not meet all of our eligibility criteria (e.g., compared isoniazid with placebo using a longer duration of treatment or used different doses than currently recommended) found an RR of 0.31 (95% CI, 0.24 to 0.41; I^2 , 0%; 5 RCTs, N=36,823). A head-to-head open label, noninferiority RCT that compared a combination of once-weekly rifapentine plus isoniazid for 3 months with daily isoniazid for 9 months found the combination therapy to be noninferior to isoniazid alone for preventing the development of active TB.

For harms, the IUAT trial reported an RR for hepatotoxicity of 4.59 (95% CI, 2.03 to 10.39; number needed to harm [NNH], 279) for 24 weeks of isoniazid as compared with placebo. Sensitivity analyses pooling the IUAT with 3 RCTs that used a longer duration of isoniazid yielded a similar result: pooled RR, 5.04 (95% CI, 2.50 to 10.15; I^2 , 0%; 4 RCTs, N=35,161). The RR of treatment discontinuation because of adverse effects across all treatment duration arms in IUAT was 1.50 (95% CI, 1.18 to 1.89; N=27,830; NNH, 167). For isoniazid compared with rifampin, the pooled RR for hepatotoxicity was 3.29 (95% CI, 1.72 to 6.28; I^2 , 0%; 3 RCTs, N=1,327) and the pooled RR for treatment discontinuation because of adverse events was 1.61 (95% CI, 0.57 to 4.57; I^2 , 40.0%; 3 RCTs, N=1,327).

Limitations: No test for the direct diagnosis of LTBI exists; thus, studies of test accuracy use populations of confirmed active TB subjects to establish sensitivity, and of healthy, low-risk individuals to establish specificity. Thus, applicability to other populations is uncertain. The single trial meeting all eligibility criteria that established the benefits of a currently recommended treatment (isoniazid 300 mg daily for 24 weeks) for preventing active TB was published over 30 years ago and was conducted among subjects with pulmonary fibrotic lesions; whether it may overestimate the benefits of treatment for populations with lower risk for progression is not clear. No trials evaluated the effectiveness (compared with placebo) of regimens other than isoniazid. Contemporary treatment studies have not included placebo arms; when available, information on benefits and harms of newer treatments are derived from comparative studies (vs. isoniazid). The evidence on harms is limited by heterogeneous specification of outcomes across studies. This review is not applicable to the highest-risk individuals, for whom testing and treatment is considered part of disease management or public health surveillance.

Conclusions: We did not find any studies evaluating the direct benefits and harms of screening for LTBI in the adult populations and settings included in this review. Both types of currently available tests (TST and IGRAs) are moderately sensitive and, within low TB-burden countries, highly specific. Isoniazid treatment reduces the risk of progression to active TB for people with LTBI and pulmonary fibrotic lesions. The evidence is limited or not available for other regimens and outcomes (e.g., deaths due to TB, all-cause mortality) among the populations included in this review. Isoniazid is associated with higher rates of hepatotoxicity than placebo and rifampin regimens.

Table of Contents

Chapter 1. Introduction	1
Scope and Purpose	1
Condition Definition	1
Etiology and Natural History	1
Risk Factors	2
Prevalence and Burden	3
Rationale for Screening	4
Screening Strategies	4
Treatment Approaches	6
Current Clinical Practice in the United States	6
Previous USPSTF Recommendation	7
Chapter 2. Methods	8
Key Questions and Analytic Framework	8
Data Sources and Searches	8
Study Selection	9
Quality Assessment and Data Abstraction	10
Data Synthesis and Analysis	10
Expert Review and Public Comment	12
USPSTF Involvement	12
Chapter 3. Results	13
Literature Search	13
Results by Key Question	13
Key Question 1. Direct Evidence for Targeted Screening for Latent Tuberculosis Infection	13
Key Question 2a. Accuracy and Reliability of Screening Tests	13
Key Question 2b. Accuracy and Reliability of Sequential Screening Strategies	17
Key Question 3. Benefits of Treatment for LTBI	17
Key Question 4. Harms of Screening for LTBI	19
Key Question 5. Harms of Treatment for LTBI	19
Chapter 4. Discussion	24
Summary of Evidence	24
Evidence for Benefit and Harms of Screening	24
Accuracy and Reliability of Screening Tests	24
Benefits and Harms of Treatment for LTBI	25
Hypothetical Outcomes of a Screening Program	26
Limitations of the Review	28
Future Research Needs	29
Conclusion	30
References	31

Figures

Figure 1. Analytic Framework: Screening for Latent Tuberculosis Infection in Adults

Figure 2. Preferred Reporting of Systematic Review and Meta-Analysis (PRISMA) Tree

Figure 3. Individual Study and Pooled Estimates (Where Applicable) of Sensitivity for Various Thresholds of Tuberculin Skin Test for Tuberculosis Infection

Figure 4. Individual Study and Pooled Estimates (Where Applicable) of Sensitivity for Interferon Gamma Release Assay Tests for Tuberculosis Infection

Figure 5. Individual Study and Pooled Estimates (Where Applicable) of Specificity for Various Thresholds of Tuberculin Skin Test and Interferon Gamma Release Assay Tests for Tuberculosis Infection

Figure 6. Isoniazid Compared With Placebo, Relative Risk of Developing Active TB in the IUAT Trial

Figure 7. Isoniazid Compared With Placebo, Relative Risk of Developing Active TB: Sensitivity Analysis Including Data From the IUAT Trial and Four Additional RCTs

Figure 8. Isoniazid Compared With Placebo, Relative Risk of Developing Hepatotoxicity in the IUAT Trial

Figure 9. Isoniazid Compared With Rifampin, Relative Risk of Developing Hepatotoxicity: Data From the Three RCTs

Figure 10. Isoniazid Compared With Rifampin, Relative Risk of Treatment Discontinuation Due to Adverse Events: Data From Three RCTs

Tables

Table 1. Summary of Sensitivity Estimates for Various Thresholds of Tuberculin Skin Test and Interferon Gamma Release Assays Among Patients With Bacteriologic-Confirmed TB

Table 2. Summary of Specificity Estimates for Various Thresholds of Tuberculin Skin Test and Interferon Gamma Release Assays Among Healthy Subjects Without TB Exposures or Risks

Table 3. Summary of Evidence of Accuracy and Reliability of TST or IGRA Tests for Targeted Screening of Latent Tuberculosis Infection—KQ 2a

Table 4. Summary of Evidence of Latent Tuberculosis Infection CDC-Recommended Pharmacotherapy Treatment Regimens to Reduce Active TB Incidence, Morbidity, Mortality, Transmission, or Improve Quality of Life—KQ 3

Table 5. Summary of Evidence of Harms Associated With CDC-Recommended Pharmacotherapy Treatment Regimens for Latent Tuberculosis Infection—KQ 5

Table 6. Projected 5-Year Outcomes of Screening 100,000 Asymptomatic Adults for Latent Tuberculosis Infection

Appendixes

Appendix A. LTBI Prevalence and Treatments

Appendix B. Methods

Appendix C. Excluded Studies

Appendix D. Evidence Tables

Appendix E. Quality Ratings

Appendix F. KQ 2 Supplemental Analyses

Appendix G. KQ 5 Supplemental Analyses

Chapter 1. Introduction

Scope and Purpose

The U.S. Preventive Services Task Force (USPSTF) last made a recommendation on latent tuberculosis (TB) screening in 1996. With new tests and treatments for latent TB infection (LTBI), an updated assessment of the evidence to inform recommendations on screening is warranted.

The purpose of this report is to systematically evaluate the current evidence on targeted screening and treatment of LTBI for populations and settings relevant to primary care in the United States. In this report, we summarize the evidence on the benefits and harms of screening for LTBI in selected high-risk adult populations, the test characteristics of Food and Drug Administration (FDA)-approved screening tests, and the benefits and harms of Centers for Disease Control and Prevention (CDC)-recommended treatments for LTBI. This review also identifies key gaps in this scientific literature.

This review was scoped to provide the USPSTF with answers to key questions needed to support making a recommendation about LTBI screening in asymptomatic, generally healthy adults in settings relevant to primary care. Thus, the review does not focus on the testing of close contacts of people with active TB, or medically vulnerable populations for whom LTBI testing is considered part of disease management. Further, this review does not focus on important issues related to TB epidemiology, the public health infrastructure for TB, or adherence with all steps involved in testing and treatment.

Condition Definition

TB is a disease caused by *Mycobacterium tuberculosis* (MTB) that is spread through airborne transmission. TB usually affects the lungs, but it can also affect other parts of the body, such as the brain, kidneys, or spine. When a person with active pulmonary TB coughs or sneezes, droplet nuclei containing MTB are expelled into the air. If another person inhales air containing these droplet nuclei, three conditions are possible: clearance of the organism; onset of active disease (primary TB disease); or latent infection without signs, symptoms, or radiographic or bacteriologic evidence of TB disease.¹ Individuals with LTBI are not infectious to others. LTBI can later reactivate when previously dormant MTB, seeded at the time of exposure, proliferate and progress to cause active TB disease.

Etiology and Natural History

After exposure to MTB, approximately 30 percent of individuals are thought to develop LTBI as diagnosed based on a positive tuberculin skin test (TST).^{2,3} Five to 10 percent of healthy (immunocompetent) people with a positive TST will progress from LTBI to active TB disease (referred to as reactivation) in their lifetime. This estimate is based on epidemiologic data and

data from placebo arms of treatment trials conducted before treatment for LTBI was routinely recommended.^{4,5} However, this range underestimates the risk of progression to active TB for some patients and overestimates the risk for others, because risks vary greatly according to age, the size of the TST reaction, and the presence or absence of specific medical conditions.⁶

A recently published observational study of contacts of people with active TB in Amsterdam who were diagnosed with LTBI between 2002 and 2011 reported a 5-year risk of incident TB of 2.4 percent (95% CI, 1.2 to 4.7) among those who did not take preventive therapy.⁷ A recent report using 2006–2008 United States data estimated the rate of TB reactivation among individuals with LTBI as 0.084 cases per 100 person-years (95% CI, 0.083 to 0.085).⁸ Among people who tested positive versus negative for HIV, rates were 1.82 (95% CI, 1.74 to 1.89) and 0.073 (95% CI, 0.070 to 0.075) cases per 100 person-years, respectively. Reactivation rates were higher among foreign-born people (0.098 cases/100 person-years; 95% CI, 0.096 to 0.10) than among those born in the United States (0.082 cases/100 person-years; 95% CI, 0.080 to 0.083).

Risk Factors

People may be considered “high risk” for LTBI for several reasons.^{1,4,9} Some may be high risk because of increased likelihood of exposure to active TB. Others may be high risk because of increased likelihood of latent infection if exposed because of medical conditions or other factors that influence the immune system. Some people with increased chance of latent infection due to underlying immune factors may also have an increased risk of LTBI reactivation. The estimates for prevalence of LTBI among some higher-risk populations and estimates for risk of reactivation of LTBI are frequently based on older studies that may not correspond to current risks and practice patterns.

Risks for incident LTBI associated with increased exposure to active TB include being born outside the United States, having lower household income or less than a high school education, being male, being a member of a racial/ethnic minority population, being an active smoker, and living with someone with active TB.¹⁰ Substantially increased exposure to active TB has also been observed among homeless individuals, injection drug users, people with HIV, and residents or employees of a high-risk congregate setting (e.g., prison, long-term care facility, hospital, homeless shelter).⁴

Risks associated with a higher likelihood of latent infection if exposed, or reactivation to active TB, or both, vary.⁴ These risks include HIV infection, injection drug use, radiographic evidence of prior healed TB, low body weight (10% below ideal), and certain medical conditions (e.g., silicosis, poorly controlled diabetes mellitus, chronic renal failure, gastrectomy, solid organ transplant, smoking, head and neck cancer, and conditions that require prolonged use of corticosteroids or other immunosuppressive agents).^{1,4,6,9} However, there is uncertainty in the estimated risk of developing active TB for people with these conditions. One review found that estimates of the relative risk (RR) of developing active TB for people with medical conditions that impair the host’s immune system are either lower than expected or have significant methodological flaws (e.g., failure to distinguish among the risk of exposure, risk of infection, and risk of reactivation; small sample sizes).⁶ This same review offers RR estimates for reactivation to active TB for a number of high-risk populations; the two populations most

relevant to primary care settings are Hispanic patients with poorly controlled diabetes (RR, 1.7; 95% CI, 1.5 to 2.2) and smokers (RR 1.5 , 95% CI, 1.1 to 2.2).¹¹

Prevalence and Burden

The prevalence and burden of active TB disease affects the prevalence and burden of LTBI. TB is a substantial health issue globally, with nearly 9 million cases of active TB and 1.5 million TB-related deaths worldwide in 2013.¹² In the United States, active TB is a much more limited health problem, with cases declining in recent decades. In 2014, a total of 9,412 new active TB cases were reported in the United States, corresponding to an incidence rate of 3.0 cases per 100,000 population.¹³ There were 563 deaths from TB disease (0.2 deaths per 100,000 population) in the United States in 2011, the most recent year for which these data are available.¹⁴ Based on 2006 data, 32 percent of deaths among active TB cases occurred in people with HIV infection, though this population represented only 12 percent of active TB cases.¹⁵ The proportion of TB deaths occurring among HIV-infected people compared with non-HIV-infected people is difficult to estimate because deaths among coinfecting people are typically attributed in vital statistics to HIV infection, as opposed to TB disease. Further, HIV status is unknown in some active TB cases.^{16,17} In 2014, the prevalence of unknown HIV status among TB cases was 14 percent and the prevalence of HIV among cases of active TB with known HIV status was 6.3 percent.¹³

Among individuals with known national origin, 6,181 active TB cases were among foreign-born individuals (66.5% of all cases) in 2014, for a rate of 15.3 cases per 100,000 population compared with 1.1 cases per 100,000 population among U.S.-born individuals.¹³ Among active TB cases ages 15 or older, 5.5 percent were homeless, 2.2 percent were long-term care facility residents, and 4.2 percent were in a correctional facility.¹³ Active TB rates also vary by race and ethnicity; rates per 100,000 population in 2013 were 0.7, 5.0, 18.7, 5.4, 11.3, and 5.4 among non-Hispanic whites, Hispanics or Latinos, Asians, non-Hispanic blacks or African Americans, Native Hawaiians or other Pacific Islanders, and American Indians or Alaska Natives, respectively.¹⁸ The incidence of active TB varies significantly by geographical location (state) within the United States: California, Texas, New York, and Florida combined accounted for more than half of all U.S. TB cases reported in 2014.¹³

Estimating the prevalence of LTBI overall and among higher-risk groups is challenging because no direct test for latent MTB exists and latent infection is not reportable to CDC's National Notifiable Disease Surveillance System.¹⁹ Unlike active TB disease, which is diagnosed on the basis of clinical signs and symptoms confirmed by bacteriologic or molecular identification of MTB from fluid or tissue specimens, existing tests for latent infection measure memory T cell response, an indirect measure of host sensitization to MTB.⁹ In general, estimates of the prevalence of LTBI are based on studies using tuberculin skin testing (TST) and/or interferon gamma release assay (IGRA) to define infection.

The largest prevalence studies use data from the National Health and Nutrition Examination Survey (NHANES), a nationally representative sample of the civilian, noninstitutionalized U.S. population, to estimate the prevalence of LTBI based on an induration ≥ 10 mm on TST or a

positive IGRA. Using 2011-2012 NHANES data, the population prevalence of LTBI among individuals aged 6 years or greater is 4.7 percent (95% CI, 3.4 to 6.3) based on a positive TST alone, 5.0 percent (95% CI, 4.2 to 5.8) based on positive IGRA alone, and 2.1 percent (95% CI, 1.5 to 2.8) based on a positive TST and IGRA. Among the foreign-born U.S. population aged 6 years or greater, the prevalence of LTBI is 20.5 percent (95% CI, 16.1 to 25.8) based on a positive TST alone, 15.9 percent (95% CI, 13.5 to 18.7) based on a positive IGRA alone, and 9.3 percent (95% CI, 7.4 to 11.7) based on a positive TST and IGRA.²⁰ Other than foreign-born individuals, NHANES does not include (or does not include enough of) persons at higher risk for TB; thus, nationally representative population estimates among higher-risk groups other than foreign born individuals are not available.

Published estimates of LTBI prevalence among higher-risk groups may have limited generalizability based on the specific population(s) used to collect the estimates, the number of participants included, the tests and definitions for a positive test, and whether studies were conducted within a single or multicenter setting. For example, a retrospective study estimated the LTBI prevalence among the homeless New York City population over the years 1992 to 2005 to be 27.1 percent based on convincing self-reported history of positive TST, but prevalence based on actual testing with TST (threshold for positivity was not specified) was 12.5 percent.²¹ A review published in May 2015 offers LTBI prevalence and active TB disease incidence estimates by high-risk categories based on studies published in English, French, or Spanish between 2009 and 2014. These estimates vary by test used (TST or IGRA) and in some cases are based on a single study. These estimates are summarized in Appendix A, Table A1.⁹

Rationale for Screening

The prevention of active TB by treating LTBI is a major goal of the national strategy for eliminating TB in the United States.^{22,23} CDC does not recommend universal or untargeted population screening for LTBI. That is, CDC discourages the use of tests for LTBI among individuals and populations at low risk for TB infection, and discourages a testing approach that is independent of a risk assessment.¹ Rather, screening for LTBI is recommended for those in population subgroups with higher prevalence of LTBI (i.e., prevalence substantially greater than that of the general U.S. population) or who have increased risk of progression from LTBI to active TB disease.

Screening Strategies

For the purposes of this review, we define screening as the use of a test in asymptomatic individuals for the purpose of identifying candidates for medication to prevent progression to active TB. No direct test for the presence of latent MTB is available as a reference standard for LTBI screening tests. The diagnosis of LTBI is based on medical and social history, physical exam, and the results of TST or interferon gamma release assay (IGRA) results (both discussed below). The presence of active TB disease should be excluded before treatment for LTBI is initiated; failure to do so may result in inadequate treatment and development of drug resistance.¹ Chest x-ray is often recommended in patients who have a positive screening test for LTBI, along with a symptom questionnaire to help differentiate LTBI and active TB disease.

TST

TST is administered by injecting 0.1 ml of an intermediate-strength dose of purified protein derivative (PPD) intradermally using the Mantoux technique with interpretation within 48 to 72 hours. In the United States, an intermediate-strength dose is 5 tuberculin units (TU) of PPD-S. In other countries, PPD RT-23 is used and an approximately equivalent intermediate-strength dose is 2 to 2.5 TU.^{24,25} If a person is infected with TB, a delayed-type hypersensitivity reaction is typically detectable 2 to 8 weeks after initial infection. Health care providers should be trained in the administration and interpretation of TST.⁴

Based on the sensitivity and specificity of the TST and the prevalence of TB in different groups, three cut-points have been recommended for defining a positive reaction: ≥ 5 mm, ≥ 10 mm, and ≥ 15 mm of induration.^{1,4} A TST reaction of ≥ 5 mm of induration is considered positive in people with the highest risk of developing active TB: patients with HIV infection; patients with organ transplants and other immunosuppressed patients (e.g., patients taking the equivalent of ≥ 15 mg/day of prednisone for 1 month or those taking tumor necrosis factor-alpha [TNF- α] antagonists); patients with recent contact with a person having infectious TB disease; or people with fibrotic changes on a chest x-ray consistent with prior TB. A TST reaction of ≥ 10 mm of induration is considered positive for LTBI in the following individuals: recent arrivals to the United States (within the last 5 years) from high-TB prevalence countries; injection drug users; residents or employees of high-risk congregate settings (e.g., correctional facilities, long-term care facilities, hospitals and other health care settings, residential facilities for people with HIV infection, and homeless shelters); and people with clinical conditions that increase the risk of progression to TB disease.¹ A TST of ≥ 15 mm of induration is considered positive in individuals with no known risk factors for TB.^{1,4}

IGRAs

IGRA test are performed using fresh whole blood specimens. Blood is mixed with assay peptides that simulate antigens derived from MTB. If infected with MTB, white blood cells recognize the simulated antigens and release interferon-gamma. Currently, two FDA-approved IGRAs are commercially available: QuantiFERON[®] TB-Gold-In-Tube (QFT-GIT) approved by FDA in 2007, and T-SPOT.TB[®], approved by FDA in 2010.^{1,26} The antigens used by both commercially available tests (ESAT-6 and CFP-10) are absent in BCG and most nontuberculosis mycobacteria. QFT-GIT was approved as a modification to the second-generation test approved in 2005 (QuantiFERON TB-Gold[®] (QFT-G)), and includes an additional antigen (TB7.7).

The interpretation of IGRA tests is based on the measured amount of interferon-gamma released (QFT-G and QFT-GIT) or on the number of visible spots that form in response to cells that release interferon-gamma (T-SPOT.TB). CDC recommends that laboratories provide both the qualitative and quantitative results. Qualitative results are reported as positive, negative, indeterminate, or borderline. Quantitative results are reported as numerical values that include a response to the TB antigen and two controls (nil and mitogen). Quantitative results may be useful for clinical decisionmaking in individual cases, in combination with risk factors. IGRAs have some advantages over TST. They require a single patient visit to conduct the test, results can be available within 24 hours, they do not cause a “booster phenomenon,” and they are unaffected by

BCG vaccination and most environmental mycobacteria.¹ However, blood samples must be processed relatively quickly (within 8 to 30 hours after collection), and limited data exist in certain groups (e.g., people recently exposed to TB, immunocompromised people, and those who will be tested repeatedly).

Among individuals at higher risk for TB, CDC considers IGRAs the preferred method of testing for groups of people who have poor rates of return for TST reading and interpretation (e.g., homeless people) and people who have received BCG vaccination.¹ CDC indicates that either TST or IGRAs can be used for other high-risk individuals. CDC does not recommend routine testing with both TST and IGRAs, but suggests situations where results from both tests could be useful. For example, IGRA testing might follow a positive TST among individuals who have a low risk of both infection and progression from infection to TB disease, or follow a negative TST when the risk for infection, progression to disease, and/or a poor outcome is high, such as among HIV-infected people.¹

Because no direct test for LTBI infection exists, the test characteristics for both TST and IGRAs are difficult to establish for latent infection. Sensitivity is generally extrapolated from the sensitivity of these tests in populations with active TB. Similarly, specificity is generally extrapolated from evaluating the test within populations of healthy individuals, free of TB risks or exposures, and without underlying medical conditions that increase risk for TB infection.

Treatment Approaches

Individuals who screen positive for LTBI and in whom active infection has been excluded are generally offered treatment with antituberculosis medications.⁴ For decades, isoniazid (INH) was the only medication used for treating LTBI. However, concerns about adverse events, primarily hepatotoxicity, and difficulty with patient adherence to long treatment regimens prompted the evaluation of alternative regimens.

The American Thoracic Society, CDC, and the Infectious Diseases Society of America recommend several regimens for treating LTBI; these regimens vary by drug, dose frequency, and duration of treatment.²⁷ The regimens include INH and drugs in the rifamycin class (rifampin [RIF] and rifapentine [RPT]). In 2011, CDC issued additional recommendations based on findings from three randomized controlled trials that demonstrated that a weekly regimen with INH and RPT for 12 weeks was as effective as traditional 6- or 9-month INH regimens for healthy individuals.²⁸ The recommended regimens for treating LTBI among adults with LTBI are summarized in Appendix A, Table A2.

Current Clinical Practice in the United States

Current guidelines from several organizations reflect a movement toward targeted testing and treatment. In developed countries with a low prevalence of TB such as the United States, most authorities recommend that LTBI screening be done only among high-risk groups and when treatment is feasible. In 2005, CDC, in collaboration with the American Thoracic Society and the

Infectious Diseases Society of America, issued its most recent joint recommendations for controlling TB.²⁷ In 2011, CDC convened an expert panel to review evidence from new trials that resulted in a recommendation for a new alternative regimen (weekly INH/RPT for 12 weeks).^{27,28} In 2015, the World Health Organization released new guidelines on the management of LTBI that offer a public health approach for testing, treating, and managing LTBI primarily geared toward high and upper middle-income countries.²⁹ These guidelines recommend testing and treatment of LTBI for individuals at the highest risk of progression to active disease, including individuals with HIV, close contacts of active pulmonary TB, and patients with selected conditions or undergoing treatment commonly associated with immunosuppression (e.g., transplant, use of anti-tumor necrosis factor treatments).

Estimates for the current prevalence of screening for LTBI in primary care settings are not available. Further, estimates of the proportion of high-risk groups and individuals cared for in primary care settings are difficult to determine and likely vary by the type of primary care setting (e.g., a public health clinic or safety net provider in a high-TB region of the United States versus a private primary care practice in a low-TB burden community).

Previous USPSTF Recommendation

In 1996, the USPSTF recommended screening with TST for asymptomatic high-risk people (A recommendation) and BCG vaccination only for selected high-risk individuals (B recommendation). Prior to the present update, the USPSTF Web site referred to CDC for this recommendation, stating, “The USPSTF recognizes the importance of targeted screening for tuberculosis. However, the USPSTF does not wish to duplicate the work of the Centers for Disease Control and Prevention (CDC) in this area and will not update its 1996 recommendations.”

Chapter 2. Methods

Key Questions and Analytic Framework

The EPC investigators, U.S. Preventive Services Task Force (USPSTF) members, and Agency for Healthcare Research and Quality (AHRQ) Medical Officers developed the scope and key questions (KQs) for this review. The analytic framework illustrates the KQs that guided the review (Figure 1).

1. Is there direct evidence that targeted screening for latent tuberculosis infection (LTBI) in primary care settings in asymptomatic adults at increased risk for developing active tuberculosis disease (TB) (e.g., individuals in populations with a high prevalence of active TB disease or with documented increased risk for progression from LTBI to active TB disease) improves quality of life, or reduces active TB disease incidence, or reduces transmission of TB, or reduces disease-specific or overall mortality?
- 2a. What is the accuracy and reliability of the tuberculin skin test (TST) or the interferon gamma release assay (IGRA) for screening asymptomatic adults who are at increased risk for developing active TB disease?
- 2b. What is the accuracy and reliability of sequential screening strategies that include both TST and IGRA testing in asymptomatic adults who are at increased risk for developing active TB disease?
3. Does treatment of LTBI with Centers for Disease Control and Prevention (CDC)-recommended pharmacotherapy regimens improves quality of life or reduces progression to active TB disease, or reduces transmission of TB, or reduces disease-specific or overall mortality?
4. Are there harms associated with screening for LTBI?
 - a. Do these harms differ by screening method or strategy?
 - b. Do these harms differ by population?
5. Are there harms associated with treatment for LTBI with CDC-recommended pharmacotherapy regimens?

Data Sources and Searches

With the assistance of a librarian with extensive experience conducting searches in support of systematic reviews, we searched PubMed/MEDLINE and the Cochrane Library for English-language articles published through August 3, 2015. We used Medical Subject Headings (MeSH) as search terms when available and keywords when appropriate, focusing on terms to describe relevant populations, tests, interventions, outcomes, and study designs. Complete search terms and limits are listed in Appendix B1. We conducted targeted searches for unpublished literature by searching ClinicalTrials.gov and the World Health Organization International Clinical Trials Registry Platform (WHO ICTRP). To supplement electronic searches, we reviewed the reference lists of pertinent review articles and studies that met our inclusion criteria, and added all previously unidentified relevant articles. We reviewed all studies suggested by peer reviewers or public comment respondents and, if appropriate, incorporated them into the final review.

Study Selection

We developed inclusion and exclusion criteria for populations, interventions, comparators, outcomes, timing, settings, and study designs (Appendix B2). We excluded studies in which more than 25 percent of the study population were younger than 18 years of age or were known to be HIV positive, unless results were stratified by these characteristics. For KQ 1, we included randomized controlled trials (RCTs) or prospective cohort studies that compared screening with no screening in primary care settings and focused on asymptomatic adults belonging to populations at increased risk for developing active TB (e.g., injection drug users, people who are homeless or residing in homeless shelters, former prisoners, people born in or former residents of countries with high TB prevalence, and people who work with such individuals). We excluded studies on close contacts of people with active TB because testing and treatment of such populations is considered part of contact tracing for public health as opposed to a primary care function. We also excluded studies of people with underlying immunosuppression and for whom LTBI screening and treatment would be part of standard disease management by specialty care providers (e.g., individuals with HIV, head and neck cancer, leukemia or lymphoma, silicosis, history of or planned organ transplant, planned or active use of TNF- α inhibitors, and planned or active use of chemotherapy) because testing and treatment typically need to be individualized and managed with respect to the patient's comorbidities and medication regimens.

For KQ 2, because there is no reference standard test that would allow us to determine sensitivity or specificity of the screening tests for LTBI,²⁶ we relied on data from studies of people with bacteriologically confirmed active TB (for sensitivity) or studies of healthy subjects known to be at low risk for TB and free of TB exposure (for specificity). We included studies assessing the accuracy or reliability of three IGRAs (T-SPOT.TB, QFT-G, and QFT-GIT) using the commercially specified threshold, but also reported results based on other thresholds when available. For studies assessing the accuracy of the TST using the Mantoux method, we required the use of intermediate-strength PPD. Systematic reviews or primary studies of test accuracy were eligible for KQ 2.

For KQs 3 and 5, we included RCTs of people with LTBI, comparing a CDC-recommended treatment (medication, dose, and duration) with placebo, delayed treatment, no treatment, or another CDC-recommended treatment. For KQ 5, prospective cohort studies and case-control studies were also eligible. For KQ 4, systematic reviews, RCTs, and prospective cohort studies reporting false-positive results leading to unnecessary testing (e.g., chest x-ray) or treatment, labeling, stigma, anxiety, or cellulitis were eligible.

For KQs 1, 3, 4, and 5, we included studies conducted in primary settings in countries categorized as “very high” on the Human Development Index (United Nations Human Development Programme). We defined primary care broadly to include public health settings or specialized clinics providing primary care functions (e.g., prison clinics). For KQ 2 sensitivity outcomes, we did not set any exclusion criteria based on setting or country; for KQ 2 specificity outcomes, we excluded studies conducted in high TB-burden countries as defined by the World Health Organization (Appendix B2).³⁰

Two investigators independently reviewed titles and abstracts; those marked for potential

inclusion by either reviewer were retrieved for evaluation of the full text. Then, two investigators independently reviewed the full texts to determine final inclusion or exclusion. Disagreements were resolved by discussion and consensus.

Quality Assessment and Data Abstraction

For each included study, one investigator extracted pertinent information about the methods, populations, interventions, comparators, outcomes, timing, settings, and study designs. A second team member reviewed all data extractions for completeness and accuracy.

We assessed the quality of studies as good, fair, or poor, using predefined criteria developed by the USPSTF and adapted for this topic (Appendix B3).³¹ Two independent reviewers assigned quality ratings for each study. Disagreements were resolved by discussion with an experienced team member. For our main analyses, we included only studies rated as having good or fair quality.

Data Synthesis and Analysis

We qualitatively synthesized findings for each key question by summarizing the characteristics and results of included studies in tabular or narrative format. To determine whether meta-analyses were appropriate, we assessed both the number of studies available, and the clinical and methodological heterogeneity of the studies following established guidance.³² To do this, we qualitatively assessed the populations, similarities and differences in screening tests or treatments used, and similarities in outcomes and timing of outcomes assessed.

For KQ 2, when at least three similar studies were available, we conducted quantitative synthesis of studies with random-effects models using the inverse-variance weighted method (DerSimonian and Laird) to determine pooled estimates of sensitivity and specificity.³³ We generated pooled estimates by test for sensitivity and stratified by important covariates, such as the timing of testing with respect to when pharmacotherapy for TB was started,²⁶ the prevalence of HIV among the study population, the TB burden in the country where the study was conducted, and for T-SPOT.TB, the threshold used to consider the test positive (FDA or European threshold).^{34,35} For specificity, we also generated pooled estimates by test and stratified by important covariates such as the prevalence of BCG vaccination in the study population and TB burden of the country in which the study was conducted. For T-SPOT.TB, we also generated estimates stratified by the threshold used to consider the test positive. We qualitatively summarized reliability outcomes and sensitivity and specificity outcomes for some tests or test thresholds using tables and narrative because we did not have a sufficient number of studies to conduct quantitative syntheses.

We conducted several types of sensitivity analyses. First, because DerSimonian and Laird random-effects models may not perform well for small meta-analyses (when few studies are included), we conducted sensitivity analyses using maximum likelihood random-effects methods.³⁶⁻⁴⁰ Results were essentially the same as for our main analyses, with some minor

variation in width of confidence intervals for some estimates. Therefore, the results from these analyses are only provided in the appendix and are not discussed in the text. Next, we did not include studies rated as poor quality in any main analyses, but did include them in sensitivity analyses.

For KQ 3 and for most comparisons and outcomes related to KQ 5, we did not conduct meta-analyses for our main analysis because we did not have a sufficient number of studies meeting all eligibility criteria that made the same comparison and reported similar outcomes. Therefore, we synthesized the included studies qualitatively, using narrative and tables. We calculated the RR for outcomes of interest (e.g., development of active TB disease, mortality from TB, development of hepatotoxicity) using the number of all randomized patients as the denominator to reflect a true intention-to-treat analysis. For our main analyses for KQ 5, we conducted quantitative synthesis of RCTs comparing INH to RIF for the following outcomes: hepatotoxicity and discontinuation due to adverse events. For sensitivity analyses for KQs 3 and 5, we conducted quantitative synthesis of RCTs by adding excluded studies that compared isoniazid with placebo that met many of our inclusion criteria, but that used a longer duration of treatment than is currently recommended (e.g., they used 1 year of isoniazid),⁴¹⁻⁴⁴ used lower or higher doses than currently recommended,^{42,43} or did not require LTBI confirmation for subjects to be eligible.^{41,43,44} For RCTs to be included in sensitivity analyses, we required that they either confirmed LTBI for subjects to be eligible (e.g., by enrolling only those who were tuberculin positive), reported data for those with confirmed LTBI (e.g., for the tuberculin positive subset of subjects), or that the vast majority of subjects (over 75%) were tuberculin positive. For these analyses, we used random-effects models with the inverse-variance weighted method (DerSimonian and Laird) to estimate RRs.⁴⁵ We conducted quantitative synthesis for the following outcomes: development of active TB disease; hepatotoxicity (e.g., isoniazid-induced hepatitis), and discontinuation of treatment due to adverse events. Because DerSimonian and Laird random-effects models may not perform well for small meta-analyses (when few studies are included), we also conducted sensitivity analyses using profile likelihood random-effects methods.³⁶⁻⁴⁰ Results were essentially the same as for those using DerSimonian and Laird random-effects models, with some minor variation in width of confidence intervals for some estimates.

For all quantitative syntheses, the chi-squared statistic and the I^2 statistic (the proportion of variation in study estimates due to heterogeneity) were calculated to assess statistical heterogeneity in effects between studies.^{46,47} An I^2 from 0 to 40 percent might not be important, 30 percent to 60 percent may represent moderate heterogeneity, 50 percent to 90 percent may represent substantial heterogeneity, and 75 percent or greater represents considerable heterogeneity.⁴⁸ The importance of the observed value of I^2 depends on the magnitude and direction of effects and on the strength of evidence for heterogeneity (e.g., p-value from the chi-squared test or a confidence interval for I^2). However, as precision and the number of subjects increase, I^2 may become inflated toward 100 percent, and may not reflect clinically relevant heterogeneity.⁴⁹

All quantitative analyses were conducted using Stata® version 13.1.⁵⁰

Expert Review and Public Comment

The draft analytic framework and draft research questions were made available for public comment and subsequently revised. This report was reviewed by content and methodologic experts, USPSTF members, and AHRQ Medical Officers, and was revised based on comments.

USPSTF Involvement

This review was funded by AHRQ. Staff of AHRQ and members of the USPSTF participated in developing the scope of the work and reviewed draft manuscripts, but the authors are solely responsible for the content.

Chapter 3. Results

Literature Search

We identified 4,406 unique records and assessed 612 full texts for eligibility (Figure 2). We excluded 541 studies for various reasons detailed in Appendix C and we included 71 published studies of good or fair quality in our main analyses (Appendix D). Of the included studies, 66 were primary studies of screening test characteristics (KQ 2a). Five studies were RCTs focused on the benefits (KQ 3) or harms (KQ 5) of pharmacotherapy for latent tuberculosis infection (LTBI). We identified no eligible studies for KQ 1 (direct evidence of screening for LTBI) or KQ 4 (harms of screening). Details of quality assessments are provided in Appendix E.

Results by Key Question

Key Question 1. Direct Evidence for Targeted Screening for Latent Tuberculosis Infection

We found no eligible studies that addressed this question.

Key Question 2a. Accuracy and Reliability of Screening Tests

We identified 66 studies of good or fair quality assessing the sensitivity, specificity, or reliability for one or more of the included screening tests.

Sensitivity of Screening Tests

We relied on evidence from 50 primary studies of good or fair quality in subjects with bacteriologically confirmed, active TB because no reference standard for direct diagnosis of LTBI exists.⁵¹⁻¹⁰⁰

Study Characteristics

Thirteen studies estimated sensitivity for TST,^{51-55,57,62,69,70,73-75,98} 16 studies for T-SPOT.TB,^{56,60,61,64,67,69,71,75,78-80,82,83,86-88} 16 studies for QFT-G,^{55,56,59,65-72,77,84,85,87,88} and 24 studies for QFT-GIT.^{51,57,58,63-65,73,74,76,81,83,86,88-96,98-100} Characteristics of studies are provided in Appendix D, Tables D1 (TST) and D2 (IGRA). One study started using QFT-G but converted to using QFT-GIT midway through the study.⁹⁷ Eight studies estimating sensitivity were conducted in high TB-burden countries,^{51,57,58,63,76,78,95,96} 29 were conducted in intermediate TB-burden countries,^{55,64-73,75,79-85,88-94,97,98,100} and 10 were conducted in low TB-burden countries,^{52-54,56,59-62,74,77} including 4 in the United States. Three multinational studies were conducted in a mix of low and intermediate TB-burden countries.^{86,87,99} Sixteen studies were conducted in countries with a human development index of less than “very high.”^{51,57,58,63,69,70,75,76,78,82,83,87,90,91,95,96} Twenty-nine studies provided stratified results for the HIV-negative segment of their study population or

excluded patients with HIV from the study population.^{51,53-57,60,62-64,66-70,73,74,76,77,80,81,85,88,89,91,92,95,96,98} Three studies were conducted with a study population with fewer than 25 percent of the study participants with BCG vaccination.^{58,76,77} Thirteen studies included between 25 and 75 percent vaccinated study populations,^{54,55,57,62,63,65,67,68,72,74,91,93,94} and 12 studies included study populations with more than 75 percent BCG vaccination.^{51,56,69-71,75,84,85,87,95,96,98} Twenty-two studies did not report the BCG vaccination prevalence in the study population. The timing of testing with respect to starting antituberculosis treatment varied across studies by the following categories: prior to or within 7 days (20 studies),^{53,56,57,62,63,65,67,69-71,74,76,77,81,84,87,92,95,96,100} within 14 days (4 studies),^{59,60,64,86} within 30 days (1 study),⁷⁵ or not reported (25 studies).^{51,52,54,55,58,61,66,68,72,73,78-80,82,83,85,88-91,93,94,97-99}

Results

We calculated pooled estimates for sensitivity of TST by test threshold and of IGRA by test (Table 1, Figures 3 and 4). The pooled sensitivity for TST was 0.79 (95% CI, 0.69 to 0.89; $I^2=94.6\%$) for the 5-mm threshold, 0.79 (95% CI, 0.71 to 0.87; $I^2=91.4\%$) for the 10-mm threshold, and 0.52 (95% CI, 0.35 to 0.68, $I^2=95.5\%$) for the 15-mm threshold. For T-SPOT.TB, we found no difference in estimates based on whether the FDA or European threshold for a positive test was used, so we combined all studies for a pooled estimate of 0.90 (95% CI, 0.87 to 0.93; $I^2=63.6\%$) (Appendix F, Figure F19). We found lower estimates for sensitivity of the QFT tests; the pooled estimate for sensitivity of QFT-G was 0.77 (95% CI, 0.74 to 0.81; $I^2=55.3\%$) and was 0.80 (95% CI, 0.77 to 0.84; $I^2=74.3\%$) for QFT-GIT. The percent of IGRA tests with indeterminate results ranged from 3 to 7 percent in studies reporting this information.

In a sensitivity analysis, we added the 14 studies^{81,85,101-112} that were excluded for poor quality, and found that estimates did not appreciably change (Appendix D, Tables D5 and D6; Appendix F, Figures F1 and F2). We repeated all analyses using a maximum likelihood-based method for random effects, and found similar results except for a slightly higher point estimate for TST at 5-mm (Appendix F, Figures F4 and F5).

Because we found moderate to substantial statistical heterogeneity, we stratified results for all tests based on factors that were consistently reported across studies and could impact the accuracy of the test (Appendix F, Figures F7 through F31). Factors that might lower sensitivity include testing that occurs after antituberculosis treatment has been started and higher proportion of study subjects with HIV or other immunosuppressing conditions. Other factors that might affect accuracy include country TB burden and BCG vaccination prevalence among the study population. Our stratified analyses by these two factors were similar because the prevalence of BCG vaccination among the study population is often correlated with the country's TB burden; BCG vaccination is common in intermediate and high TB burden countries as compared to low TB burden countries.

The stratified analyses identified heterogeneity between strata for several of these factors but are limited by few studies in some of the strata we evaluated. Factors influencing sensitivity estimates were not consistent among all tests (or test thresholds for TST), limiting our ability to draw definitive conclusions. For TB country burden, estimates for sensitivity of TST at 10- and 15-mm thresholds were higher in low TB-burden countries as compared to intermediate- and

high-burden countries (Appendix F, Figures F13 and F17). We could not identify sources of heterogeneity for sensitivity estimates that were consistently present across the studies of the three IGRA tests.

Specificity of Screening Tests

Similarly, because no reference standard for direct diagnosis of LTBI exists, we relied on 18 primary studies in low or intermediate TB-burden countries among healthy individuals known to be free of TB risks and exposures to estimate specificity.^{53,54,69,75,86,93,113-124}

Study Characteristics

Fourteen studies estimated specificity for TST,^{53,54,69,75,113-118,120-123} 5 studies for T-SPOT.TB,^{69,75,86,121,123} 3 studies for QFT-G,^{69,118,119} and 4 studies for QFT-GIT.^{86,93,123,124} Characteristics of studies are described in Appendix D, Tables D3 and D4. All studies were conducted in countries with a human development index of “very high.” Three studies were conducted in intermediate TB-burden countries^{69,75,93}; 1 study was conducted in two countries, one of which is low TB burden and the other intermediate⁸⁶; and the remaining 14 studies were conducted in low TB burden countries (10 of these were in the United States). Four studies were conducted with study populations with more than 75 percent BCG vaccination,^{69,75,93,122} 9 studies included fewer than 5 percent BCG vaccinated participants,^{53,54,113,114,117,118,120,121,123} and BCG vaccination was not reported in 5 studies.^{86,116,119,124,125}

Results

We calculated a pooled estimate for specificity of TST at the 10-mm threshold of 0.97 (95% CI, 0.96 to 0.99, $I^2=94.3\%$) and at the 15-mm threshold of 0.99 (95% CI, 0.98 to 0.99; $I^2=91.7\%$). The pooled estimate for specificity of T-SPOT.TB was 0.95 (95% CI, 0.92 to 0.98; $I^2=79.1\%$). The pooled estimate for specificity of QFT-G was 0.98 (95% CI, 0.90 to 1.0) and the pooled estimate for QFT-GIT was 0.97 (95% CI, 0.94 to 0.99, $I^2=93.4\%$). A limited number of available studies precluded quantitative synthesis for TST at the 5-mm threshold. Estimates are summarized in Table 2 and Figure 5. The percentage of IGRA tests with indeterminate results ranged from 0 to 3 percent in studies reporting this information.

As part of a sensitivity analysis, we added five studies^{94,103,105,108,126} that had been excluded for poor quality and found similar results (Appendix D, Tables D7 and D8; Appendix F, Figure F3). We repeated all analyses using a maximum likelihood-based method for random effects and found similar point estimates, but with slightly larger confidence intervals (Appendix F, Figure F6). Because of substantial heterogeneity among studies, we stratified results based on country TB burden and BCG vaccination prevalence (Appendix F, Figures F32 through F44). Across all tests, specificity was substantially lower in intermediate TB-burden countries than in low TB-burden countries, yet removing these studies had marginal effect on the overall pooled estimates and inconsistency as measured by I^2 . For example, at the 10-mm threshold of the TST, we removed the one study conducted in an intermediate TB burden country with a specificity of 0.45; the pooled estimate changed from 0.97 to 0.98 and the I^2 statistic was reduced from 94.3 percent to 91.2 percent (Appendix F, Table F34).

Reliability of Screening Tests

We identified nine studies of good or fair quality assessing the reliability for at least one of the included screening tests.^{75,113,114,123,127-131}

Study Characteristics

Study characteristics are shown in Appendix D, Table D9. Three studies assessed the interrater reliability of TST.^{113,114,123} Two studies assessed the interrater reliability of T-SPOT.*TB*,^{75,128} one assessed the interrater reliability of QFT-GIT,¹³⁰ and one assessed the interlaboratory reliability of QFT-GIT.¹²⁹ Two studies assessed the test-retest reliability of T-SPOT.*TB* and QFT-GIT 1 to 4 weeks after an initial test.^{127,131,132} Eight studies were conducted in a low TB-burden country (seven in the United States and one in The Netherlands), one study was conducted in an intermediate burden TB country⁷⁵ (Turkey), and one study enrolled Nepalese military recruits who had left Nepal and recently entered the United Kingdom.¹³¹ Two studies reported the percentage of the study population that had HIV; less than 1 percent in both studies were HIV positive.^{127,131} In two studies the majority of participants were BCG vaccinated.^{75,131}

Results

Interrater reliability. Three studies (N=1,826,¹²³ N=1,189,¹¹⁴ and N=127¹¹³) measured the interrater reliability for TST results by reporting the kappa statistic for agreement by TST reaction size; results ranged from 0.55 to 0.79, indicating moderate to substantial agreement between two observers. One study (N=91) found substantial agreement between two observers for manually reading T-SPOT.*TB* results (kappa=0.92) and manual versus automatic ELISPOT readings (kappa=0.73).⁷⁵ One study (N=313) evaluated agreement among six individual ELISPOT readers; all kappa values were above 0.6.¹²⁸ One study (N=146) assessed interrater reliability for manual versus automated ELISA readings for QFT-GIT; each study participant had two blood draws and each sample was sent for both automated and manual readings.¹³⁰ Across all samples, 88.6 percent of results were concordant and 11.0 percent were discordant; the discordance rates for specific comparisons were 4.8 percent (between two different automated readings, kappa= 0.85), 6.9 percent (between two different manual readings, kappa=0.80), and 3.4 percent (manual compared with automated, kappa ranged from 0.73 to 0.90 across comparisons).¹³⁰

Interlaboratory reliability. One study (N= 91) evaluated the interlaboratory reliability of QFT-GIT by sending three blood specimens from each participant to three different laboratories noted to have extensive experience and proficiency with IGRA testing and interpretation.¹²⁹ Across all three labs, 7.7 percent of participants had discordant results (none had indeterminate results); kappas of pairwise lab sample comparisons ranged from 0.87 to 0.93.¹²⁹

Reproducibility and test-retest reliability. One study (N= 130) assessed the reliability of IGRA results by processing two blood samples from each study participant (using the same laboratory and same type of test interpretation); 5.8 percent of participants had discordant results for QFT-GIT and 6.5 percent had discordant results for T-SPOT.*TB*.¹²⁷ Two studies measured the test-retest reliability for QFT-GIT. One study enrolled U.S. health care workers (HCWs)¹²⁷

and one enrolled a population from a high TB-burden country (Nepal).^{131,132} In the study (N=130) enrolling HCWs, 8 percent of baseline T-SPOT.*TB* negative tests changed to positive and 53 percent of positive tests changed to negative on repeat testing at 2 weeks; for QFT-GIT, 8 percent of negative tests changed to positive and 33 percent of positive tests changed to negative.¹²⁷ Finally, in the study enrolling a Nepalese population, the kappa statistic for agreement between initial QFT-GIT test and retest at 1 week was 0.48 (95% CI, 0.26 to 0.70) and was 0.66 (95% CI, 0.5 to 0.83) for T-SPOT.*TB*.¹³¹

Key Question 2b. Accuracy and Reliability of Sequential Screening Strategies

We found no eligible studies that addressed this question.

Key Question 3. Benefits of Treatment for LTBI

We included three RCTs (Thompson 1982, Menzies 2008, Sterling 2011) assessing treatment of LTBI that met all eligibility criteria (Appendix D, Table D10).¹³³⁻¹³⁵ One compared isoniazid with placebo;¹³⁵ one compared rifampin with isoniazid;¹³³ and one compared rifapentine plus isoniazid with isoniazid alone.¹³⁴

We identified four additional RCTs (Bush 1965, Falk 1978, Ferebee 1963, Veening 1968) that compared isoniazid with placebo that did not meet all eligibility criteria, but that we used in sensitivity analyses (Appendix D, Table D13). For RCTs to be included in sensitivity analyses, we required that they either confirmed LTBI for subjects to be eligible (e.g., by enrolling only those who were tuberculin positive), reported data for those with confirmed LTBI (e.g., for the tuberculin-positive subset of subjects), or that the vast majority of subjects (over 75%) were tuberculin positive. These trials met many of our eligibility criteria, but they all used a longer duration of treatment than is currently recommended by the CDC⁴¹⁻⁴⁴ (i.e., they used 1 year or more of isoniazid), and some used lower or higher doses than currently recommended^{42,43} or did not require LTBI confirmation for subjects to be eligible.^{41,43,44} One of the four trials was rated poor quality for high risk of selection bias, attrition bias, confounding, and measurement bias.⁴²

Our searches identified additional RCTs that compared isoniazid with placebo, which we excluded from this review. Reasons for excluding studies from this review are listed in Appendix C. For example, several trials focused on the use of isoniazid for household contacts of active cases, but did not require LTBI for study entry;¹³⁶⁻¹³⁸ over half of the participants in these trials were children; they evaluated 1 year or more of isoniazid; and 1 trial used a higher dose¹³⁸ than is currently recommended by CDC. Two other trials randomized households or villages to evaluate the prophylactic use of isoniazid in areas with a high prevalence of active TB at the time of the study (Greenland¹³⁹ or Alaska¹⁴⁰). These two did not require LTBI for study entry. One evaluated an unusual isoniazid regimen (400 mg x 3 months, nothing for 3 months, then 400 mg x 3 months¹³⁹); the other evaluated 1 year of isoniazid and included many children.¹⁴⁰ Other excluded RCTs evaluated patients with silicosis¹⁴¹ or renal transplant and dialysis patients.¹⁴²

Isoniazid Compared With Placebo

The International Union Against Tuberculosis (IUAT) trial was the single trial meeting all eligibility criteria that compared isoniazid with placebo.¹³⁵ It randomized 27,830 adults from seven European countries with fibrotic pulmonary lesions but without active TB or previous anti-TB treatment to four groups: isoniazid 300 mg daily for 12 weeks, isoniazid 300 mg daily for 24 weeks (currently a CDC-approved regimen), isoniazid 300 mg daily for 52 weeks, or placebo. Participants were required to have a 6 mm or greater Mantoux test. The median age was 50 years. Just over half were men.

After 5 years of followup, 76 (1.1%), 34 (0.5%), 24 (0.3%), and 97 (1.4%) participants developed active TB in the four groups, respectively (Appendix D, Table D11). The RRs for developing active TB compared with placebo were 0.79 (95% CI, 0.58 to 1.06), 0.35 (95% CI, 0.24 to 0.52), and 0.25 (95% CI, 0.16 to 0.39), respectively (Figure 6). For the 24-week CDC approved regimen, we calculated a number needed to treat (NNT) of 112 to prevent one case of active TB. Our sensitivity analyses using data from the 24- and 52-week groups from the IUAT trial and four additional RCTs, including a total of 36,823 participants, found an RR of 0.31 (95% CI, 0.24 to 0.41) and no statistical heterogeneity in effects between studies (I^2 , 0.0%) (Figure 7; Appendix D, Table D14).

The IUAT trial found that people with larger fibrotic pulmonary lesions had a greater risk of developing active TB. The incidence of active TB in the placebo group was half as great among people with lesions $< 2\text{cm}^2$ (11.6 per 1000) as among people with larger lesions (21.3 per 1000).

There were no deaths due to TB in any of the isoniazid groups in the IUAT trial; three people died from TB in the placebo group. The RR for death due to TB was 0.14 (95% CI, 0.01 to 2.78) for each of the isoniazid groups compared with placebo. All-cause mortality was not reported separately for the four groups. The trial reported benefit-to-risk ratios (defined as cumulative TB cases prevented/cumulative hepatitis cases incurred) of 1.2, 2.6, and 2.1 for the isoniazid groups compared with placebo, respectively.

Rifampin Compared With Isoniazid

The one included RCT making this comparison was an open-label trial conducted in Canada, Brazil, and Saudi Arabia that randomized 847 participants to 4 months of rifampin or 9 months of isoniazid to compare adverse events and treatment completion.¹³³ Because this RCT was focused largely on adverse events, it is described in greater detail with the results for KQ 5. We mention it briefly in this section because it reported zero deaths from TB in either group. It also reported all-cause mortality, with zero deaths in the rifampin group and one in the isoniazid group.

Rifapentine Plus Isoniazid Compared With Isoniazid Alone

The one included RCT making this comparison, the PREVENT TB study, was an open-label, noninferiority trial conducted in the United States, Canada, Brazil, and Spain, that randomized 7,731 people age 12 or older to directly observed once-weekly rifapentine (900 mg) plus

isoniazid (900 mg) for 3 months or to daily self-administered isoniazid (300 mg) for 9 months.¹³⁴ The primary endpoint was development of confirmed TB. Subjects were primarily from the United States and Canada (89% of those randomized) and were high-risk individuals with positive tuberculin skin tests. Most (71%) had a close contact with a patient with culture-confirmed, active TB within the past 2 years; 25 percent were included solely because of recent conversion to skin-test positivity. Less than 3 percent of participants were HIV positive; the participants with HIV were not required to have a positive TST. Risk factors for TB included a history of incarceration (5.1%), history of injection-drug use (3.7%), and homelessness (27.8%).

Almost 90 percent of subjects randomized completed 33 months of followup. Active TB developed in 7 people in the combination-therapy group and in 15 people in the isoniazid-only group. The combination-therapy group was found to be noninferior to the isoniazid-only group. The trial identified 70 deaths from any cause (31 vs. 39, $p=0.22$).

From among the 7,731 randomized, we obtained data from CDC for the subset of participants most directly relevant for this review: the 6,886 adults (18 years of age or older) who were HIV negative and were TST or IGRA positive. The median age for this subset was 37. Just over half were men; 57 percent were white. For this subset, active TB developed in 5 people in the combination-therapy group and in 10 people in the isoniazid-only group. The combination-therapy group was found to be noninferior to the isoniazid-only group. Overall mortality was similar for the two groups (30 vs. 34, respectively, $p=0.42$).

Key Question 4. Harms of Screening for LTBI

We did not identify any studies addressing this question.

Key Question 5. Harms of Treatment for LTBI

We included five RCTs assessing harms associated with the treatment of LTBI that met all eligibility criteria (Appendix D, Table D10).^{133-135,143,144} One compared isoniazid with placebo;¹³⁵ three compared rifampin with isoniazid;^{133,143,144} and one compared rifapentine plus isoniazid with isoniazid alone.¹³⁴

We identified five additional RCTs that evaluated harms associated with treatment of LTBI that did not meet all eligibility criteria, but that we used in sensitivity analyses (Appendix D, Table D13). Criteria for RCTs to be included in sensitivity analyses for KQ 5 are the same as those described for KQ 3. The five additional trials met many of our eligibility criteria, but four of the five used a longer duration of treatment than is currently recommended by CDC^{41,43,44,145} (i.e., they used 1 year or more of isoniazid), one used a shorter duration than is currently recommended by CDC (3 months of isoniazid),¹⁴⁶ and some used a lower dose than currently recommended⁴³ or did not require LTBI confirmation for subjects to be eligible.^{41,43,44,145} All five of these trials compared isoniazid with placebo, and were rated as having fair quality. Our searches identified additional RCTs and one observational study that compared isoniazid with placebo, which we excluded from this review. Reasons for excluding studies from this review are listed in Appendix C.

From this body of evidence, we were able to quantitatively synthesize harms related to hepatotoxicity and discontinuation of medication due to adverse events. Studies also reported a variety of gastrointestinal (GI) adverse events, but we were unable to quantitatively synthesize these outcomes because of heterogeneity in how GI adverse events were measured across included studies. For example, GI adverse events were reported as a single combined value per treatment arm in one study,⁴³ as rates of treatment discontinuation due to GI events in another study,¹⁴⁷ and reported by separate types of GI events (i.e., nausea, clay-colored stools, or anorexia) with no summary rate in a third study.¹⁴⁶ No studies reported harms related to peripheral neuropathy or development of drug-resistant TB.

Isoniazid Compared With Placebo

The IUAT trial was the single trial meeting all eligibility criteria that compared isoniazid with placebo.^{135,147} Study characteristics for this trial were previously described (See KQ 3 results); the quality of this study was rated as fair for KQ 5 outcomes because harm outcomes were not prespecified, and ascertainment techniques were not adequately described except for the hepatotoxicity outcomes.

Hepatotoxicity

The IUAT trial reported rates of hepatotoxicity development (Appendix D, Table D12).¹³⁵ The RRs for developing hepatotoxicity associated with isoniazid compared with placebo were 3.45 (95% CI, 1.49 to 7.99) for 12 weeks of treatment, 4.59 (95% CI, 2.03 to 10.39) for 24 weeks of treatment, and 6.21 (95% CI, 2.79 to 13.79) for 52 weeks of treatment (Figure 8). For the study arms comparing the 24-week CDC-approved regimen to placebo (N=13,955), we calculated that one case of hepatotoxicity would result from treating 279 people with isoniazid (i.e., a number needed to harm [NNH] of 279). Our sensitivity analyses using data from the IUAT trial (three treatment arms combined) and three additional RCTs,^{41,145,146} including a total of 35,161 participants, found an RR of 5.04 (95% CI, 2.50 to 10.15) and no statistical heterogeneity among studies (I^2 , 0.0%, $p=0.630$) (Appendix G, Figure G1).

The one RCT included in the main analysis comparing isoniazid with placebo for treatment of LTBI¹³⁵ reported mortality rates from hepatotoxicity of 0.03 percent, 0.0 percent, and 0.01 percent for the 12-, 24-, and 52-week isoniazid treatment groups. This study had zero deaths from hepatotoxicity among placebo-treated patients. The authors reported that the mortality rate from hepatitis associated with isoniazid was 0.14 per 1,000 people receiving isoniazid, for a calculated RR of 2.35 (95% CI, 0.12 to 45.46; NNH, 6,947).

Treatment Discontinuation Because of Adverse Events

Rates of treatment discontinuation because of adverse events in the IUAT study were presented only for all three isoniazid treatment groups combined. A total of 345 patients (1.8%) receiving isoniazid discontinued treatment because of adverse events compared with 84 patients (1.2%) receiving placebo. The RR of discontinuation owing to adverse events among patients treated with isoniazid vs. placebo was 1.50 (95% CI, 1.18 to 1.89; 1 RCT, N=27,830, NNH, 167). Our sensitivity analysis using data from the IUAT study and three additional RCTs,^{43,44,146} including

a total of 55,398 participants, found an RR of 1.58 (95% CI, 1.00 to 2.49; I^2 , 70.2) (Appendix G, Figure G2).

Gastrointestinal (GI) Adverse Events

The IUAT trial reported that 1.2 percent of isoniazid patients and 0.9 percent of placebo patients discontinued treatment due to GI distress.¹⁴⁷ The RR of discontinuation due to GI distress was among isoniazid versus placebo patients was 1.33 (95% CI, 1.01 to 1.75). Among studies included in sensitivity analyses, one⁴³ reported gastrointestinal adverse events (0.7% in isoniazid group vs. 0.3% in placebo), one study¹⁴⁶ reported nausea (3.3% in isoniazid group vs. 1.7% in placebo), clay-colored stools (10.0% in isoniazid group vs. 5.0% in placebo), and anorexia (8.3% in both isoniazid and placebo groups).

Other Harms

No other adverse events were reported in the IUAT trial.¹³⁵ A variety of other adverse events were reported in the RCTs included in sensitivity analyses. Rates of other adverse events were generally similar among isoniazid and placebo patients (Appendix D, Table D15). One study reported an increased risk for rash (0.9% of isoniazid patients and 0.3% of placebo patients; RR, 2.7; 95% CI, 1.27 to 5.73).^{41,148}

Rifampin Compared With Isoniazid

Three open-label RCTs compared rifampin with isoniazid (Appendix D, Table D10). One trial was conducted in Canada (N=116) compared 4 months of rifampin (10 mg/kg of body weight, up to 600 mg/day) with 9 months of isoniazid (5 mg/kg, up to 300 mg/day).¹⁴³ Participants were 18 years of age or older with documented LTBI; more than half were male. A later study by the same authors conducted in Canada, Brazil, and Saudi Arabia randomized 847 participants to the same two treatments.¹³³ Participants were 18 years of age or older with documented LTBI; just over half were male. The third trial randomized inmates (N=365) in the San Francisco City and County Jail diagnosed with LTBI at jail entry to 9 months of isoniazid (900 mg twice per week) or 4 months of rifampin (600 mg/day).¹⁴⁴ Ninety-three percent of study participants were male.

Hepatotoxicity

Rates of hepatotoxicity in these three RCTs among individuals receiving isoniazid were 5.2 percent,¹⁴³ 3.7 percent,¹³³ and 11.4 percent¹⁴⁴ (Appendix D, Table D12). Rates among rifampin-treated patients were 0.0 percent, 0.7 percent, and 4.4 percent, respectively. The RRs of hepatotoxicity from these three RCTs for isoniazid compared with rifampin were 7.00 (95% CI, 0.37 to 132.56), 5.25 (95% CI, 1.54 to 17.87), and 2.57 (95% CI, 1.17 to 5.65), respectively. Our meta-analysis of these three RCTs (total N=1,327) found a greater risk of hepatotoxicity for patients treated with isoniazid than for those treated with rifampin, (RR, 3.29; 95% CI, 1.72 to 6.28; I^2 , 0.0%) (Figure 9). All studies reported zero deaths from hepatotoxicity.

Treatment Discontinuation Because of Adverse Events

Rates of discontinuations because of adverse events were reported in all three included RCTs. Rates were 13.8 percent (isoniazid) and 3.4 percent (rifampin);¹⁴³ 5.6 percent (isoniazid) and 3.8 percent (rifampin);¹³³ and 0.0 percent (isoniazid) and 1.1 percent (rifampin)¹⁴⁴ (Appendix D, Table D12). The RR of discontinuation due to adverse events for isoniazid compared with rifampin for these three studies was 4.0 (95% CI, 0.89 to 18.04), 1.48 (95% CI, 0.80 to 2.74), and 0.20 (95% CI, 0.01 to 4.05). Our meta-analysis found no statistically significant difference between treatments (RR, 1.61; 95% CI, 0.57 to 4.57; I^2 , 40.0%, N=1,327) (Figure 10).

Gastrointestinal Adverse Events

Among the three included RCTs, one reported GI adverse events in 3.4 percent of the study population, not separated by treatment arm.¹⁴³ One reported grade one/two GI intolerance among 0.2 percent of rifampin patients and 0.5 percent of isoniazid patients.¹³³ The third study reported GI adverse events among 9 percent of rifampin patients and 10 percent of isoniazid patients.¹⁴⁴ The pooled RR among the two studies reporting adverse events by treatment arm was 1.60; 95% CI, 0.76 to 3.40; I^2 , 0%; N=1,211).

Other Harms

The three RCTs in the main analysis reported on various other harms (Appendix D, Table D12). None of these harms involved statistically significant differences in RR for isoniazid compared with rifampin. The pooled RR for adverse events categorized as “other” by two RCTs was RR, 0.82; 95% CI, 0.42 to 1.59; 3 RCTs; I^2 , 0%; N=480).^{143,144}

Rifapentine Plus Isoniazid Compared With Isoniazid

The one included RCT making this comparison, the PREVENT TB study, was an open-label, noninferiority trial conducted in the United States, Canada, Brazil, and Spain, that randomized 7,731 people 12 years of age or older to directly observed once-weekly rifapentine (RPT) at 900 mg plus isoniazid (INH) at 900 mg (RPT+INH) for 3 months, or to daily self-administered isoniazid at 300 mg (INH only) for 9 months.¹³⁴ More details regarding this study are presented in the results section on benefits of treatment (KQ 3).

Hepatotoxicity

Rates of grade 3 and 4 hepatotoxicity were 4.9 and 1.0 percent in the RPT+INH arm and 5.5 and 1.1 percent in the INH-only arm, respectively. The RR for grade 3 or 4 hepatotoxicity was 0.90 (95% CI, 0.75 to 1.08). Mortality from hepatotoxicity was reported in 1.0 percent of isoniazid patients and 0.8 percent of isoniazid plus rifapentine patients (RR, 0.83; 95% CI, 0.51 to 1.35).

Treatment Discontinuation Because of Adverse Events

Rates of discontinuation because of adverse events were 5.2 percent in the RPT+INH arm and 4.1 percent in the INH-only arm. The RR of treatment discontinuation due to adverse events for

RPT+INH versus INH-only was 1.28 (95% CI, 1.03 to 1.59).

Other Harms

Possible hypersensitivity was reported in 0.5 percent of isoniazid patients and 4.1 percent of isoniazid plus rifapentine patients. The RR of possible hypersensitivity for RPT+INH versus INH-only was 8.04 (95% CI, 4.88 to 13.26).

Chapter 4. Discussion

Summary of Evidence

Tables 3 through 5 provide a summary of findings in this evidence review. These tables, which are presented by key question (KQ), provide a summary of outcomes organized by test or intervention along with a description of precision, risk of bias, and applicability.

Evidence for Benefit and Harms of Screening

We did not identify any randomized controlled trials (RCTs) or prospective cohort studies directly assessing the effectiveness or harms of screening for latent tuberculosis infection (LTBI) compared with no screening in the populations and outcomes specified for this review.

Accuracy and Reliability of Screening Tests

The evidence on accuracy and reliability was based on fair quality evidence overall. Because of the lack of tests for the direct diagnosis of LTBI, evaluating accuracy of tests relies on extrapolation from test characteristics among populations with active, confirmed TB (sensitivity) or healthy individuals known to be free of TB risks and exposures (specificity). The evidence suggests that for the populations studied, currently available tests are moderately sensitive, and in low TB-burden countries, highly specific. Sensitivity estimates for tuberculin skin testing (TST) (at 5-mm and 10-mm, but not 15-mm thresholds) and QuantiFERON[®] (QFT) interferon-gamma release assays (IGRAs) were consistent, with pooled estimates ranging from 0.77 to 0.80. Sensitivity estimates for the T-SPOT.*TB* IGRA test were higher at 0.90, and estimates of sensitivity for IGRA tests were more precise than those for TST. Pooled estimates for specificity for TST at 10- and 15-mm thresholds and all IGRA tests ranged from 0.95 to 0.99. We judged specificity estimates to be consistent and precise in low TB-burden countries. Our findings for sensitivity and specificity are generally consistent with other systematic reviews evaluating accuracy, despite differences in study inclusion and exclusion criteria.¹⁴⁹⁻¹⁵² We found limited evidence for the reliability of these tests and, of those identified, few assessed reliability in the same way.

The applicability of the evidence on accuracy and reliability of screening tests to primary care practice settings and populations is uncertain, since the lack of a direct test for LTBI requires test accuracy studies to be performed in specific populations (e.g., active, confirmed TB, healthy low TB-risk populations) to ensure the validity of findings. We found lower estimates for specificity in studies conducted with populations from intermediate TB-burden countries. This could be the result of unintentional inclusion of subjects with unknown past TB exposure, resulting in increased frequency of positive results, inclusion of bacilli Calmette–Guérin (BCG) vaccinated subjects increasing false positive TST results, or because of other factors that affect the administration or interpretation of tests among populations in these countries.

Despite this uncertainty, the evidence is likely applicable to primary care practice settings that

serve high-risk populations (e.g., public health settings, residents of high-risk congregate settings, clinics serving foreign-born populations), where the use of a highly specific test among a higher prevalence population minimizes false positives and results from a moderately sensitive test can be combined with a clinical risk assessment to determine the likelihood of infection to inform treatment decisions. In these settings, clinical risk assessment prior to testing may already be a part of standard clinical workflow, and clinic and laboratory staff may have extensive experience with appropriate testing techniques and interpretation. However, many primary care practice settings may not serve large populations with high risk for LTBI; thus, an approach that relies on an individualized clinical assessment for LTBI risk to inform decisions regarding testing may not be part of standard workflow. Systematic identification of high-risk individuals cared for in low-prevalence practice settings may be challenging and associated with opportunity costs.

Benefits and Harms of Treatment for LTBI

The best evidence on effectiveness of treatment for LTBI with a CDC-recommended regimen was from the International Union Against Tuberculosis (IUAT) trial, a large (N=27,830) good-quality study. It found a 65 percent relative reduction in progression to active TB at 5 years for 24 weeks of isoniazid compared with placebo (NNT 112). Our sensitivity analysis adding four RCTs (that did not meet all of our eligibility criteria) that used a longer duration of treatment than is currently recommended and some different doses than currently recommended found a similar reduction. The IUAT trial enrolled subjects with pulmonary fibrotic lesions, a group thought to be at the highest risk for progression to active TB. In this trial, those with smaller lesions progressed to active TB at lower rates than those with larger lesions. Further, the populations included in the other treatment studies used in our sensitivity analysis were not people identified to have LTBI via screening in primary care settings; rather, they were household contacts of active cases,⁴³ veterans with inactive pulmonary TB,^{41,148} people residing in mental institutions,⁴⁴ and military members exposed to an active TB case.⁴² Thus, the available evidence may not be applicable to individuals in primary care settings who screen positive on TST or IGRAs, but who have normal chest radiographs or who are not recent converters or close contacts. Thus, estimates of treatment effectiveness may represent the upper bounds of effectiveness, which may be lower in other screen-positive populations. Further, all of the RCTs that assessed the effectiveness of isoniazid compared with placebo were published more than 30 years ago (1963, 1965, 1968, 1978, 1982). Most of them evaluated 1 year of treatment with isoniazid, because that was the recommended treatment for many years; shorter durations and other regimens were later studied with a focus on reducing harms (and little attention on evidence for benefits). It is unclear whether changes in the prevalence of TB, treatments for active TB, or likelihood of LTBI progressing to active TB would significantly change estimates of effectiveness.

We found limited evidence on efficacy for other CDC-recommended regimens meeting our eligibility criteria and scant evidence on effectiveness of treatments for reducing mortality due to TB or all-cause mortality. No studies compared rifampin or rifapentine plus isoniazid with placebo, or compared a 9-month course of isoniazid with placebo. However, the included head-to-head open label, noninferiority RCT (the PREVENT TB trial) that compared a combination of once-weekly rifapentine plus isoniazid for 3 months with daily isoniazid for 9 months found the

combination therapy to be noninferior (with estimates trending in favor of combination therapy) to isoniazid alone for preventing the development of active TB.

The evidence on harms was fair quality overall and suggests a more than fourfold increased risk for hepatotoxicity for treatment with 6 months of isoniazid compared with placebo and a more than three times increased risk when compared with rifampin. Deaths because of hepatotoxicity were rare across all studies included such that estimates were imprecise. In the IUAT study, all three subjects who died from hepatitis had continued to take INH after liver abnormalities were recognized.¹³⁵ Two studies used in sensitivity analysis for harms reported normalization of liver enzyme levels among those experiencing asymptomatic elevation¹⁴⁵ or mild hepatitis.^{41,148} Discontinuation of treatment because of adverse events was modestly increased for isoniazid as compared with placebo, but estimates of no difference between isoniazid and rifampin were inconsistent and imprecise. Gastrointestinal distress, an outcome that represents a heterogeneous group of harms in both type and severity, was inconsistently reported by included studies. Other harms reported were limited by inconsistent and imprecise findings. Other adverse events occurred infrequently and may be subject to more bias in determination than are hepatotoxicity or discontinuation because of adverse events.

The overall benefits and harms of screening and treatment are influenced by several factors. The NNT is driven both by the effectiveness of treatment as compared with no treatment, and by the rate of progression to active TB among an untreated group. Given that treatment of LTBI has been the standard of care for decades, contemporary data for estimating efficacy/effectiveness are not available. A recent study to estimate the cost-effectiveness of screening for latent tuberculosis using TST and IGRA among different risk groups specified in current CDC screening guidelines reported similar difficulties in establishing robust estimates of TB reactivation, and uncertainty in test characteristics as a result of the lack of a referent standard for diagnosis for LTBI.¹⁵³ Proponents for screening suggest benefits on outcomes related to TB transmission and through case-finding of active TB that occurs during screening. However, we identified no studies meeting our study selection criteria that reported on outcomes related to TB transmission.

Hypothetical Outcomes of a Screening Program

The hypothetical outcomes of a screening program for LTBI are illustrated in Table 6. These outcomes are a crude approach to estimating the overall benefits and harms of screening a population, and several scenarios are illustrated to provide alternative outcomes based on differing prevalence of infection, and differing rates of progression from latent to active TB. A detailed list of assumptions and relevant citations for assumptions are provided in the table notes.

We calculate outcomes for two different prevalence estimates for LTBI (20.5% for the foreign-born U.S. population and 4.7% for the overall U.S. population) and provide the range of outcomes based on the lower and upper confidence intervals associated with these prevalence estimates (shown in brackets in Table 6). For the sensitivity and specificity of tests, we use the pooled estimates from our meta-analysis for TST at the 10-mm threshold, although in practice, the threshold used is typically individualized to the risk of the person being screened. We assume all individuals who test positive receive a chest x-ray to rule out active disease, and we assume

that a proportion of individuals will not be offered treatment based on a history of prior treatment for LTBI or active TB disease. We assume rates of progression in the absence of treatment based on rates of progression in the placebo arm of the IUAT trial (1.4% at 5 years) and an alternative rate of progression based on more recent estimates (0.084/100 person-years for overall population and 0.098/100 person-years for foreign-born U.S. population).⁸ We use an estimate for the RR reduction in progression to active TB for treatment with 6 months of isoniazid based on the IUAT trial (0.35). We assume rates of hepatotoxicity and discontinuation owing to adverse events based on estimates calculated for this review.

For the base case using the foreign-born U.S. population prevalence estimate and rate of progression from the IUAT trial, we estimate that for 100,000 asymptomatic patients screened and eligible for treatment, 18,580 will have a positive test, require a chest x-ray and be offered treatment. Of these, 2,385 are patients with false positive tests that do not have any potential to benefit from treatment. Of those treated, 79 patients will progress to active TB despite treatment, compared with 225 cases if no screening and subsequent treatment were offered. This is equivalent to a number needed to treat to prevent one case of progression to active TB of 111. Under an alternative assumption regarding rate of progression to active TB, we estimate that fewer cases will progress to active TB with (28) or without (79) treatment, and the number needed to treat to prevent one case from progressing to active TB increases to 314. We had insufficient evidence to estimate benefits relating to prevention of TB deaths, TB transmission, and improvements in quality of life.

With respect to harms, 85 patients treated with isoniazid for 6 months will experience hepatotoxicity as compared with 19 if no screening and subsequent treatment were offered, for a number needed to harm to cause one case of hepatotoxicity of 279. Likewise, 334 subjects would discontinue INH treatment owing to adverse events as compared with 223 if no screening and treatment were offered for a number needed to harm to cause one discontinuation owing to adverse events of 167. Fewer cases of hepatotoxicity would occur with treatment with rifampin (26) compared with no treatment (19) for a number needed to harm to cause one case of hepatotoxicity of 2,531. We had insufficient data to estimate outcomes for other types of harms such as psychological harms, peripheral neuropathy, hematologic reactions, and dermatologic or hypersensitivity reactions.

Overall, the estimated number of active TB cases prevented ranges from 52 to 146, depending on which assumption for progression to active TB is used. Sixty-seven cases of hepatotoxicity would be caused if using INH; 9 of those cases would be caused by unnecessary treatment in people with false positive screening results and no potential to benefit from treatment. One hundred and eleven cases of treatment discontinuation owing to adverse events would occur; 14 of those would occur in individuals with no potential to benefit (false positives).

Table 6 also shows hypothetical outcomes using estimates of LTBI prevalence for the overall U.S. community-dwelling population, a lower prevalence population than the U.S. foreign-born population. For this population, fewer absolute numbers of individuals screen positive and are subjected to treatment and thus experience treatment-related harms. However, over 40 percent (2,859) of subjects offered treatment (6,572) are subjects with false positive tests that have no potential for benefit yet are subjected to the risk of harms from treatment.

Limitations of the Review

This review is limited in the ability to directly assess the effectiveness of targeted screening for LTBI because we identified no studies comparing screened against unscreened populations among the populations considered in this review. LTBI screening and treatment among some high-risk individuals is a standard of practice. Thus, trials comparing screening with no screening in these populations have not been conducted.

We could not assess screening test characteristics specifically for LTBI because of the absence of a reference standard for direct diagnosis. We relied on extrapolation from studies in active TB populations for sensitivity, and healthy subjects for specificity, an approach consistent with other studies estimating sensitivity and specificity of these tests. We identified a substantial amount of statistical heterogeneity in some of our pooled estimates, although we believe this heterogeneity is not clinically meaningful and we suspect inflation of I^2 (the proportion of variation in study estimates due to heterogeneity) among specificity outcomes because of very precise individual study estimates.⁴⁹ When possible, we stratified analyses by study features possibly contributing to the heterogeneity, but few studies were available for some of the strata used in analysis, and with rare exceptions, findings were not consistent for explaining heterogeneity across tests (or test thresholds for TST). We did not stratify findings by reagent used for TST (PPD vs. RT-23); the equivalence between these two reagents has not been established in recent years and may have contributed to heterogeneity in findings.

The studies of screening tests in our review did not consistently report comorbidities of the study population tested. Although we excluded studies and results from populations with more than 25 percent HIV-infected individuals, patients with active TB often have underlying comorbidities related to immunosuppression, and the extent to which sensitivity of tests is blunted by this underlying immunosuppression is not known and may result in lower estimates for sensitivity than would otherwise be found in populations with latent infection. On the other hand, the presence of active disease may result in more host sensitization than would occur compared with latent infection such that this population may overestimate the true sensitivity of the tests for latent infection. We did not identify any eligible studies evaluating the sequential use of tests; studies that used more than one test typically performed both tests on the study population to assess concordance rates, or used a second test only in the case of an indeterminate or unexpected result on the first test.

Evidence on reliability of tests was limited. For the T-SPOT.*TB* test, manual versus automated reading of specimens could affect reliability, but few studies using T-SPOT.*TB* formally evaluated this. Further, test-retest reliability may vary by baseline prevalence of LTBI; the U.S. study assessing this had a higher rate of reversion from positive to negative as compared to the study conducted among Nepalese immigrants. For TST, test-retest reliability is challenging to measure because repeated testing in someone with a positive test is not clinically recommended because of the risk of stimulating an even larger hypersensitivity reaction. Moreover, interpretation of repeat testing among subjects initially testing negative is complicated by the well-known booster phenomenon. Studies of reliability included in this review were not conducted in primary care settings. Both tests (TST and IGRA) have fairly detailed test procedures for administration, handling, and interpretation. The one study assessing IGRA

interlaboratory reliability sent specimens to labs that have extensive expertise and experience with IGRA testing and interpretation. Thus, the applicability of reliability evidence to primary care practice settings or labs that may not have the expertise or economies of scale to perform tests with high fidelity to recommended instructions for testing is uncertain.

We identified no studies assessing the harms of screening compared with no screening. Potential harms include overdiagnosis and treatment for LTBI that would have never progressed to active TB. Potential harms also include incidental findings on chest x-ray for people who screen positive for LTBI, which result in the need for follow-up CT scans or serial x-rays for findings such as lung nodules unrelated to TB disease. This review was also limited in the ability to determine the burden of repeat testing required for people who have indeterminate results on IGRAs. Last, we did not identify any evidence about psychosocial harms for people who screen positive and may experience anxiety or stigma associated with being labeled as infected with TB.

This review was limited to the evaluation of existing CDC-recommended LTBI treatment regimens. Isoniazid was established as an effective treatment of LTBI several decades ago; the IUAT trial and the RCTs in our sensitivity analysis were published more than 30 years ago. CDC-treatment recommendations have evolved based on interval studies comparing shorter durations and alternative regimens against the standard isoniazid regimen to reduce harms, improve adherence, or both, rather than to assess efficacy. Since the original isoniazid trials were conducted, the prevalence of TB has declined, yet the prevalence of resistant strains among those infected has increased; thus, the applicability of evidence from an era before multidrug TB resistance is unclear. We identified little information on the rate of progression from LTBI to active TB in the modern era, which is an important determinant for making decisions about treatment.

Our review excluded treatments that are not recommended by CDC and also excluded several populations (e.g., children, people with HIV). A recent network meta-analysis of treatment for LTBI that used a mixed-treatment comparison methodology suggests that some of the more recently recommended regimens are efficacious for preventing active TB (e.g., rifampin for 3–4 months, rifapentine-isoniazid combination), potentially more so than isoniazid alone, and may have fewer adverse effects.¹⁵⁴ This analysis included studies among children; HIV-infected individuals; household or close contacts of active TB without confirmed LTBI; and individuals with renal transplant, silicosis, or rheumatoid arthritis taking immunosuppressive biologic medication, which were all populations excluded from the present review. It also included treatment regimens not eligible for our review. A systematic review conducted for the Cochrane Collaboration on isoniazid for preventing TB in non-HIV-infected people found a significant reduction in active TB over 2 years or longer using data from 11 RCTs (RR, 0.40; 95% CI, 0.31 to 0.52). The review included studies among children, household or close contacts of active TB patients in the absence of confirmed LTBI, individuals with renal transplant, and those with silicosis, which were all populations excluded from the present review.¹⁵⁵

Future Research Needs

Continuing declines in TB incidence in the United States during the past several decades suggest

progress toward reaching the public health goal of TB elimination. Most active TB cases are reactivations of latent TB as opposed to new transmission. Risk for LTBI and progression to active TB is on a continuum, and although there is certainty about individuals and populations at the absolute highest risk, there is uncertainty in the LTBI prevalence and rates of progression for individuals and populations at increased risk but perhaps not at the highest absolute risk—for example, people with diabetes and smokers. More research to elucidate the epidemiology of LTBI in these groups could inform future screening and treatment strategies to better tailor individual screening and treatment recommendations. Screening benefits are maximized when treatment is directed to those most likely to progress to active TB and harms minimized when unnecessary evaluation and treatment are avoided in those unlikely to have been exposed to TB or who are unlikely to progress if latently infected. Future research to develop more accurate screening tests, more effective LTBI treatments with fewer harms and side effects, and treatments requiring shorter duration with higher rates of patient adherence would also improve the overall benefit of an LTBI screening program.

In addition to research to improve the accuracy of screening tests and the effectiveness or safety of treatment, research is needed to determine efficient ways of identifying candidates for LTBI testing that take advantage of varied data sources and alternative venues for risk assessment beyond primary care office settings. Primary care settings serving the general population are different from primary care provided in specialized clinics that care for high-risk populations (e.g., prison clinics, clinics serving large proportions of foreign-born populations) and TB-specific public health settings; thus, an approach to clinical risk assessment and testing that can be tailored based on setting and practice characteristics is needed. For example, operations research may be needed to identify efficient ways of identifying high-risk individuals seen in low-prevalence community practice settings. Further, research that informs our understanding of the incremental net benefit of more or less frequent screening could also help determine optimal approaches to screening.

Conclusion

We identified no studies that directly evaluated the benefits and harms of a screening program for LTBI as compared with no screening among the populations considered in this review. Both types of currently available screening tests for LTBI (TST and IGRAs) are moderately sensitive, and within low TB-burden countries are highly specific. Isoniazid treatment reduces the risk of progression to active TB for people with LTBI and pulmonary fibrotic lesions. The evidence for benefit on other outcomes (e.g., TB mortality, all-cause mortality) or other regimens is limited or not available for the populations considered in this review. Isoniazid is associated with higher rates of hepatotoxicity than placebo and rifampin regimens. Isoniazid is also associated with higher risk for discontinuation of treatment owing to adverse events than placebo, but this risk was similar to the risk for rifampin regimens.

References

1. Centers for Disease Control and Prevention (CDC). Latent Tuberculosis Infection: A Guide for Primary Health Care Providers. Developed in partnership with the New Jersey Medical School Global Tuberculosis Institute Atlanta, GA: CDC, National Centers for HIV/AIDS, Viral Hepatitis, STD, and TB Prevention; 2013.
<http://www.cdc.gov/tb/publications/LTBI/default.htm>
2. Grzybowski S, Barnett GD, Styblo K. Contacts of cases of active pulmonary tuberculosis. *Bull Int Union Tuberc*. 1975;50(1):90-106. Epub: 1975/01/01. PMID: 1218291.
3. Marks SM, Taylor Z, Qualls NL, et al. Outcomes of contact investigations of infectious tuberculosis patients. *Am J Respir Crit Care Med*. 2000 Dec;162(6):2033-8. Epub: 2000/12/09. PMID: 11112109.
4. American Thoracic Society, Centers for Disease Control and Prevention. Targeted tuberculin testing and treatment of latent tuberculosis infection. *MMWR Recomm Rep*. 2000 Jun 9;49(RR-6):1-51. Epub: 2000/07/06. PMID: 10881762.
5. Ferebee SH. Controlled chemoprophylaxis trials in tuberculosis. A general review. *Bibl Tuberc*. 1970;26:28-106. Epub: 1970/01/01. PMID: 4903501.
6. Horsburgh CR, Jr. Priorities for the treatment of latent tuberculosis infection in the United States. *N Engl J Med*. 2004 May 13;350(20):2060-7. Epub: 2004/05/14. PMID: 15141044.
7. Sloot R, Schim van der Loeff MF, Kouw PM, et al. Risk of tuberculosis after recent exposure. A 10-year follow-up study of contacts in Amsterdam. *Am J Respir Crit Care Med*. 2014 Nov 1;190(9):1044-52. Epub: 2014/09/30. PMID: 25265362.
8. Shea KM, Kammerer JS, Winston CA, et al. Estimated rate of reactivation of latent tuberculosis infection in the United States, overall and by population subgroup. *Am J Epidemiol*. 2014 Jan 15;179(2):216-25. PMID: 24142915.
9. Getahun H, Matteelli A, Chaisson RE, et al. Latent Mycobacterium tuberculosis infection. *N Engl J Med*. 2015 May 28;372(22):2127-35. PMID: 26017823.
10. Lindsay RP, Shin SS, Garfein RS, et al. The association between active and passive smoking and latent tuberculosis infection in adults and children in the United States: results from NHANES. *PLoS One*. 2014 Mar 24;9(3):e93137. PMID: 24664240.
11. Horsburgh CR, Jr., Rubin EJ. Clinical practice. Latent tuberculosis infection in the United States. *N Engl J Med*. 2011 Apr 14;364(15):1441-8. PMID: 21488766.
12. Centers for Disease Control and Prevention (CDC). Data and Statistics. Atlanta, GA: CDC; 2014 March 7. <http://www.cdc.gov/tb/statistics/default.htm>. Accessed May 22, 2015.
13. Scott C, Kirking HL, Jeffries C, et al. Tuberculosis trends--United States, 2014. *MMWR Morb Mortal Wkly Rep*. 2015 Mar 20;64(10):265-9. PMID: 25789741.
14. Hoyert DL, Xu J. Deaths: preliminary data for 2011. *Natl Vital Stat Rep*. 2012 Oct 10;61(6):1-51. PMID: 24984457.
15. Centers for Disease Control and Prevention, National Center for HIV/AIDS VH, STD, and TB Prevention, Prevention DoHA. HIV and Tuberculosis. Atlanta, GA: Centers for Disease Control and Prevention; 2013 March.
<http://www.cdc.gov/hiv/resources/factsheets/hivtb.htm>. Accessed 9 June 2015.

16. Centers for Disease Control and Prevention. National Tuberculosis Indicators. 2013 State Comparison. Atlanta, GA: CDC, National Center for HIV/AIDS, Viral Hepatitis, STD, and TB Prevention, Division of Tuberculosis Elimination
http://www.cdc.gov/tb/statistics/pdf/statetbindicators_2013statecomparison_final.pdf. Accessed 8 Jun 2015.
17. World Health Organization. Global tuberculosis report 2014. World Health Organization WHO/HTM/TB/2014.08. Geneva, Switzerland: 2014.
http://www.who.int/tb/publications/global_report/en/
18. Centers for Disease Control and Prevention. Reported Tuberculosis in the United States, 2013. Atlanta, GA: U.S. Department of Health and Human Services, Centers for Disease Control and Prevention 2014.
19. Centers for Disease Control and Prevention. Tuberculosis (TB) (Mycobacterium tuberculosis). Atlanta, GA: Centers for Disease Control and Prevention; 2015 May 6.
<http://www.cdc.gov/nndss/conditions/tuberculosis/> Accessed 8 June 2015.
20. Miramontes R, Hill AN, Yelk Woodruff RS, et al. Tuberculosis Infection in the United States: Prevalence Estimates from the National Health and Nutrition Examination Survey, 2011-2012. PLoS One. 2015;10(11):e0140881. PMID: 26536035.
21. McAdam JM, Bucher SJ, Brickner PW, et al. Latent tuberculosis and active tuberculosis disease rates among the homeless, New York, New York, USA, 1992-2006. Emerg Infect Dis. 2009 Jul;15(7):1109-11. PMID: 19624932.
22. Advisory Council for the Elimination of Tuberculosis (ACET). Tuberculosis elimination revisited: obstacles, opportunities, and a renewed commitment. MMWR Recomm Rep. 1999 Aug 13;48(RR-9):1-13. Epub: 1999/09/15. PMID: 10485562.
23. Geiter L, ed Ending neglect: the elimination of tuberculosis in the United States. Washington, DC: National Academy Press; 2000.
24. Comstock GW, Edwards LB, Philip RN, et al. A COMPARISON IN THE UNITED STATES OF AMERICA OF TWO TUBERCULINS, PPD-S AND RT 23. Bull World Health Organ. 1964;31:161-70. Epub: 1964/01/01. PMID: 14253239.
25. Lee E, Holzman RS. Evolution and current use of the tuberculin test. Clin Infect Dis. 2002 Feb 1;34(3):365-70. PMID: 11774084.
26. Mazurek GH, Jereb J, Vernon A, et al. Updated guidelines for using Interferon Gamma Release Assays to detect Mycobacterium tuberculosis infection - United States, 2010. MMWR Recomm Rep. 2010 Jun 25;59(RR-5):1-25. Epub: 2010/06/26. PMID: 20577159.
27. Taylor Z, Nolan CM, Blumberg HM. Controlling tuberculosis in the United States. Recommendations from the American Thoracic Society, CDC, and the Infectious Diseases Society of America. MMWR Recomm Rep. 2005 Nov 4;54(Rr-12):1-81. Epub: 2005/11/04. PMID: 16267499.
28. Centers for Disease Control and Prevention. Recommendations for use of an isoniazid-rifapentine regimen with direct observation to treat latent Mycobacterium tuberculosis infection. MMWR Morb Mortal Wkly Rep. 2011 Dec 9;60(48):1650-3. Epub: 2011/12/14. PMID: 22157884.
29. World Health Organization. 2, Recommendations. In: Guidelines on the Management of Latent Tuberculosis Infection. Geneva: World Health Organization; 2015.

30. World Health Organization. Tuberculosis Country Profiles. Geneva, Switzerland: World Health Organization; 2014 www.who.int/tb/country/data/profiles/en/. Accessed 21 May 2015.
31. Harris RP, Helfand M, Woolf SH, et al. Current methods of the US Preventive Services Task Force: a review of the process. *Am J Prev Med*. 2001 Apr;20(3 Suppl):21-35. PMID: 11306229.
32. West SL, Gartlehner G, Mansfield AJ, et al. Comparative Effectiveness Review Methods: Clinical Heterogeneity. Methods Research Report AHRQ Publication No. 10-EHC070-EF. Rockville, MD: Agency for Healthcare Research and Quality; 2010.
33. Nyaga VN, Arbyn M, Aerts M. Metaprop: a Stata command to perform meta-analysis of binomial data. *Arch Public Health*. 2014;72(1):39. PMID: 25810908.
34. TSPOT. TB Package Insert UK. Oxfordshire, UK: Oxford Immunotec Ltd; 2013 <http://www.oxfordimmunotec.com/international/wp-content/uploads/sites/3/PI-TB-IVD-UK-V2.pdf>. Accessed 4 May 2015.
35. T-SPOT.TB. Package Insert US. Oxfordshire, UK: Oxford Immunotec; 2013 <http://www.oxfordimmunotec.com/international/wp-content/uploads/sites/3/PI-TB-IVD-UK-V2.pdf> Accessed 4 May 2015.
36. Kontopantelis E, Reeves D. metaan: random-effects meta-analysis. *Stata J*. 2010;10(3):395-407.
37. Kontopantelis E, Springate DA, Reeves D. A re-analysis of the Cochrane Library data: the dangers of unobserved heterogeneity in meta-analyses. *PLoS One*. 2013;8(7):e69930. PMID: 23922860.
38. Hardy RJ, Thompson SG. A likelihood approach to meta-analysis with random effects. *Stat Med*. 1996 Mar 30;15(6):619-29. PMID: 8731004.
39. Jackson D, White IR, Thompson SG. Extending DerSimonian and Laird's methodology to perform multivariate random effects meta-analyses. *Stat Med*. 2010 May 30;29(12):1282-97. PMID: 19408255.
40. Cornell JE, Mulrow CD, Localio R, et al. Random-effects meta-analysis of inconsistent effects: a time for change. *Ann Intern Med*. 2014 Feb 18;160(4):267-70. PMID: 24727843.
41. Falk A, Fuchs GF. Prophylaxis with isoniazid in inactive tuberculosis. A Veterans Administration Cooperative Study XII. *Chest*. 1978 Jan;73(1):44-8. Epub: 1978/01/01. PMID: 340155.
42. Veening GJ. Long term isoniazid prophylaxis. Controlled trial on INH prophylaxis after recent tuberculin conversion in young adults. *Bull Int Union Tuberc*. 1968 Dec;41:169-71. Epub: 1968/12/01. PMID: 4885378.
43. Bush OB, Jr., Sugimoto M, Fujii Y, et al. Isoniazid prophylaxis in contacts of persons with known tuberculosis. Second report. *Am Rev Respir Dis*. 1965 Nov;92(5):732-40. Epub: 1965/11/01. PMID: 5321147.
44. Ferebee SH, Mount FW, Murray FJ, et al. A controlled trial of isoniazid prophylaxis in mental institutions. *Am Rev Respir Dis*. 1963 Aug;88:161-75. Epub: 1963/08/01. PMID: 14045220.
45. Sutton AJ, Abrams KR, Jones DR, et al. Methods for meta-analysis in medical research. In: *Wiley Series in Probability and Statistics - Applied Probability and Statistics Section*. London: Wiley; 2000.

46. Higgins JP, Thompson SG. Quantifying heterogeneity in a meta-analysis. *Stat Med*. 2002 Jun 15;21(11):1539-58. PMID: 12111919.
47. Higgins JP, Thompson SG, Deeks JJ, et al. Measuring inconsistency in meta-analyses. *BMJ*. 2003 Sep 6;327(7414):557-60. PMID: 12958120.
48. Higgins JPT, Green S. *Cochrane Handbook for Systematic Reviews of Interventions: The Cochrane Collaboration*. London: The Cochrane Collaboration; 2014 17 April 2015. www.cochrane-handbook.org. Accessed 26 May 2015.
49. Rücker G, Schwarzer G, Carpenter JR, et al. Undue reliance on I(2) in assessing heterogeneity may mislead. *BMC Med Res Methodol*. 2008;8:79. PMID: 19036172.
50. StataCorp. *Stata Statistical Software: Release 13.1*. College Station, TX: StataCorp LP; 2013.
51. Painter JA, Graviss EA, Hai HH, et al. Tuberculosis screening by tuberculosis skin test or QuantiFERON-TB Gold In-Tube Assay among an immigrant population with a high prevalence of tuberculosis and BCG vaccination. *PLoS One*. 2013 12/19;8(12):e82727. PMID: 24367546.
52. Seibert AF, Haynes J, Jr., Middleton R, et al. Tuberculous pleural effusion. Twenty-year experience. *Chest*. 1991 Apr;99(4):883-6. Epub: 1991/04/11. PMID: 1901261.
53. Fietta A, Meloni F, Cascina A, et al. Comparison of a whole-blood interferon-gamma assay and tuberculin skin testing in patients with active tuberculosis and individuals at high or low risk of *Mycobacterium tuberculosis* infection. *Am J Infect Control*. 2003 Oct;31(6):347-53. Epub: 2003/11/11. PMID: 14608301.
54. Berkel GM, Cobelens FG, de Vries G, et al. Tuberculin skin test: estimation of positive and negative predictive values from routine data. *Int J Tuberc Lung Dis*. 2005 Mar;9(3):310-6. Epub: 2005/03/25. PMID: 15786896.
55. Kang YA, Lee HW, Yoon HI, et al. Discrepancy between the tuberculin skin test and the whole-blood interferon gamma assay for the diagnosis of latent tuberculosis infection in an intermediate tuberculosis-burden country. *JAMA*. 2005 Jun 8;293(22):2756-61. Epub: 2005/06/09. PMID: 15941805.
56. Goletti D, Carrara S, Vincenti D, et al. Accuracy of an immune diagnostic assay based on RD1 selected epitopes for active tuberculosis in a clinical setting: a pilot study. *Clin Microbiol Infect*. 2006 Jun;12(6):544-50. Epub: 2006/05/17. PMID: 16700703.
57. Tsiouris SJ, Coetzee D, Toro PL, et al. Sensitivity analysis and potential uses of a novel gamma interferon release assay for diagnosis of tuberculosis. *J Clin Microbiol*. 2006 Aug;44(8):2844-50. PMID: 16891501.
58. Adetifa IM, Lugos MD, Hammond A, et al. Comparison of two interferon gamma release assays in the diagnosis of *Mycobacterium tuberculosis* infection and disease in The Gambia. *BMC Infect Dis*. 2007;7:122. Epub: 2007/10/27. PMID: 17961228.
59. Dewan PK, Grinsdale J, Kawamura LM. Low sensitivity of a whole-blood interferon-gamma release assay for detection of active tuberculosis. *Clin Infect Dis*. 2007 Jan 1;44(1):69-73. Epub: 2006/12/05. PMID: 17143818.
60. Janssens JP, Roux-Lombard P, Perneger T, et al. Quantitative scoring of an interferon-gamma assay for differentiating active from latent tuberculosis. *Eur Respir J*. 2007 Oct;30(4):722-8. Epub: 2007/06/01. PMID: 17537773.
61. Losi M, Bossink A, Codecasa L, et al. Use of a T-cell interferon-gamma release assay for the diagnosis of tuberculous pleurisy. *Eur Respir J*. 2007 Dec;30(6):1173-9. Epub: 2007/08/24. PMID: 17715165.

62. Mazurek GH, Weis SE, Moonan PK, et al. Prospective comparison of the tuberculin skin test and 2 whole-blood interferon-gamma release assays in persons with suspected tuberculosis. *Clin Infect Dis*. 2007 Oct 1;45(7):837-45. Epub: 2007/09/07. PMID: 17806047.
63. Pai M, Joshi R, Bandyopadhyay M, et al. Sensitivity of a whole-blood interferon-gamma assay among patients with pulmonary tuberculosis and variations in T-cell responses during anti-tuberculosis treatment. *Infection*. 2007 Apr;35(2):98-103. Epub: 2007/04/03. PMID: 17401714.
64. Chee CB, Gan SH, Khinmar KW, et al. Comparison of sensitivities of two commercial gamma interferon release assays for pulmonary tuberculosis. *J Clin Microbiol*. 2008 Jun;46(6):1935-40. Epub: 2008/04/11. PMID: 18400912.
65. Harada N, Higuchi K, Yoshiyama T, et al. Comparison of the sensitivity and specificity of two whole blood interferon-gamma assays for *M. tuberculosis* infection. *J Infect*. 2008 May;56(5):348-53. Epub: 2008/04/09. PMID: 18395264.
66. Kobashi Y, Mouri K, Yagi S, et al. Usefulness of the QuantiFERON TB-2G test for the differential diagnosis of pulmonary tuberculosis. *Intern Med*. 2008;47(4):237-43. Epub: 2008/02/16. PMID: 18277023.
67. Kobashi Y, Mouri K, Yagi S, et al. Clinical evaluation for diagnosing active TB disease and transitional change of two commercial blood tests. *Scand J Infect Dis*. 2008;40(8):629-34. Epub: 2008/07/22. PMID: 18642159.
68. Kobashi Y, Mouri K, Yagi S, et al. Clinical utility of the QuantiFERON TB-2G test for elderly patients with active tuberculosis. *Chest*. 2008 May;133(5):1196-202. Epub: 2008/02/12. PMID: 18263689.
69. Soysal A, Torun T, Efe S, et al. Evaluation of cut-off values of interferon-gamma-based assays in the diagnosis of *M. tuberculosis* infection. *Int J Tuberc Lung Dis*. 2008 Jan;12(1):50-6. Epub: 2008/01/05. PMID: 18173877.
70. Ak O, Dabak G, Ozer S, et al. The evaluation of the Quantiferon-TB Gold test in pulmonary and extrapulmonary tuberculosis. *Jpn J Infect Dis*. 2009 Mar;62(2):149-51. Epub: 2009/03/24. PMID: 19305058.
71. Higuchi K, Kawabe Y, Mitarai S, et al. Comparison of performance in two diagnostic methods for tuberculosis infection. *Med Microbiol Immunol*. 2009 Feb;198(1):33-7. Epub: 2008/11/27. PMID: 19034505.
72. Kobashi Y, Sugiu T, Shimizu H, et al. Clinical evaluation of the T-SPOT.*TB* test for patients with indeterminate results on the QuantiFERON TB-2G test. *Intern Med*. 2009;48(3):137-42. Epub: 2009/02/03. PMID: 19182423.
73. Park SY, Jeon K, Um SW, et al. Clinical utility of the QuantiFERON-TB Gold In-Tube test for the diagnosis of active pulmonary tuberculosis. *Scand J Infect Dis*. 2009;41(11-12):818-22. Epub: 2009/11/20. PMID: 19922063.
74. Bocchino M, Chairadonna P, Matarese A, et al. Limited usefulness of QuantiFERON-TB Gold In-Tube for monitoring anti-tuberculosis therapy. *Respir Med*. 2010 Oct;104(10):1551-6. Epub: 2010/06/15. PMID: 20542675.
75. Dilektasli AG, Erdem E, Durukan E, et al. Is the T-cell-based interferon-gamma releasing assay feasible for diagnosis of latent tuberculosis infection in an intermediate tuberculosis-burden country? *Jpn J Infect Dis*. 2010 Nov;63(6):433-6. Epub: 2010/11/26. PMID: 21099095.

76. Legesse M, Ameni G, Mamo G, et al. Performance of QuantiFERON-TB Gold In-Tube (QFTGIT) for the diagnosis of Mycobacterium tuberculosis (Mtb) infection in Afar Pastoralists, Ethiopia. BMC Infect Dis. 2010;10:354. Epub: 2010/12/18. PMID: 21162756.
77. Metcalfe JZ, Cattamanchi A, Vittinghoff E, et al. Evaluation of quantitative IFN-gamma response for risk stratification of active tuberculosis suspects. Am J Respir Crit Care Med. 2010 Jan 1;181(1):87-93. Epub: 2009/10/03. PMID: 19797760.
78. Tan CK, Lai CC, Chen HW, et al. Enzyme-linked immunospot assay for interferon-gamma to support the diagnosis of tuberculosis in diabetic patients. Scand J Infect Dis. 2010 Oct;42(10):752-6. Epub: 2010/06/02. PMID: 20513167.
79. Boyd AE, Ashcroft A, Lipman M, et al. Limited added value of T-SPOT.TB blood test in diagnosing active TB: a prospective bayesian analysis. J Infect. 2011 Jun;62(6):456-61. Epub: 2011/05/17. PMID: 21570124.
80. Cho OH, Park KH, Kim SM, et al. Diagnostic performance of T-SPOT.TB for extrapulmonary tuberculosis according to the site of infection. J Infect. 2011 Nov;63(5):362-9. Epub: 2011/07/26. PMID: 21781986.
81. Kim EY, Park MS, Kim YS, et al. Risk factors for false-negative results of QuantiFERON-TB Gold In-Tube assay in non-HIV-infected patients with culture-confirmed tuberculosis. Diagn Microbiol Infect Dis. 2011 Jul;70(3):324-9. Epub: 2011/05/07. PMID: 21546200.
82. Lai CC, Tan CK, Lin SH, et al. Diagnostic value of an enzyme-linked immunospot assay for interferon-gamma in cutaneous tuberculosis. Diagn Microbiol Infect Dis. 2011 May;70(1):60-4. Epub: 2011/04/26. PMID: 21513844.
83. Lai CC, Tan CK, Lin SH, et al. Diagnostic performance of whole-blood interferon-gamma assay and enzyme-linked immunospot assay for active tuberculosis'. Diagn Microbiol Infect Dis. 2011 Oct;71(2):139-43. Epub: 2011/08/16. PMID: 21840675.
84. Lui G, Lee N, Cheung SW, et al. Interferon gamma release assay for differentiating tuberculosis among pneumonia cases in acute healthcare setting. J Infect. 2011 Jun;62(6):440-7. Epub: 2011/05/18. PMID: 21575991.
85. Ra SW, Lyu J, Choi CM, et al. Distinguishing tuberculosis from Mycobacterium avium complex disease using an interferon-gamma release assay. Int J Tuberc Lung Dis. 2011 May;15(5):635-40. Epub: 2011/07/16. PMID: 21756514.
86. Ruhwald M, Dominguez J, Latorre I, et al. A multicentre evaluation of the accuracy and performance of IP-10 for the diagnosis of infection with M. tuberculosis. Tuberculosis (Edinb). 2011 May;91(3):260-7. Epub: 2011/04/05. PMID: 21459676.
87. Walsh MC, Camerlin AJ, Miles R, et al. The sensitivity of interferon-gamma release assays is not compromised in tuberculosis patients with diabetes. Int J Tuberc Lung Dis. 2011 Feb;15(2):179-84, i-iii. Epub: 2011/01/12. PMID: 21219678.
88. Kobashi Y, Abe M, Mouri K, et al. Usefulness of tuberculin skin test and three interferon-gamma release assays for the differential diagnosis of pulmonary tuberculosis. Intern Med. 2012;51(10):1199-205. Epub: 2012/06/13. PMID: 22687790.
89. Lee J, Lee SY, Won DI, et al. Comparison of whole-blood interferon-gamma assay and flow cytometry for the detection of tuberculosis infection. J Infect. 2013 Apr;66(4):338-45. Epub: 2012/09/27. PMID: 23010554.

90. Taki-Eddin L, Monem F. Utility of an interferon-gamma release assay as a potential diagnostic aid for active pulmonary tuberculosis. *J Infect Dev Ctries*. 2012 Jan;6(1):67-72. Epub: 2012/01/14. PMID: 22240431.
91. Feng JY, Huang SF, Lee MC, et al. Characteristics of IFN-gamma responses in IGRA among pulmonary TB suspects in a TB-endemic area. *Diagn Microbiol Infect Dis*. 2013 Sep;77(1):46-52. Epub: 2013/07/23. PMID: 23867329.
92. Jeon YL, Nam YS, You E, et al. Factors influencing discordant results of the QuantiFERON-TB Gold In-tube test in patients with active TB. *J Infect*. 2013 Oct;67(4):288-93. Epub: 2013/06/26. PMID: 23796867.
93. Kim S, Kim YK, Lee H, et al. Interferon gamma mRNA quantitative real-time polymerase chain reaction for the diagnosis of latent tuberculosis: a novel interferon gamma release assay. *Diagn Microbiol Infect Dis*. 2013 Jan;75(1):68-72. Epub: 2012/10/30. PMID: 23102550.
94. Min JW, Lee HY, Lee JS, et al. Effect of prolonged incubation time on results of the QuantiFERON TB gold in-tube assay for diagnosis of latent tuberculosis infection. *Clin Vaccine Immunol*. 2013 Sep;20(9):1377-80. Epub: 2013/07/05. PMID: 23825190.
95. Qian F, Wang W, Qiu Z, et al. Evaluation of a new tuberculosis-related interferon gamma release assay for tuberculosis infection diagnosis in Huzhou, eastern China. *Indian J Pathol Microbiol*. 2013 Apr-Jun;56(2):125-8. Epub: 2013/09/24. PMID: 24056648.
96. Wang S, Chen J, Zhang Y, et al. Mycobacterium tuberculosis region of difference (RD) 2 antigen Rv1985c and RD11 antigen Rv3425 have the promising potential to distinguish patients with active tuberculosis from M. bovis BCG-vaccinated individuals. *Clin Vaccine Immunol*. 2013 Jan;20(1):69-76. Epub: 2012/11/09. PMID: 23136116.
97. Lee YJ, Lee J, Kim YY, et al. Performance of whole-blood interferon-gamma release assay in patients admitted to the emergency department with pulmonary infiltrates. *BMC Infect Dis*. 2011;11:107. Epub: 2011/04/26. PMID: 21513568.
98. Włodarczyk M, Rudnicka W, Janiszewska-Drobinska B, et al. Interferon-gamma assay in combination with tuberculin skin test are insufficient for the diagnosis of culture-negative pulmonary tuberculosis. *PLoS One*. 2014;9(9):e107208. Epub: 2014/09/16. PMID: 25221998.
99. Erdem H, Ozturk-Engin D, Elaldi N, et al. The microbiological diagnosis of tuberculous meningitis: results of Haydarpasa-1 study. *Clin Microbiol Infect*. 2014 Oct;20(10):O600-8. Epub: 2014/05/23. PMID: 24849547.
100. Kim CH, Lim JK, Yoo SS, et al. Diagnostic performance of the QuantiFERON-TB Gold In-Tube assay and factors associated with nonpositive results in patients with miliary tuberculosis. *Clin Infect Dis*. 2014 Apr;58(7):986-9. Epub: 2014/01/25. PMID: 24457341.
101. Memish ZA, Mah MW, Mahmood SA, et al. Clinico-diagnostic experience with tuberculous lymphadenitis in Saudi Arabia. *Clin Microbiol Infect*. 2000 Mar;6(3):137-41. Epub: 2001/02/13. PMID: 11168089.
102. Kang YA, Lee HW, Hwang SS, et al. Usefulness of whole-blood interferon-gamma assay and interferon-gamma enzyme-linked immunospot assay in the diagnosis of active pulmonary tuberculosis. *Chest*. 2007 Sep;132(3):959-65. Epub: 2007/05/17. PMID: 17505029.

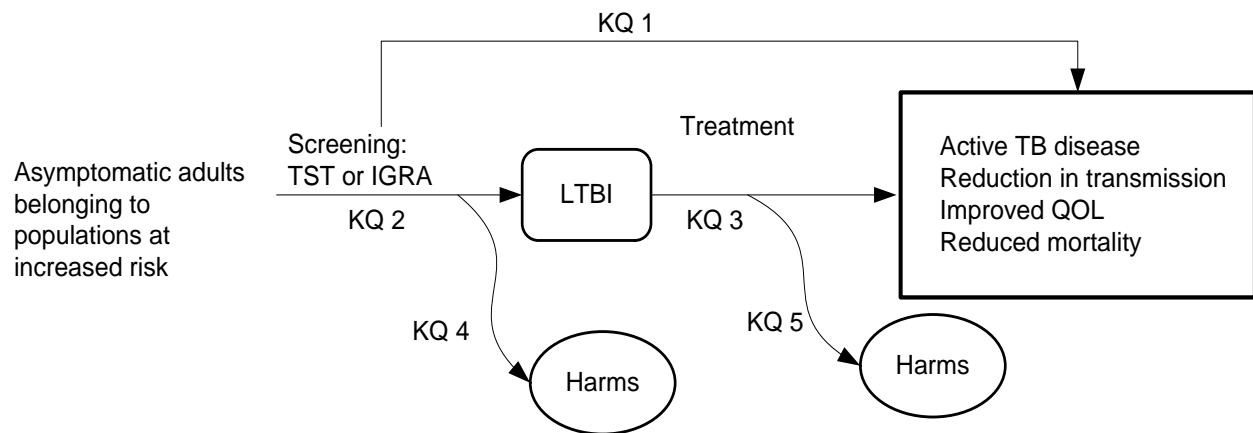
103. Ozekinci T, Ozbek E, Celik Y. Comparison of tuberculin skin test and a specific T-cell-based test, T-SPOT.*TB*, for the diagnosis of latent tuberculosis infection. *J Int Med Res*. 2007 Sep-Oct;35(5):696-703. Epub: 2007/10/20. PMID: 17944056.
104. Eum SY, Lee YJ, Kwak HK, et al. Evaluation of the diagnostic utility of a whole-blood interferon-gamma assay for determining the risk of exposure to *Mycobacterium tuberculosis* in Bacille Calmette-Guerin (BCG)-vaccinated individuals. *Diagn Microbiol Infect Dis*. 2008 Jun;61(2):181-6. Epub: 2008/02/26. PMID: 18296002.
105. Palazzo R, Spensieri F, Massari M, et al. Use of whole-blood samples in in-house bulk and single-cell antigen-specific gamma interferon assays for surveillance of *Mycobacterium tuberculosis* infections. *Clin Vaccine Immunol*. 2008 Feb;15(2):327-37. Epub: 2007/11/23. PMID: 18032595.
106. Kalantri Y, Hemvani N, Chitnis DS. Evaluation of whole blood IFNgamma test using PPD and recombinant antigen challenge for diagnosis of pulmonary and extra-pulmonary tuberculosis. *Indian J Exp Biol*. 2009 Jun;47(6):463-8. Epub: 2009/07/29. PMID: 19634712.
107. Kobashi Y, Shimizu H, Ohue Y, et al. False negative results of QuantiFERON TB-2G test in patients with active tuberculosis. *Jpn J Infect Dis*. 2009 Jul;62(4):300-2. Epub: 2009/07/25. PMID: 19628910.
108. Shalabi NM, Houssen ME. Discrepancy between the tuberculin skin test and the levels of serum interferon-gamma in the diagnosis of tubercular infection in contacts. *Clin Biochem*. 2009 Nov;42(16-17):1596-601. Epub: 2009/09/08. PMID: 19732759.
109. Shrestha R, Gyawali P, Yadav BK, et al. In-vitro assessment of cell-mediated immunity by demonstrating effector-t cells for diagnosis of tuberculosis in Nepalese subjects. *Nepal Med Coll J*. 2011 Dec;13(4):275-8. Epub: 2012/09/29. PMID: 23016479.
110. Li H, Yang L, Zheng CY, et al. Use of bronchoalveolar lavage enzyme-linked immunospot for diagnosis of smear-negative pulmonary tuberculosis. *Int J Tuberc Lung Dis*. 2012 Dec;16(12):1668-73. Epub: 2012/11/08. PMID: 23131267.
111. Turtle L, Kemp T, Davies GR, et al. In routine UK hospital practice T-SPOT.*TB* is useful in some patients with a modest pre-test probability of active tuberculosis. *Eur J Intern Med*. 2012 Jun;23(4):363-7. Epub: 2012/05/09. PMID: 22560387.
112. Kamiya H, Ikushima S, Kondo K, et al. Diagnostic performance of interferon-gamma release assays in elderly populations in comparison with younger populations. *J Infect Chemother*. 2013 Apr;19(2):217-22. Epub: 2012/10/31. PMID: 23108426.
113. Villarino ME, Burman W, Wang YC, et al. Comparable specificity of 2 commercial tuberculin reagents in persons at low risk for tuberculous infection. *Jama*. 1999 Jan 13;281(2):169-71. Epub: 1999/01/23. PMID: 9917121.
114. Villarino ME, Brennan MJ, Nolan CM, et al. Comparison testing of current (PPD-S1) and proposed (PPD-S2) reference tuberculin standards. *Am J Respir Crit Care Med*. 2000 Apr;161(4 Pt 1):1167-71. Epub: 2000/04/14. PMID: 10764307.
115. Mazurek GH, LoBue PA, Daley CL, et al. Comparison of a whole-blood interferon gamma assay with tuberculin skin testing for detecting latent *Mycobacterium tuberculosis* infection. *Jama*. 2001 Oct 10;286(14):1740-7. Epub: 2001/10/12. PMID: 11594899.
116. Bellete B, Coberly J, Barnes GL, et al. Evaluation of a whole-blood interferon-gamma release assay for the detection of *Mycobacterium tuberculosis* infection in 2 study populations. *Clin Infect Dis*. 2002 Jun 1;34(11):1449-56. Epub: 2002/05/17. PMID: 12015690.

117. Taggart EW, Hill HR, Ruegner RG, et al. Evaluation of an in vitro assay for gamma interferon production in response to *Mycobacterium tuberculosis* infections. *Clin Diagn Lab Immunol*. 2004 Nov;11(6):1089-93. Epub: 2004/11/13. PMID: 15539511.
118. Taggart EW, Hill HR, Ruegner RG, et al. Evaluation of an in vitro assay for interferon gamma production in response to the *Mycobacterium tuberculosis*-synthesized peptide antigens ESAT-6 and CFP-10 and the PPD skin test. *Am J Clin Pathol*. 2006 Mar;125(3):467-73. Epub: 2006/04/15. PMID: 16613353.
119. Bua A, Mollicotti P, Delogu G, et al. QuantiFERON TB Gold: a new method for latent tuberculosis infection. *New Microbiol*. 2007 Oct;30(4):477-80. Epub: 2007/12/18. PMID: 18080685.
120. Mazurek GH, Zajdowicz MJ, Hankinson AL, et al. Detection of *Mycobacterium tuberculosis* infection in United States Navy recruits using the tuberculin skin test or whole-blood interferon-gamma release assays. *Clin Infect Dis*. 2007 Oct 1;45(7):826-36. Epub: 2007/09/07. PMID: 17806046.
121. Bienek DR, Chang CK. Evaluation of an interferon-gamma release assay, T-SPOT.TB, in a population with a low prevalence of tuberculosis. *Int J Tuberc Lung Dis*. 2009 Nov;13(11):1416-21. Epub: 2009/10/29. PMID: 19861016.
122. Katsenos S, Nikolopoulou M, Konstantinidis AK, et al. Interferon-gamma release assay clarifies the effect of bacille Calmette-Guerin vaccination in Greek army recruits. *Int J Tuberc Lung Dis*. 2010 May;14(5):545-50. Epub: 2010/04/16. PMID: 20392346.
123. Mancuso JD, Mazurek GH, Tribble D, et al. Discordance among commercially available diagnostics for latent tuberculosis infection. *Am J Respir Crit Care Med*. 2012 Feb 15;185(4):427-34. Epub: 2011/12/14. PMID: 22161162.
124. Lempp JM, Margan JZ, Hankinson AL, et al. Assessment of the QuantiFERON-TB Gold In-Tube Test for the Detection of *Mycobacterium tuberculosis* Infection in US Navy Recruits. In: *Prevention CfDCA*, editor; 2015.
125. Saigal S, Agarwal SR, Nandeesh HP, et al. Safety of an ofloxacin-based antitubercular regimen for the treatment of tuberculosis in patients with underlying chronic liver disease: a preliminary report. *J Gastroenterol Hepatol*. 2001 Sep;16(9):1028-32. Epub: 2001/10/12. PMID: 11595068.
126. Franken WP, Timmermans JF, Prins C, et al. Comparison of Mantoux and QuantiFERON TB Gold tests for diagnosis of latent tuberculosis infection in Army personnel. *Clin Vaccine Immunol*. 2007 Apr;14(4):477-80. Epub: 2007/02/16. PMID: 17301213.
127. Dorman SE, Belknap R, Graviss EA, et al. Interferon-gamma release assays and tuberculin skin testing for diagnosis of latent tuberculosis infection in healthcare workers in the United States. *Am J Respir Crit Care Med*. 2014 Jan 1;189(1):77-87. Epub: 2013/12/05. PMID: 24299555.
128. Franken WP, Thijsen S, Wolterbeek R, et al. Variation in T-SPOT.TB spot interpretation between independent observers from different laboratories. *Clin Vaccine Immunol*. 2009 Oct;16(10):1439-42. Epub: 2009/08/28. PMID: 19710293.
129. Whitworth WC, Hamilton LR, Goodwin DJ, et al. Within-subject interlaboratory variability of QuantiFERON-TB gold in-tube tests. *PLoS One*. 2012;7(9):e43790. Epub: 2012/09/13. PMID: 22970142.
130. Whitworth WC, Goodwin DJ, Racster L, et al. Variability of the QuantiFERON(R)-TB gold in-tube test using automated and manual methods. *PLoS One*. 2014;9(1):e86721. Epub: 2014/01/28. PMID: 24466211.

131. O'Shea MK, Fletcher TE, Beeching NJ, et al. Tuberculin skin testing and treatment modulates interferon-gamma release assay results for latent tuberculosis in migrants. *PLoS One*. 2014;9(5):e97366. Epub: 2014/05/13. PMID: 24816576.
132. Cummings KJ, Smith TS, Shogren ES, et al. Prospective comparison of tuberculin skin test and QuantiFERON-TB Gold In-Tube assay for the detection of latent tuberculosis infection among healthcare workers in a low-incidence setting. *Infect Control Hosp Epidemiol*. 2009 Nov;30(11):1123-6. Epub: 2009/10/07. PMID: 19803719.
133. Menzies D, Long R, Trajman A, et al. Adverse events with 4 months of rifampin therapy or 9 months of isoniazid therapy for latent tuberculosis infection: a randomized trial. *Ann Intern Med*. 2008 Nov 18;149(10):689-97. Epub: 2008/11/20. PMID: 19017587.
134. Sterling TR, Villarino ME, Borisov AS, et al. Three months of rifapentine and isoniazid for latent tuberculosis infection. *N Engl J Med*. 2011 Dec 8;365(23):2155-66. Epub: 2011/12/14. PMID: 22150035.
135. Thompson MJ. Efficacy of various durations of isoniazid preventive therapy for tuberculosis: five years of follow-up in the IUAT trial. *International Union Against Tuberculosis Committee on Prophylaxis. Bull World Health Organ*. 1982;60(4):555-64. Epub: 1982/01/01. PMID: 6754120.
136. Mount FW, Ferebee SH. The effect of isoniazid prophylaxis on tuberculosis morbidity among household contacts of previously known cases of tuberculosis. *Am Rev Respir Dis*. 1962 Jun;85:821-7. Epub: 1962/06/01. PMID: 14476668.
137. Ferebee SH, Mount FW. Tuberculosis morbidity in a controlled trial of the prophylactic use of isoniazid among household contacts. *Am Rev Respir Dis*. 1962 Apr;85:490-510. Epub: 1962/04/01. PMID: 13892318.
138. Egsmose T, Ang'awa JO, Poti SJ. The use of isoniazid among household contacts of open cases of pulmonary tuberculosis. *Bull World Health Organ*. 1965;33(3):419-33. Epub: 1965/01/01. PMID: 5321762.
139. Horwitz O, Payne PG, Wilbek E. Epidemiological basis of tuberculosis eradication. 4. The isoniazid trial in Greenland. *Bull World Health Organ*. 1966;35(4):509-26. Epub: 1966/01/01. PMID: 5335457.
140. Comstock GW, Ferebee SH, Hammes LM. A controlled trial of community-wide isoniazid prophylaxis in Alaska. *Am Rev Respir Dis*. 1967 Jun;95(6):935-43. Epub: 1967/06/01. PMID: 6026165.
141. Girling DJ, Chan SL, Hong Kong Chest Service/Tuberculosis Research Centre, Madras/British Medical Research Council. A double-blind placebo-controlled clinical trial of three antituberculosis chemoprophylaxis regimens in patients with silicosis in Hong Kong. *Am Rev Respir Dis*. 1992;145:36-41.
142. John GT, Thomas PP, Thomas M, et al. A double-blind randomized controlled trial of primary isoniazid prophylaxis in dialysis and transplant patients. *Transplantation*. 1994 Jun 15;57(11):1683-4. Epub: 1994/06/15. PMID: 8009608.
143. Menzies D, Dion MJ, Rabinovitch B, et al. Treatment completion and costs of a randomized trial of rifampin for 4 months versus isoniazid for 9 months. *Am J Respir Crit Care Med*. 2004 Aug 15;170(4):445-9. Epub: 2004/06/03. PMID: 15172892.
144. White MC, Tulskey JP, Lee JR, et al. Isoniazid vs. rifampin for latent tuberculosis infection in jail inmates: toxicity and adherence. *J Correct Health Care*. 2012 Apr;18(2):131-42. Epub: 2012/03/16. PMID: 22419641.

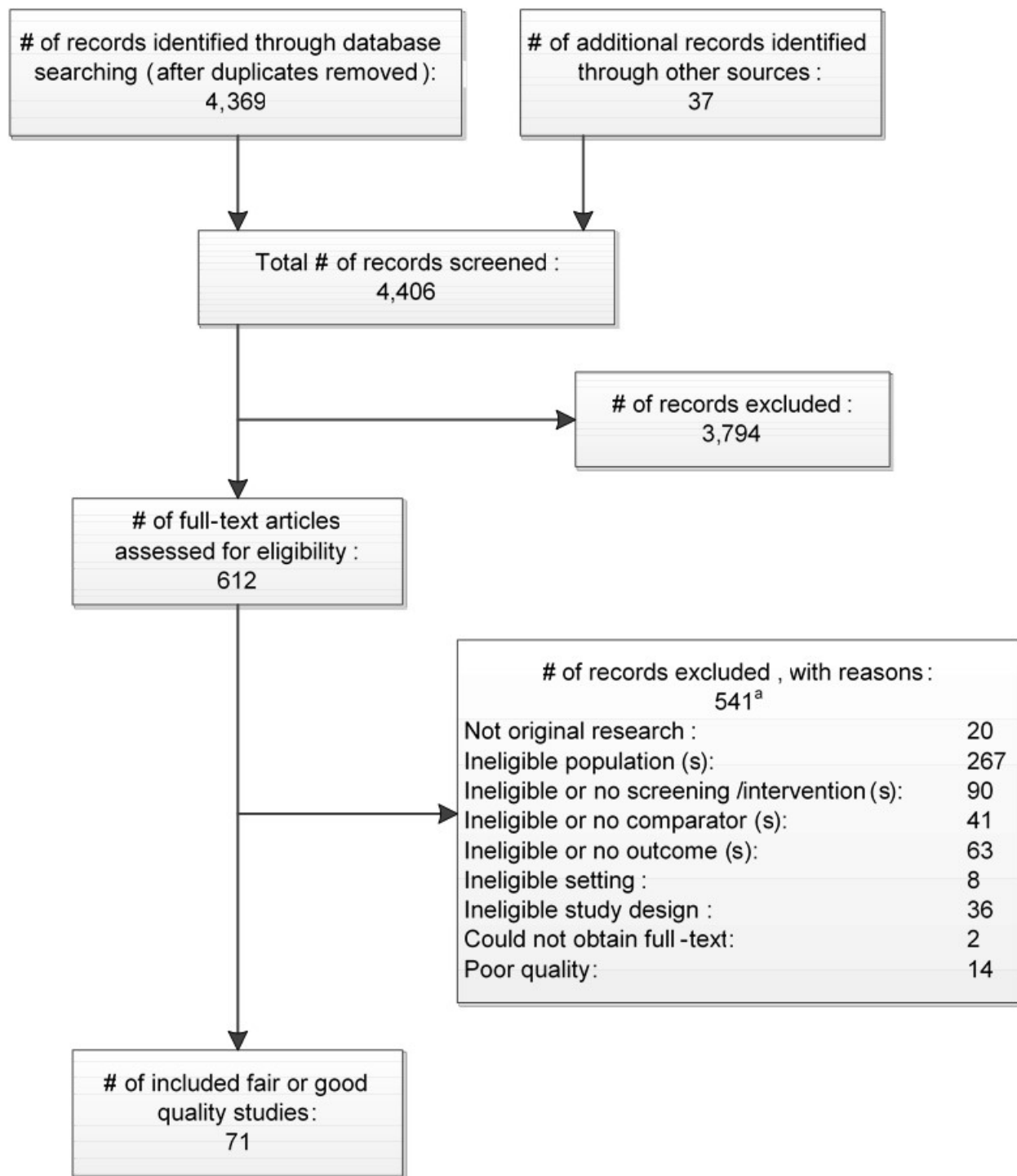
145. Bailey WC, Weill H, DeRouen TA, et al. The effect of isoniazid on transaminase levels. *Ann Intern Med.* 1974 Aug;81(2):200-2. Epub: 1974/08/01. PMID: 4843577.
146. Byrd RB, Horn BR, Griggs GA, et al. Isoniazid chemoprophylaxis. Association with detection and incidence of liver toxicity. *Arch Intern Med.* 1977 Sep;137(9):1130-3. Epub: 1977/09/01. PMID: 332099.
147. Krebs A. The IUAT trial on isoniazid preventive treatment in persons with fibrotic lung lesions. *Bull Int Union Tuberc.* 1976;51(1):193-201. Epub: 1976/01/01. PMID: 801115.
148. Falk A, Fuchs G. Isoniazid (INH) prophylaxis in inactive pulmonary tuberculosis: report of a Veterans Administration Cooperative Study. *Bull Int Union Tuberc.* 1976;51(1):219-23. Epub: 1976/01/01. PMID: 1030286.
149. Menzies D, Pai M, Comstock G. Meta-analysis: new tests for the diagnosis of latent tuberculosis infection: areas of uncertainty and recommendations for research. *Ann Intern Med.* 2007 Mar 6;146(5):340-54. Epub: 2007/03/07. PMID: 17339619.
150. Pai M, Zwerling A, Menzies D. Systematic review: T-cell-based assays for the diagnosis of latent tuberculosis infection: an update. *Ann Intern Med.* 2008 Aug 5;149(3):177-84. Epub: 2008/07/03. PMID: 18593687.
151. Diel R, Loddenkemper R, Nienhaus A. Evidence-based comparison of commercial interferon-gamma release assays for detecting active TB: a metaanalysis. *Chest.* 2010 Apr;137(4):952-68. Epub: 2009/12/22. PMID: 20022968.
152. Diel R, Goletti D, Ferrara G, et al. Interferon-gamma release assays for the diagnosis of latent *Mycobacterium tuberculosis* infection: a systematic review and meta-analysis. *Eur Respir J.* 2011 Jan;37(1):88-99. Epub: 2010/10/30. PMID: 21030451.
153. Linas BP, Wong AY, Freedberg KA, et al. Priorities for screening and treatment of latent tuberculosis infection in the United States. *Am J Respir Crit Care Med.* 2011 Sep 1;184(5):590-601. Epub: 2011/05/13. PMID: 21562129.
154. Stagg HR, Zenner D, Harris RJ, et al. Treatment of latent tuberculosis infection: a network meta-analysis. *Ann Intern Med.* 2014 Sep 16;161(6):419-28. PMID: 25111745.
155. Smieja MJ, Marchetti CA, Cook DJ, et al. Isoniazid for preventing tuberculosis in non-HIV infected persons. *Cochrane Database Syst Rev.* 2000(2):Cd001363. Epub: 2000/05/05. PMID: 10796642.
156. Polesky A, Farber HW, Gottlieb DJ, et al. Rifampin preventive therapy for tuberculosis in Boston's homeless. *Am J Respir Crit Care Med.* 1996 Nov;154(5):1473-7. Epub: 1996/11/01. PMID: 8912767.

Figure 1. Analytic Framework: Screening for Latent Tuberculosis Infection in Adults



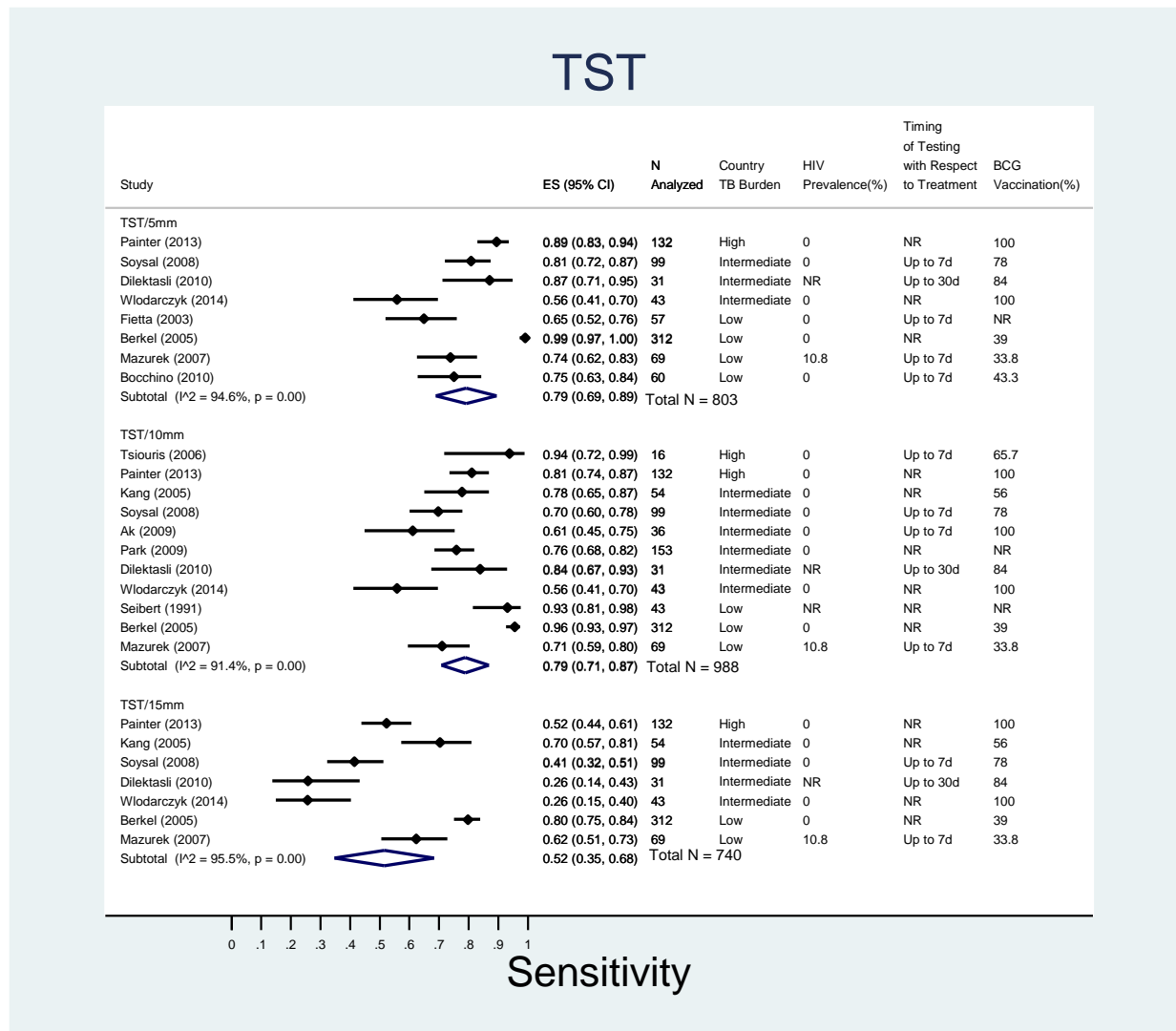
IGRA = interferon gamma release assay; KQ = key question; LTBI = latent tuberculosis infection; QOL = quality of life; TB = tuberculosis; TST = tuberculin skin test.

Figure 2. Preferred Reporting of Systematic Review and Meta-Analysis (PRISMA) Tree



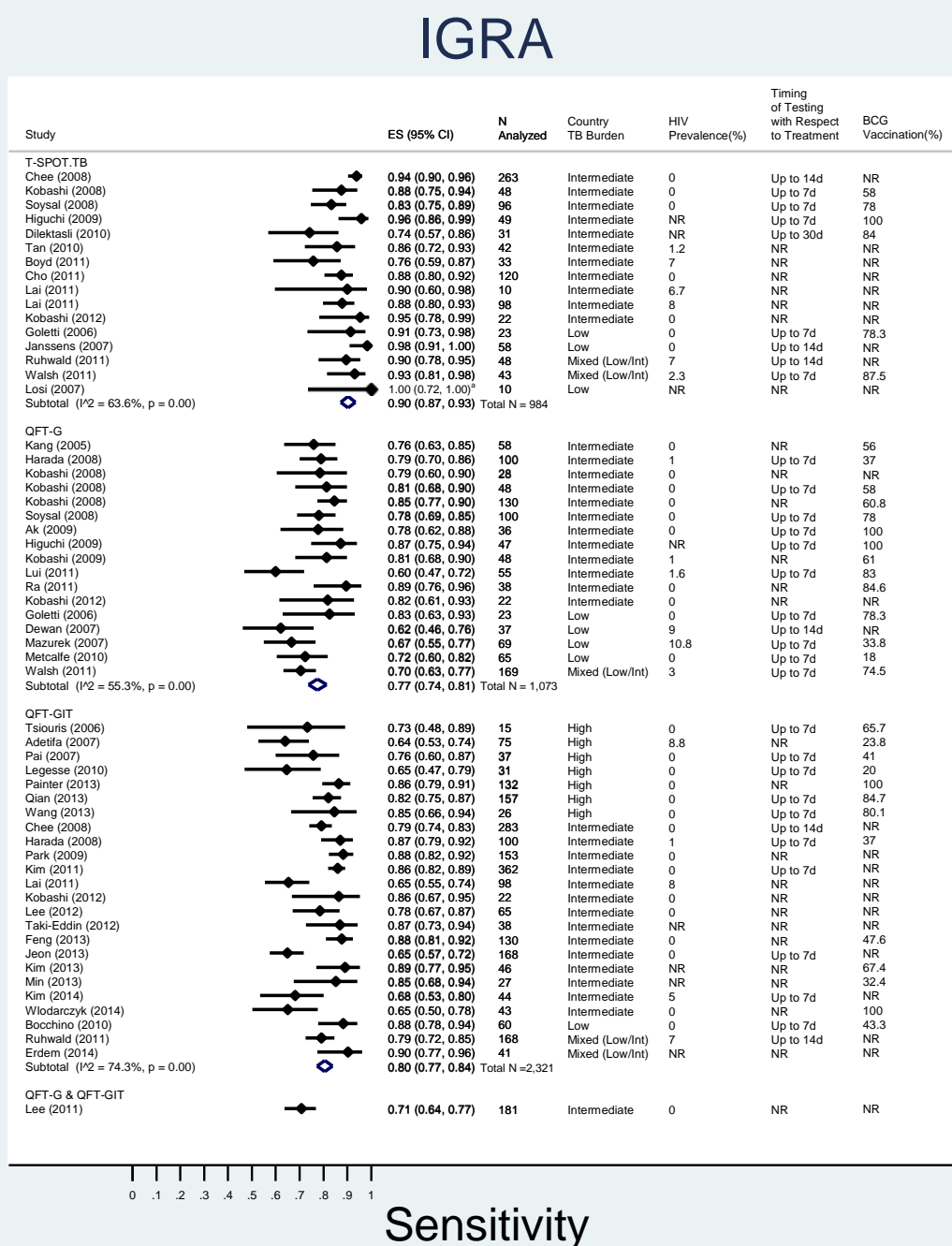
^a 19 poor quality and/or ineligible studies were excluded but used in sensitivity analyses

Figure 3. Individual Study and Pooled Estimates of Sensitivity for Various Thresholds of the TST for Tuberculosis Infection



BCG = Bacillus Calmette-Guerin; CI = confidence interval; d = day; ES = effect size; HIV = Human immunodeficiency virus; mm = millimeter; N = number; NR = not reported; TB = tuberculosis; TST = tuberculin skin test.

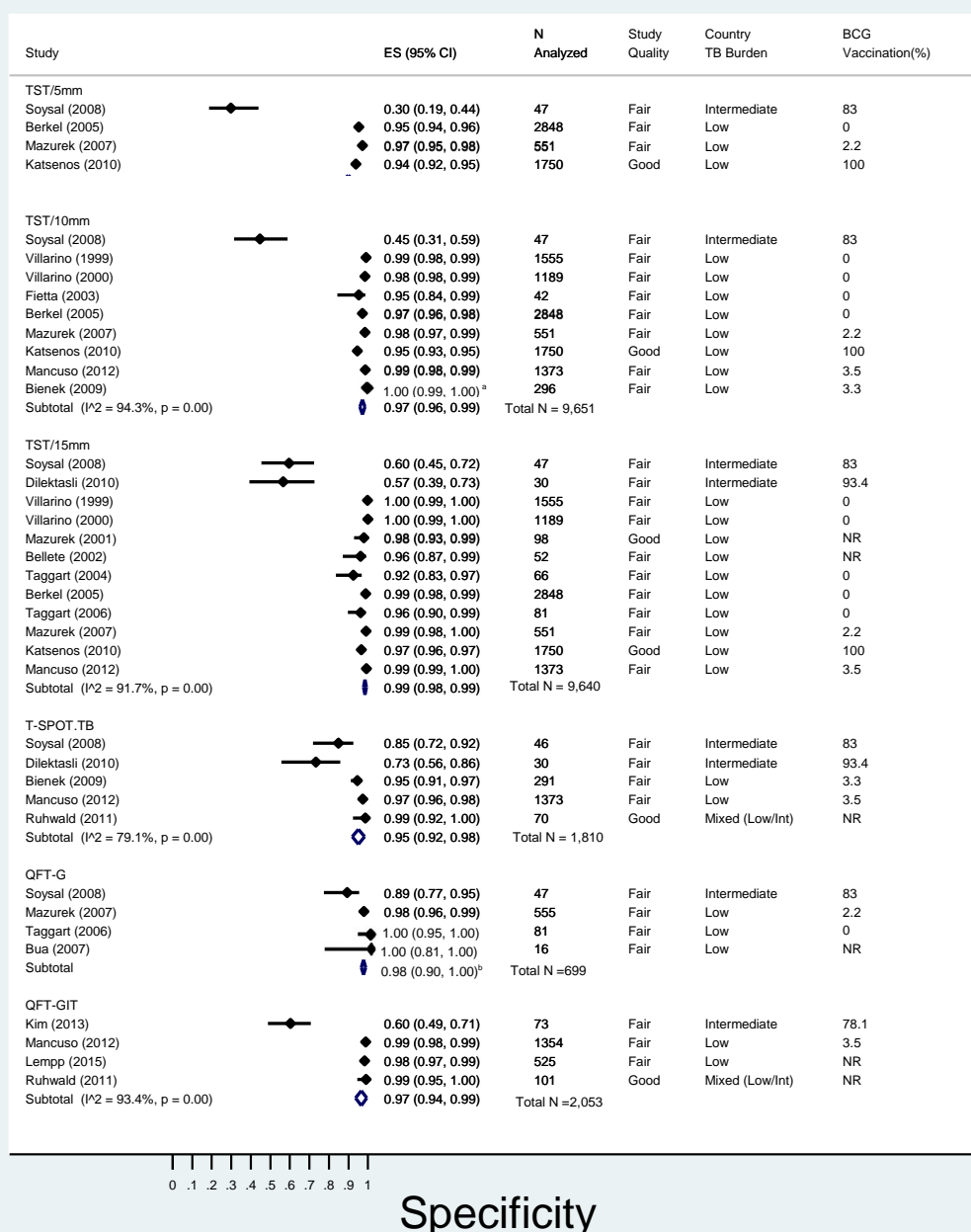
Figure 4. Individual Study and Pooled Estimates (Where Applicable) of Sensitivity for IGRA Tests for Tuberculosis Infection



^a Excluded from pooled estimate due to point estimate of 1.0.

BCG = Bacillus Calmette-Guerin; CI = confidence interval; ES = effect size; HIV = Human immunodeficiency virus; Int = intermediate; mm = millimeter; N = number; NR = not reported; QFT-G = QuantiFERON TB Gold[®] test (2nd generation test); QFT-GIT = QuantiFERON TB Gold-In-Tube[®] test (3rd generation test); TB = tuberculosis; T-SPOT.TB = Commercial ELISPOT Assay.

Figure 5. Individual Study and Pooled Estimates (Where Applicable) of Specificity for Various Thresholds of TST and IGRA Tests for Tuberculosis Infection

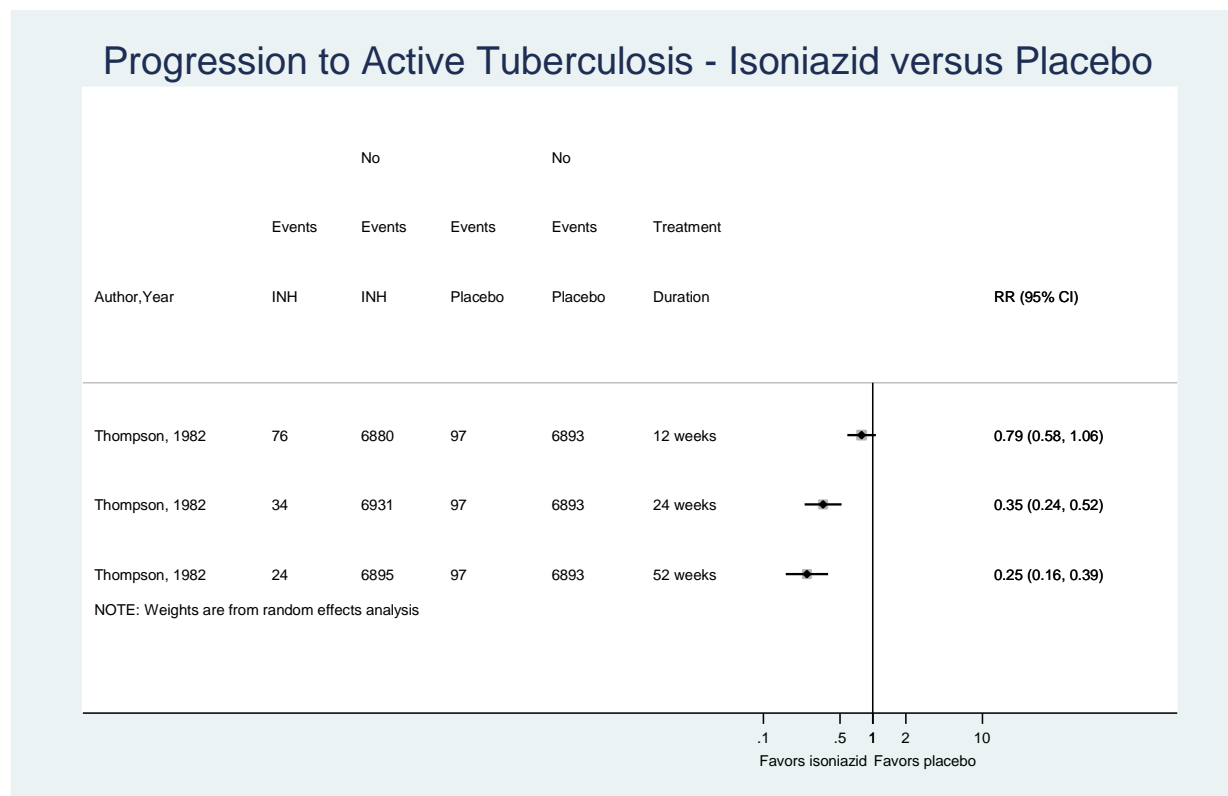


^a Excluded from pooled estimate due to point estimate of 1.0.

^b Pooled estimate from maximum likelihood estimate random effects model because of two studies with point estimates of 1.0. No I^2 statistic is calculated using this model.

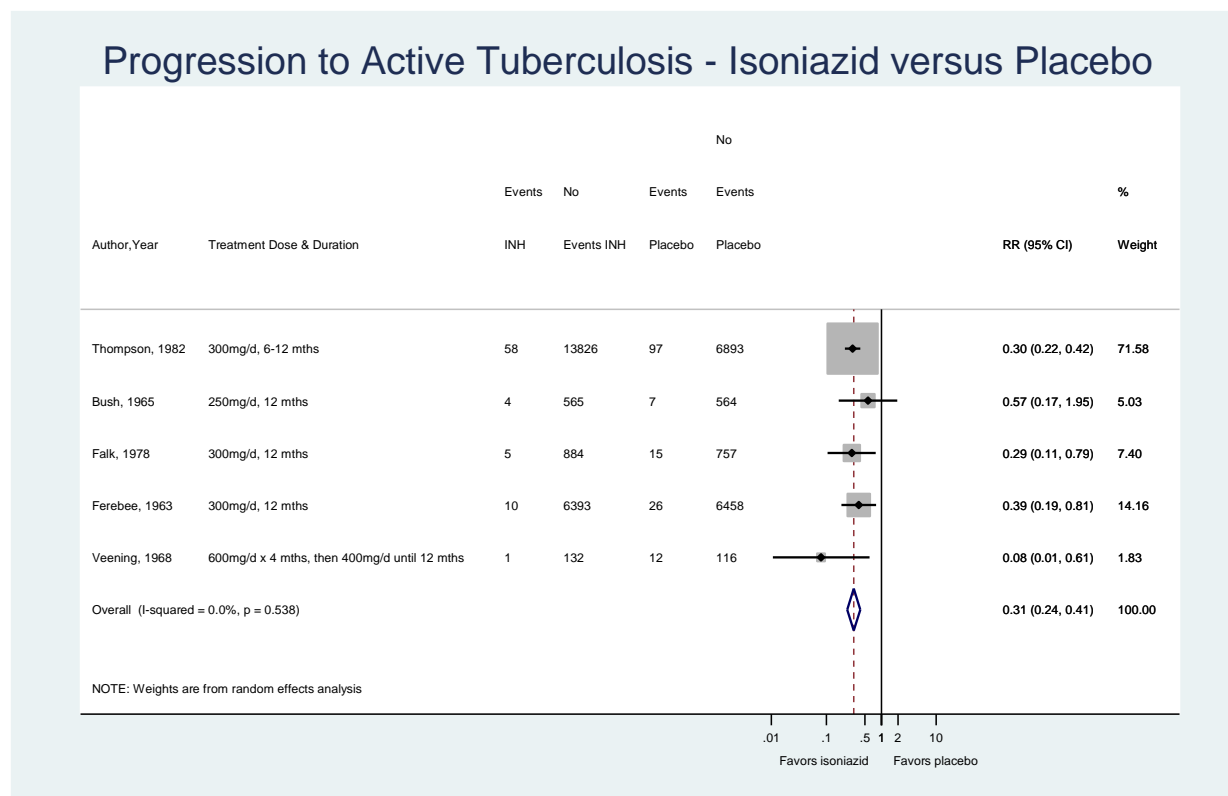
BCG = Bacillus Calmette-Guerin; CI = confidence interval; ES = effect size; Int = intermediate; mm = milligram; N = number; NR = not reported; QFT-G = QuantiFERON TB Gold[®] test (2nd generation test); QFT-GIT = QuantiFERON TB Gold-In-Tube[®] test (3rd generation test); TB = tuberculosis; T-SPOT.TB = commercial ELISPOT Assay; TST = tuberculin skin test.

Figure 6. Isoniazid Compared With Placebo, Relative Risk of Developing Active Tuberculosis in the IUAT Trial



CI = confidence interval; INH = isoniazid; RR = relative risk.

Figure 7. Isoniazid Compared With Placebo, Relative Risk of Developing Active Tuberculosis: Sensitivity Analysis Including Data From the IUAT Trial and Four Additional RCTs

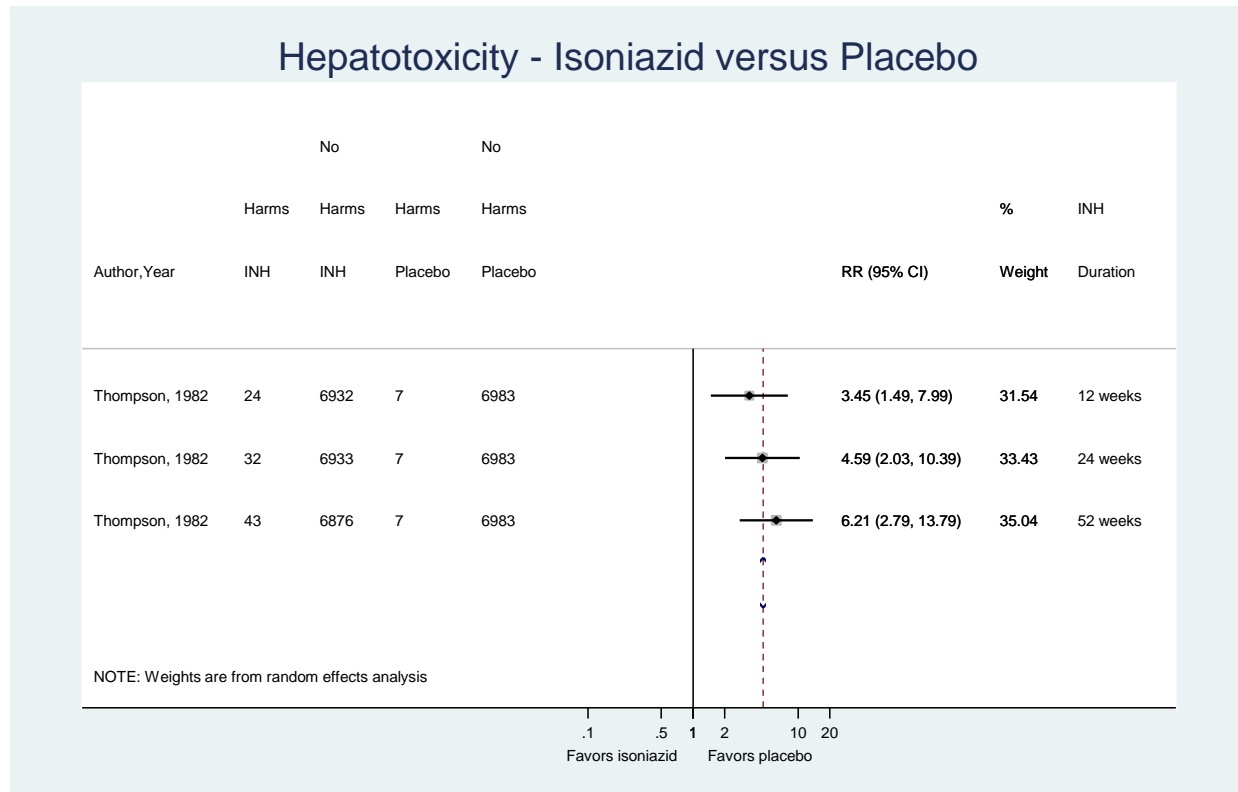


Notes: For Thompson, 1982¹³⁵ (IUAT trial), we included data from the 24- and 52-week groups. For Bush 1965,⁴³ we only used data for those ≥ 20 years of age. For Falk 1978,⁴¹ we used data for the subset with no previous TB therapy for participants in the isoniazid 1-year group (we did not include data for the isoniazid 2-year group). For Ferebee 1963,⁴⁴ we used only the subset that was tuberculin positive; we were unable to get adult-only data to enter here (for the full study sample, 34 of the 51 cases in the placebo arm were among adults, and it was not reported how many of the 19 total cases in the isoniazid arm of the study were among adults).

For RCTs other than the IUAT trial to be included in this sensitivity analysis, we required that they either confirmed LTBI for subjects to be eligible, reported data for those with confirmed LTBI, or that the vast majority of subjects (over 75 percent) were tuberculin positive. These trials met many of our eligibility criteria, but they all used a longer duration of treatment than is currently recommended by CDC (i.e., they used 1 year or longer of isoniazid), and some used lower or higher doses than currently recommended^{42,43} or did not require LTBI confirmation for subjects to be eligible.^{41,43,44} One of the four trials was rated poor quality.⁴²

CI = confidence interval; INH = isoniazid; mg/d = milligrams per day; mths = months; RR = relative risk.

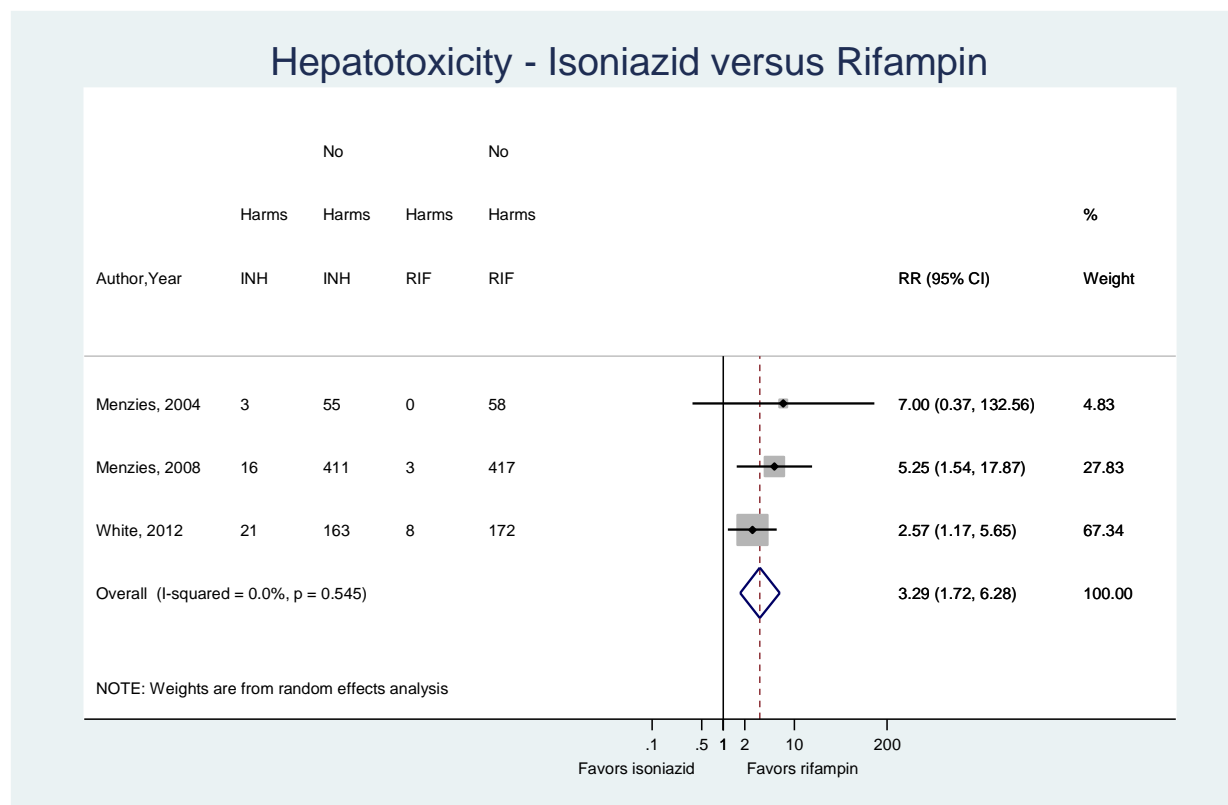
Figure 8. Isoniazid Compared With Placebo, Relative Risk of Developing Hepatotoxicity in the IUAT Trial



Notes: For Thompson, 1982¹³⁵ (IUAT trial), we included data from the 12-, 24-, and 52-week groups. A definition for hepatotoxicity (presented as “hepatitis” in this study) was not reported.

CI = confidence interval; INH= isoniazid; RR = relative risk.

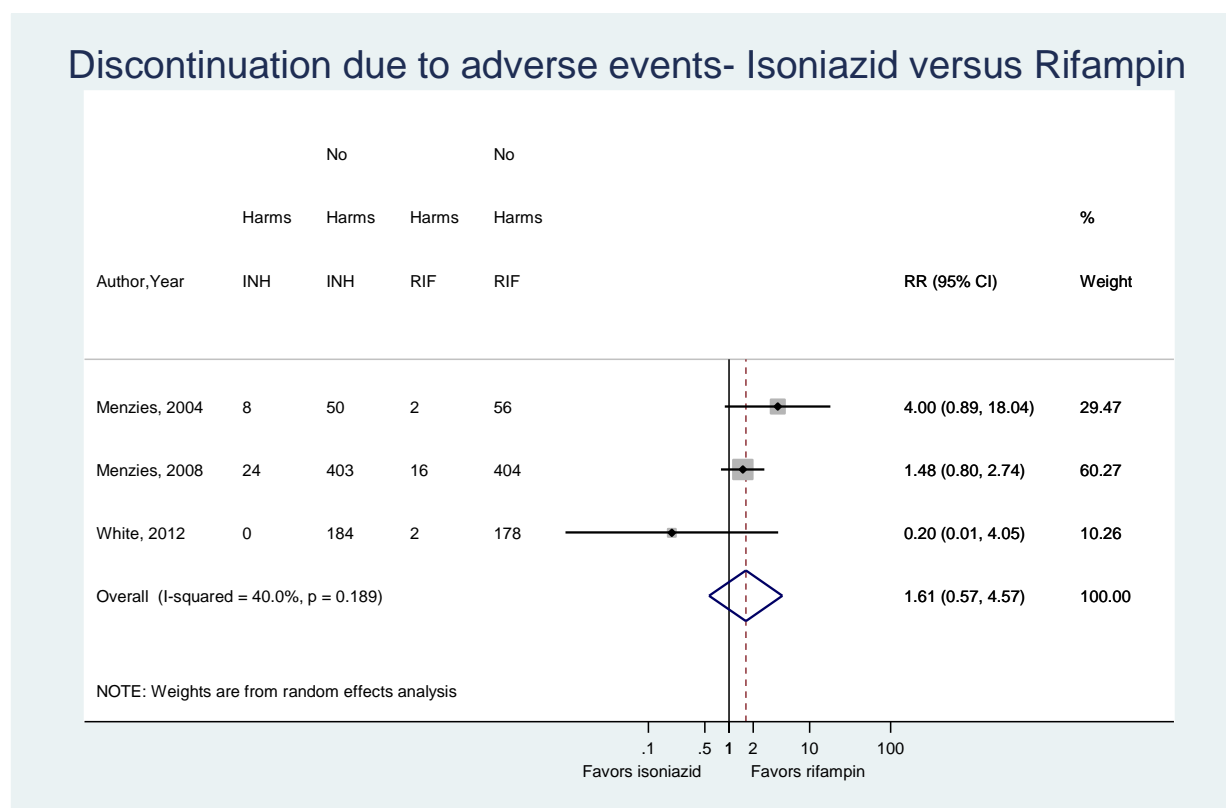
Figure 9. Isoniazid Compared With Rifampin, Relative Risk of Developing Hepatotoxicity: Data From Three RCTs



Notes: For Menzies, 2004, hepatotoxicity was defined as liver transaminase (alanine transaminase) levels more than three times the upper limits of normal with symptoms, or transaminase levels more than 5 times the upper limits of normal without symptoms. For Menzies, 2008, hepatotoxicity includes both grade 3 and grade 4 hepatotoxicity. Liver aminotransferase levels that increased to 5 to 10 or 3 to 10 times the upper limit of normal in the presence of compatible symptoms met criteria for grade 3 hepatotoxicity, whereas those that exceeded 10 times the upper limit of normal met criteria for grade 4 toxicity. For White, 2012, hepatotoxicity was defined as liver function tests (LFTs) greater than 3 times the upper limit of normal.

CI = confidence interval; INH = isoniazid; RIF = rifampin; RR = relative risk.

Figure 10. Isoniazid Compared With Rifampin, Relative Risk of Treatment Discontinuation Due to Adverse Events: Data From Three RCTs



Notes: For Menzies, 2004, adverse events that resulted in permanent discontinuation of therapy were hepatitis, severe nausea and vomiting, persistent debilitating fatigue, and rash. For Menzies, 2008, a blinded review panel judged the type and severity of the adverse events and its likely relationship to the study drug. The total presented reflects permanent discontinuation of therapy due to any adverse event (grade 1–4) judged to be probably drug-related. These adverse events were hepatotoxicity, hematologic, drug interaction, rash, and gastrointestinal intolerance. For White, 2012, treatment discontinuation adverse events were elevated liver function tests and nausea/vomiting.

CI = confidence interval; INH= isoniazid; RIF = rifampin; RR = relative risk.

Table 1. Summary of Sensitivity Estimates for Various Thresholds of Tuberculin Skin Test and Interferon Gamma-Release Assays Among Patients With Bacteriologic-Confirmed Tuberculosis

Test	Number of Studies (Total N)	Pooled Sensitivity Estimate (95% CI), I^2	Individual Study Sensitivity Estimates (95% CI), N
TST (5-mm threshold)	8 (803)	0.79 (0.69 to 0.89) ^a , 94.6%	See Figure 3
TST (10-mm threshold)	11 (988)	0.79 (0.71 to 0.87), 91.4%	See Figure 3
TST (15-mm threshold)	7 (740)	0.52 (0.35 to 0.68), 95.5%	See Figure 3
IGRA; T-SPOT.TB	16 ^b (984)	0.90 (0.87 to 0.93), 63.6%	See Figure 4
IGRA; QFT-G	17 (1,073)	0.77 (0.74 to 0.81), 55.3%	See Figure 4
IGRA; QFT-GIT	24 (2,321)	0.80 (0.77 to 0.84), 74.3%	See Figure 4
IGRA; QFT-G & GIT	1	NA	0.71 (0.64 to 0.77), 181 ⁹⁷

^a Estimates from a maximum likelihood random effects model yielded slightly different estimate (0.84; 95% CI, 0.68 to 0.92).

^b One study⁶¹ could not be included in the pooled estimate due to a point estimate for sensitivity of 1.0 (95% CI, 0.69 to 1.0). The estimate using the maximum likelihood approach, which can accommodate point estimates of 1.0, was similar (pooled Sn 0.90; 95% CI, 0.86 to 0.93).

CI = confidence interval; I^2 = the proportion of variation in study estimates due to heterogeneity; IGRA = interferon gamma release assay; mm = millimeter; N = number of patients; NA = not applicable; QFT-G= QuantiFERON TB Gold[®] test (2nd generation test); QFT-GIT= QuantiFERON TB Gold-In-Tube[®] test (3rd generation test); Sn = sensitivity; T-SPOT.TB = Commercial ELISPOT Assay; TST = tuberculin skin test.

Table 2. Summary of Specificity Estimates for Various Thresholds of Tuberculin Skin Test and Interferon Gamma-Release Assays Among Healthy Subjects Without Tuberculosis Exposures or Risks

Test	Number of Studies (Total N for pooled studies)	Pooled Specificity Estimate (95% CI), I ²	Individual Study Estimate Specificity (95% CI), N
TST (5-mm threshold)	4	NA ^a	0.30 (0.19 to 0.44), 47 ⁶⁹ 0.95 (0.94 to 0.96), 2,848 ⁵⁴ 0.94 (0.92 to 0.95), 1,750 ¹²² 0.97 (0.95 to 0.98), 551 ¹²⁰
TST (10-mm threshold)	9 ^b (9,651)	0.97 (0.96 to 0.99), 94.3%	See Figure 5
TST (15-mm threshold)	12 (9,640)	0.99 (0.98 to 0.99), 91.7%	See Figure 5
IGRA; T-SPOT.TB	5 (1,810)	0.95 (0.92 to 0.98) ^c , 79.1%	See Figure 5
IGRA; QFT-G	4 (699)	0.98 ^d (0.90, 1.0)	See Figure 5
IGRA; QFT-GIT	4 (2,053)	0.97 (0.94 to 0.99), 93.4%	See Figure 5

^a Studies not pooled as one study estimate from intermediate TB burden country was much lower than the estimates from low TB burden countries.

^b One study¹²¹ could not be included in the DerSimonian-Laird pooled estimate because of a point estimate for specificity of 1.0 (95% CI, 0.99 to 1.00). The estimate using the maximum likelihood approach, which can accommodate point estimates of 1.0, was similar (pooled Sp, 0.97; 95% CI, 0.93 to 0.99).

^c Estimates from a maximum likelihood random effects model yielded a slightly different estimate (0.93; 95% CI, 0.85 to 0.97).

^d Pooled estimate is from maximum likelihood random effects model as two studies included point estimates for specificity of 1.0. The I² statistic is not calculated when using this model.

CI = confidence interval; I² = the proportion of variation in study estimates due to heterogeneity; IGRA = interferon gamma release assay; mm = millimeter; N = number analyzed; NA = not applicable; QFT-G= QuantiFERON TB Gold® test (2nd generation test); QFT-GIT = QuantiFERON TB Gold-In-Tube® test (3rd generation test); Sp = specificity; T-SPOT.TB = Commercial ELISPOT Assay; TST = tuberculin skin test.

Table 3. Summary of Evidence of Accuracy and Reliability of TST or IGRA Tests for Targeted^a Screening of Latent Tuberculosis Infection—KQ 2a

Test or Intervention	Number of Studies (observations) Study Designs By Test or Outcome	Summary of Findings By Test or Outcome	Consistency/ Precision	Reporting Bias	Overall Quality	Body of Evidence Limitations	Applicability
TST 5-mm Accuracy	Sn 8 (803)	Sn pooled 0.79 (95% CI, 0.69 to 0.89, $I^2=94.6\%$)	Consistent but imprecise for Sn	Undetected	Fair	Independent interpretation of test often not reported	TST using Mantoux procedure with intermediate-strength dose of PPD
	Sp 4 (5,196) Observational studies of test accuracy	Sp in low TB-burden countries 0.94 (95% CI, 0.92 to 0.95) 0.95 (95% CI, 0.94 to 0.96) 0.97 (95% CI, 0.95 to 0.98) Sp in intermediate TB burden country 0.30 (95% CI, 0.19 to 0.44)	Consistent and precise for Sp in low TB-burden countries			Description of participant characteristics highly variable across studies	Lack of direct test for LTBI requires extrapolation of test characteristics from active TB (Sn) and healthy, low-risk populations (Sp)
TST 10-mm Accuracy	Sn 11 (988)	Sn pooled 0.79 (95% CI, 0.71 to 0.87, $I^2=91.4\%$)	Consistent but imprecise for Sn	Undetected	Fair	Independent interpretation of test often not reported	TST using Mantoux procedure with intermediate-strength dose of PPD
	Sp 9 (9,651) Observational studies of test accuracy	Sp pooled 0.97 (95% CI, 0.96 to 0.99, $I^2=94.3\%$) ^b	Consistent and precise for Sp in low TB-burden countries			Description of participant characteristics highly variable across studies	Lack of direct test for LTBI requires extrapolation of test characteristics from active TB (Sn) and healthy, low-risk populations (Sp)
TST 15-mm Accuracy	Sn 7 (740)	Sn pooled 0.52 (95% CI, 0.35 to 0.68, $I^2=96.3\%$)	Inconsistent and imprecise for Sn	Undetected	Fair	Independent interpretation of test often not reported	TST using Mantoux procedure with intermediate-strength dose of PPD
	Sp 12 (9,640) Observational studies of test accuracy	Sp pooled 0.99 (95% CI, 0.98 to 0.99, $I^2=91.7\%$) ^c	Consistent and precise for Sp in low TB-burden countries			Description of participant characteristics highly variable across studies	Lack of direct test for LTBI requires extrapolation of test characteristics from active TB (Sn) and healthy, low-risk populations (Sp) The 15-mm threshold is not recommended in current practice for patients at high risk for TB infection

Table 3. Summary of Evidence of Accuracy and Reliability of TST or IGRA Tests for Targeted^a Screening of Latent Tuberculosis Infection—KQ 2a

Test or Intervention	Number of Studies (observations) Study Designs By Test or Outcome	Summary of Findings By Test or Outcome	Consistency/ Precision	Reporting Bias	Overall Quality	Body of Evidence Limitations	Applicability
TST Reliability	Interrater reliability 3 (3,142) Observational studies of test accuracy	Kappa 0.69 and 0.79 in two studies assessing reliability of rater assessment of skin test reaction in healthy populations at low risk for TB Kappa 0.52 to 0.78 of rater assessment of skin test reaction as assessed in different study with populations including subjects with active TB and healthy, low -risk subjects	Consistent for moderate to substantial agreement; precision unknown	Undetected	Fair	Reliability may be affected by the populations in which it is assessed	TST using Mantoux procedure with intermediate-strength dose of PPD TST administration and interpretation dependent on the use of appropriate, standardized technique
IGRA T-SPOT.TB Accuracy	Sn 16 (984) Sp 5 (1,810) Observational studies of test accuracy	Sn pooled 0.90 (95% CI, 0.87 to 0.93, $I^2=63.6\%$) Sp pooled 0.95 (95% CI, 0.92 to 0.98, $I^2=79.1\%$) ^d	Consistent and precise for Sn and Sp	Undetected	Fair	Independent interpretation of test often not reported; description of participant characteristics highly variable across studies Studies vary with respect to how they report indeterminate results	Lack of direct test for LTBI requires extrapolation of test characteristics from active TB (Sn) and healthy, low-risk (Sp) populations T-SPOT.TB requires proper specimen handling prior to assay; interpretation of test can be done manually through visual inspection, or through use of machines that automate interpretation FDA-approved threshold for positive test is higher than threshold used in non-U.S. studies

Table 3. Summary of Evidence of Accuracy and Reliability of TST or IGRA Tests for Targeted^a Screening of Latent Tuberculosis Infection—KQ 2a

Test or Intervention	Number of Studies (observations) Study Designs By Test or Outcome	Summary of Findings By Test or Outcome	Consistency/Precision	Reporting Bias	Overall Quality	Body of Evidence Limitations	Applicability
IGRA T-SPOT. TB Reliability	Interrater reliability 2 (404)	1 study conducted in active TB patients with manual interpretation: interrater reliability 96% (kappa 0.92), manual vs. automatic interpretation: Interrater reliability 85.8% (kappa 0.73)	Consistent for interrater reliability, unknown precision	Undetected	Fair	Independent interpretation of test often not reported; description of participant characteristics highly variable across studies	T-SPOT. TB requires proper specimen handling prior to assay; interpretation of test can be done manually through visual inspection, or through use of machine that automates interpretation
	Reproducibility 1(130)	1 study conducted among immigrants who were close contacts of active TB patients with kappa > 0.6 among 6 manual readers Discordant results in participants who had 2 samples drawn simultaneously (same lab and method of interpretation): 10/153 (6.5%)	Consistency unknown for single study, unknown precision			Studies vary with respect to how they report indeterminate results	
	Test-retest 2 (296)	1 study enrolling HCWs: 9/111 (8.1%) tests changed from negative to positive and 10/19 (52.6%) changed from positive to negative at 2 weeks. 1 study enrolling Nepalese military recruits, kappa for agreement between initial test and retest= 0.66 (95% CI, 0.50 to 0.83)	Inconsistent and imprecise for test-retest reliability				
	Observational studies of test accuracy						

Table 3. Summary of Evidence of Accuracy and Reliability of TST or IGRA Tests for Targeted^a Screening of Latent Tuberculosis Infection—KQ 2a

Test or Intervention	Number of Studies (observations) Study Designs By Test or Outcome	Summary of Findings By Test or Outcome	Consistency/ Precision	Reporting Bias	Overall Quality	Body of Evidence Limitations	Applicability
IGRA QFT-G Accuracy	Sn 17 (1,073)	Sn pooled 0.77 (95% CI, 0.74 to 0.81, $I^2=55.3\%$)	Consistent and precise for Sn	Undetected	Fair	Independent interpretation of test often not reported; description of participant characteristics highly variable across studies	Lack of direct test for LTBI requires extrapolation of test characteristics from active TB (Sn) and healthy, low-risk (Sp) populations
	Sp 4 (699)	Sp pooled 0.98 ^e (95% CI, 0.90 to 1.0)	Consistent and precise for Sp in low TB burden countries, imprecise in intermediate TB-burden country				QFT-G requires proper specimen handling prior to assay
	Observational studies of test accuracy					Studies vary with respect to how they report indeterminate results	This generation of QFT test is no longer being marketed
IGRA QFT-GIT Accuracy	Sn 24 (2,321)	Sn pooled 0.80 (95% CI, 0.77 to 0.84, $I^2=74.3\%$)	Consistent and precise for Sn	Undetected	Fair	Independent interpretation of test often not reported; description of participant characteristics highly variable across studies	Lack of direct test for LTBI requires extrapolation of test characteristics from active TB (Sn) and healthy, low-risk (Sp) populations
	Sp 4 (2,053)	Sp pooled 0.97 (95% CI, 0.94 to 0.99, $I^2=93.4\%$)	Consistent and precise for Sp in low TB burden countries, imprecise in intermediate TB-burden country				QFT-GIT requires proper specimen handling prior to assay
	Observational studies of test accuracy					Studies vary with respect to how they report indeterminate results	

Table 3. Summary of Evidence of Accuracy and Reliability of TST or IGRA Tests for Targeted^a Screening of Latent Tuberculosis Infection—KQ 2a

Test or Intervention	Number of Studies (observations) Study Designs By Test or Outcome	Summary of Findings By Test or Outcome	Consistency/Precision	Reporting Bias	Overall Quality	Body of Evidence Limitations	Applicability
IGRA QFT-GIT Reliability	Interrater reliability 1 (146)	Across all 4 tests (2 samples from each participant analyzed by manual and automated ELISA): 88.6% were concordant (16% concordant positive and 72.6% concordant negative); 11% were discordant. Discordance by method of interpretation: automated vs. automated= 4.8% (kappa 0.85); manual vs. manual= 6.9% (kappa 0.80); automated vs. manual=3.4% to 9.0% across comparisons (kappa 0.73 to 0.90)	Consistency unknown for single study assessing Interrater reliability, precision unknown	Undetected	Fair	Studies vary with respect to how they assess reliability outcomes	QFT-GIT requires proper specimen handling prior to assay
	Reproducibility 1 (130)	Number of discordant results in participants who had 2 samples drawn simultaneously: 10 /172 (5.8%)	Consistency unknown for single study assessing reproducibility, precision unknown				
	Test-retest reliability 2 (296)	1 study enrolling HCWs, 10/134 (7.5%) results changed from negative to positive and 5/15 (33.3%) changed from positive to negative at 2 weeks. In the other study enrolling Nepalese military recruits, kappa for agreement between initial test and retest: 0.48 (95% CI, 0.26 to 0.70)	Inconsistent and imprecise for test-retest reliability				

Table 3. Summary of Evidence of Accuracy and Reliability of TST or IGRA Tests for Targeted^a Screening of Latent Tuberculosis Infection—KQ 2a

Test or Intervention	Number of Studies (observations) Study Designs By Test or Outcome	Summary of Findings By Test or Outcome	Consistency/Precision	Reporting Bias	Overall Quality	Body of Evidence Limitations	Applicability
IGRA QFT-GIT Reliability (continued)	Interlaboratory reliability 1 (91) Observational studies of test accuracy	Across 3 labs, 7/ 91 (7.7%) subjects had discordant results; kappas of pairwise lab sample comparisons ranged from 0.87, 0.89, and 0.93	Consistency unknown for single study assessing interlaboratory reliability, precision unknown				

^a Targeted refers to screening of subjects who have been identified as higher risk for infection; for example, recent arrivals (within the past 5 years) to the United States who are foreign born.

^b Pooled estimate includes one study conducted in an intermediate TB-burden country with a much lower estimate (0.45) as compared with the low TB-burden countries included in the pooled estimate. The pooled estimate without this study was 0.98 (95% CI, 0.97 to 0.99, I^2 91.2%).

^c Pooled estimates includes two studies in intermediate TB-burden countries with much lower estimates (0.60 and 0.57) compared with low TB-burden countries included in the pooled estimate. The pooled estimate without these studies was the same, but I^2 was reduced to 88.7 percent.

^d Pooled estimate includes two low, two intermediate, and one mixed (two countries one low and one intermediate) TB-burden countries. The estimates in the two intermediate TB-burden countries were lower (0.85 and 0.73) compared with estimates in the other studies.

^e Pooled estimate is from maximum likelihood random effects model since two studies included point estimates for specificity of 1.0. The I^2 statistic is not calculated when using this model.

CI = confidence interval; FDA = Food and Drug Administration; LTBI = latent tuberculosis infection; mm = millimeter; PPD = purified protein derivative; QFT-G = QuantiFERON[®] TB Gold (2nd generation test); QFT-GIT = QuantiFERON[®] TB Gold-In-Tube (3rd generation test); Sn = sensitivity; Sp = specificity; TB = tuberculosis; T-SPOT.TB[®] = commercial ELISPOT assay; TST= tuberculin skin test.

Table 4. Summary of Evidence for Treatment of Latent Tuberculosis Infection With CDC-Recommended Pharmacotherapy Treatment Regimens—KQ 3

Test or Intervention	Number of Studies (observations) Study Designs By Test or Outcome	Summary of Findings By Test or Outcome	Consistency/Precision	Reporting Bias	Overall Quality	Body of Evidence Limitations	Applicability
INH vs. Placebo	1 RCT (27,830) ^a	<i>Developing active TB:</i> <i>Main analysis</i> RR: 0.35 at 5 years follow-up (95% CI, 0.24, 0.52) for INH x 24 weeks ^b as compared to placebo; NNT=112	Consistency NA for the single study; reasonably precise for developing active TB	Undetected	Good (Fair to good for sensitivity analysis)	Studies used in analysis used longer duration (1 year of INH) ^d and some used doses lower or higher than currently recommended; 1 trial was poor quality for high risk of selection, attrition, and measurement bias and confounding	Study population in main analysis trial include those with fibrotic pulmonary lesions and a 6-mm or greater Mantoux test; median age 50; trials in main and sensitivity analysis published over 30 ago (1963, 1965, 1968, 1978, 1982) Trials in sensitivity analysis enrolled HH contacts of active cases veterans with inactive pulmonary TB, people residing in mental institutions, and military members exposed to an active TB case
	1 RCT (27,830) ^a	<i>Deaths due to TB:</i> 0 vs. 3 RR: 0.14 (95% CI, 0.01, 2.78) for the combined isoniazid groups vs. placebo	Imprecise for deaths due to TB	Undetected	Good	Small number of events	Same as above for main analysis applicability
	1 RCT (27,830) ^a	<i>All-cause mortality:</i> NR by group	-	-	-	Data on all-cause mortality NR by group	Same as above for main analysis applicability
RIF vs. INH	1 RCT (847)	<i>Developing active TB:</i> NR <i>Deaths due to TB:</i> 0 vs. 0 <i>All-cause mortality:</i> 0 vs. 1	Consistency NA, single study; imprecise	Undetected	Good	Open label. Lacking data for outcome of developing active TB, no events for outcome of deaths due to TB, and only one event for all-cause mortality outcome	Adults with a positive TST and physician recommendation for INH in Canada, Saudi Arabia, and Brazil. Just over half were between ages 18 and 34 and just over half were male. Trial focused on harm outcomes; subjects only followed for duration of their treatment (4 or 9 months)

Table 4. Summary of Evidence for Treatment of Latent Tuberculosis Infection With CDC-Recommended Pharmacotherapy Treatment Regimens—KQ 3

Test or Intervention	Number of Studies (observations) Study Designs By Test or Outcome	Summary of Findings By Test or Outcome	Consistency/Precision	Reporting Bias	Overall Quality	Body of Evidence Limitations	Applicability
RPT + INH vs. INH	1 RCT (6,886) ^e	<p><i>Developing active TB:</i> 5 vs. 10^f</p> <p><i>Deaths due to TB:</i> NR</p> <p><i>All-cause mortality:</i> 30 vs. 34, p=0.42</p>	<p>Consistency NA for this single study; reasonably precise for developing active TB and all-cause mortality</p> <p>NR for deaths due to TB</p>	Undetected	Fair	Open label; single study, no data for deaths due to TB	Median age 37; just over half male; 57% white; combined intervention was directly observed once weekly x 3 months; high-risk subjects; most had a close contact with someone with active TB; 25% were included solely because of recent TST conversion

^a Of the 27,830 participants in the IUAT trial, the only trial meeting all eligibility criteria for KQ 3 that compared INH with placebo, 6,965 of them were treated with a CDC-approved regimen (INH 300 mg x 24 weeks). The IUAT trial randomized 27,830 participants to INH 300 mg x 12 weeks (6,956), INH 300 mg x 24 weeks (6,965), INH 300 mg x 52 weeks (6,919), or placebo (6,990).

^b The relative risks for the other treatment groups for developing active TB compared with placebo were 0.79 (95% CI, 0.58, 1.06) and 0.25 (95% CI, 0.16, 0.39), for 12 and 52 weeks of INH, respectively.

^c Followup for the 5 RCTs included in the sensitivity analysis ranged from 2 to 10 years; one study followed patients for 2 years (Bush), one for 5 years (IUAT), two for 7 years (Falk, Veening), and one for 10 years (Ferebee).

^d No longer a CDC-recommended treatment regimen.

^e This open-label, noninferiority trial randomized 7,731 people; we obtained data from CDC for this table on the subset of participants most directly relevant for this review: the 6,886 adults (18 years or older) who were HIV negative and were TST or IGRA positive.

^f The combination therapy group was found to be noninferior to the INH-only group.

CI = confidence interval; HH = household; INH = isoniazid; NA = not applicable; RCT = randomized controlled trial; RPT = rifapentine; RR = relative risk; TB = tuberculosis; TST= tuberculin skin test.

Table 5. Summary of Evidence for Harms Associated With CDC-Recommended Pharmacotherapy Treatment Regimens for Latent Tuberculosis Infection—KQ 5

Test or Intervention	Number of Studies (observations) Study Designs By Test or Outcome	Summary of Findings By Test or Outcome	Consistency/Precision	Reporting Bias	Overall Quality	Body of Evidence Limitations	Applicability
INH vs. placebo	1 RCT (27,830) ^a	<i>Hepatotoxicity:</i> <i>Main analysis:</i> RR: 4.59 at 5 years (95% CI, 2.03 to 10.39) for 24 weeks INH compared to placebo; NNH 279	Consistency NA for the single study in main analysis; consistent across 4 studies in sensitivity analysis; imprecise	Undetected	Fair for main analysis, fair to poor for sensitivity analyses	Harm ascertainment techniques not well described Studies used in sensitivity analysis limited by measurement and ascertainment bias	Study population in main analysis trial includes those with fibrotic pulmonary lesions and a ≥6-mm Mantoux test; median age 50; trial published in 1982 Trials in sensitivity analysis published in 1974, 1977, and 1978, and enrolled employees in a U.S. hospital, individuals meeting ATS criteria referred to a U.S. military medical center, and veterans with inactive pulmonary TB
	Sensitivity analysis with 4 RCTs (35,161)	<i>Sensitivity analysis:</i> Pooled RR: 5.04 ^b (95% CI, 2.50 to 10.15, I ² =0%)					
	1 RCT (27,830) ^a	<i>Death from hepatotoxicity^c:</i> 0 deaths in placebo group, 0.14 deaths per 1,000 receiving INH; RR: 2.35 (95% CI, 0.12 to 45.46, NNH = 6,947)	Consistency NA for the single study; imprecise	Undetected	Fair	Rare number of events. Harm ascertainment techniques not well described	Same as above for main analysis
	1 RCT (27,830) ^a	<i>Discontinuation of treatment due to adverse events:</i> <i>Main analysis:</i> RR 1.50 ^b (95% CI, 1.18 to 1.89, NNH 167) <i>Sensitivity analysis:</i> Pooled RR: 1.58 (95% CI, 1.00 to 2.49, I ² = 70.2%)	Consistency NA for the single study; reasonably precise. Inconsistent across the 4 studies included in sensitivity analysis, reasonably precise	Undetected	Fair for main analysis Fair to poor for sensitivity analysis	Harm ascertainment techniques not well described Studies limited by lack of prespecification of harm outcomes, measurement and ascertainment bias	Same as above for main analysis Trials in treatment discontinuation sensitivity analysis published in 1963, 1965, and 1977, and enrolled residents of mental institutions, HH contacts of active cases, and adults meeting ATS criteria for chemoprophylaxis
	1 RCT (27,830) ^a	<i>Gastrointestinal adverse events:</i> RR 1.33 ^b (95% CI, 1.01 to 1.75) <i>Sensitivity analysis:</i> Different outcomes reported across studies; no differences among groups	Consistency NA for the single study; reasonably precise	Undetected	Fair to poor	Gastrointestinal harms not prespecified, measurement and ascertainment bias	Study population in main analysis trial includes those with fibrotic pulmonary lesions and a 6-mm or greater Mantoux test; median age 50; trial published in 1982

Table 5. Summary of Evidence for Harms Associated With CDC-Recommended Pharmacotherapy Treatment Regimens for Latent Tuberculosis Infection—KQ 5

Test or Intervention	Number of Studies (observations) Study Designs By Test or Outcome	Summary of Findings By Test or Outcome	Consistency/Precision	Reporting Bias	Overall Quality	Body of Evidence Limitations	Applicability
INH vs. RIF	3 RCTs (1,327)	<i>Hepatotoxicity:</i> Pooled RR: 3.29 (95% CI, 1.17 to 6.28, $I^2 = 0\%$) <i>Death from hepatotoxicity:</i> No events reported in any arms of any study	Consistent, imprecise	Undetected	Fair to Good	2 trials were open-label, 1 trial lost nearly half of participants to follow-up	Trials published in 2004, 2008, 2012; subjects had positive TST following Canadian guidelines in 2 trials, subjects in other trial were inmates diagnosed with LTBI at jail entry
		<i>Discontinuation of treatment due to adverse events:</i> Pooled RR: 1.61 (95% CI, 0.57 to 4.57, $I^2 = 40.0\%$)	Inconsistent, imprecise	Undetected	Fair to Good	Same as above	Same as above
	2 RCTs (1,211)	<i>Gastrointestinal adverse events:</i> Pooled RR: 1.60 (95% CI, 0.76 to 3.40, $I^2 = 0\%$)	Inconsistent and imprecise	Undetected	Fair	1 study lost nearly half of participants to follow-up; duration of follow-up may be inadequate for harms	Trials published in 2008 and 2012; subjects had positive TST following Canadian guidelines in 1 trial, subjects in other trial were inmates diagnosed with LTBI at jail entry
	2 RCTs (480)	<i>Other adverse events:</i> Pooled RR: 0.82 (95% CI, 0.42 to 1.59, $I^2 = 0\%$)	Inconsistent and imprecise	Undetected	Fair	1 study lost nearly half of participants to follow-up; duration of follow-up may be inadequate for harms, measurement and ascertainment bias	Trials published in 2004 and 2012; subjects had positive TST following Canadian guidelines in 1 trial, subjects in other trial were inmates diagnosed with LTBI at jail entry
RPT + INH vs. INH	1 RCT (6,886) ^d	<i>Hepatotoxicity:</i> (Grade 3 or 4) Calculated RR: 0.90 (95% CI, 0.75 to 1.08) <i>Death from hepatotoxicity:</i> 0.83 (95% CI, 0.51 to 1.35)	Consistency NA for the single study, imprecise	Undetected	Fair	Single study, masking unclear and high overall attrition	Trial published in 2011, data were from HIV-negative subgroup with TST or IGRA confirmation; combined intervention was directly observed once week x 3 months; high-risk individuals; most had close contact with someone with active TB; 25% were included solely because of recent TST conversion
		<i>Discontinuation of treatment due to adverse events:</i> Calculated RR: 1.28 (95% CI, 1.03 to 1.59)	Consistency NA for the single study, reasonably precise	Undetected	Fair	Single study, masking unclear and high overall attrition	Same as above
		<i>Possible hypersensitivity:</i> RR: 8.04 (95% CI, 4.88 to 13.26)	Consistency NA for the single study, imprecise	Undetected	Fair	Single study, masking unclear and high overall attrition	Same as above

Table 5. Summary of Evidence for Harms Associated With CDC-Recommended Pharmacotherapy Treatment Regimens for Latent Tuberculosis Infection—KQ 5

^a Of the 27,830 participants in the IUAT trial, the only trial meeting all eligibility criteria for KQ 3 that compared INH with placebo, 6,965 of them were treated with a CDC-approved regimen (INH 300 mg x 24 weeks). The IUAT trial randomized 27,830 participants to INH 300 mg x 12 weeks (6,956), INH 300 mg x 24 weeks (6,965), INH 300 mg x 52 weeks (6,919), or placebo (6,990).

^b Estimate includes combined data from all three INH study arms (12 weeks, 24 weeks, 52 weeks) in the IUAT trial.

^c One additional RCT used in sensitivity analysis for this outcome reported no deaths from hepatotoxicity in either the INH or placebo arm.

^d Followup for the 5 RCTs included in the sensitivity analysis ranged from 2 to 10 years; one study followed patients for 2 years (Bush), one for 5 years (IUAT), two for 7 years (Falk, Veening), and one for 10 years (Ferebee).

ATS = American Thoracic Society; CI = confidence interval; IGRA = interferon gamma release assay; INH = isoniazid; LTBI = latent tuberculosis infection; NA = not applicable; NNH = number needed to harm; RCT = randomized controlled trial; RIF = rifampin; RPT = rifapentine; RR = relative risk; TB = tuberculosis; TST = tuberculin skin test; U.S. = United States.

Table 6. Projected 5-Year Outcomes of Screening 100,000 Asymptomatic Adults for Latent Tuberculosis Infection^a

	Variable	Foreign-born Population Screened	Foreign-born Population Not Screened	Community, noninstitutionalized Population Screened	Community, noninstitutionalized Population Not Screened
Detection	Patients with LTBI ^b n (95% CI)	20,500 (16,100 to 25,800)	20,500 (16,100 to 25,800)	4,700 (3,400 to 6,300)	4,700 (3,400 to 6,300)
	Positive screening test, chest x-ray, offered LTBI treatment ^c n,(n false positive/n true positive)	18,580 (2,385/16,195)	NA	6,572 (2,859/3,713)	NA
Benefits	Progression to active TB ^d , n,(n false positive/n true positive) [Range of estimate] ^e	79 (0/79) [62 to 99]	225 (NA) [177 to 287]	18 (0/18) [13 to 24]	52 (NA) [37 to 69]
	NNT to prevent 1 case of LTBI from progressing to active TB ^f	111	-	111	-
	Progression to active TB using alternative assumption for rate of LTBI reactivation ^g . n,(n false positive/n true positive) [Range of estimate] ^e	28 (0/28) [19 to 35]	79 [55 to 100]	5 (0/5) [4 to 7]	16 (NA) [11 to 21]
	NNT to prevent 1 case of LTBI from progressing to active TB under alternative assumption for rate of LTBI reactivation ^{f,g}	314	-	366	-
	TB transmission	Unknown	Unknown	Unknown	Unknown
	Death from TB	Unknown	Unknown	Unknown	Unknown
Harms	Hepatotoxicity ^h (INH or placebo) n,(n false positive/n true positive) [Range of estimate] ^e	85 (11/74) [70 to 104]	19 (NA) [15 to 23]	30(13/17) [26 to 36]	7 (NA) [6 to 8]
	NNH to cause hepatotoxicity from treatment with INH ⁱ	279	-	279	-
	Hepatotoxicity (RIF or placebo) ^j n,(n false positive/n true positive) [Range of estimate] ^e	26 (3/23) [21 to 32]	19 (NA) [15 to 23]	9 (4/5) [8 to 11]	7 (NA) [6 to 8]
	NNH to cause hepatotoxicity from treatment with RIF ^j	2,531	-	2,531	-
	Discontinuation due to adverse effects (INH or placebo) ^k n,(n false positive/n true positive) [Range of estimate] ^e	334 (43/292) [274 to 407]	223 (NA) [183 to 271]	118 (51/67) [101 to 140]	79 (NA) [67 to 93]
	NNH to cause discontinuation due to adverse events from treatment with INH	167	-	167	-
	Potential psychological harms	Unknown		Unknown	

Table 6. Projected 5-Year Outcomes of Screening 100,000 Asymptomatic Adults for Latent Tuberculosis Infection^a

Variable	Foreign-born Population Screened	Foreign-born Population Not Screened	Community, noninstitutionalized Population Screened	Community, noninstitutionalized Population Not Screened
Summary of estimated benefits and harms	<p>52 to 146 active TB cases prevented^l</p> <p>67 cases of hepatotoxicity caused if using INH for everyone; 9 of those cases caused by unnecessary treatment (for people with false positives)^m</p> <p>7 cases of hepatotoxicity caused if using RIF for everyone; 1 of those cases caused by unnecessary treatment (for people with false positives)^m</p> <p>111 cases of discontinuation due to adverse events caused if using INH for everyone; 14 of those caused by unnecessary treatment (for people with false positives)^m</p>		<p>10 to 33 active TB cases prevented^l</p> <p>24 cases of hepatotoxicity caused if using INH for everyone; 10 of those cases caused by unnecessary treatment (for people with false positives)^m</p> <p>3 cases of hepatotoxicity caused if using RIF for everyone; 1 of those cases caused by unnecessary treatment (for people with false positives)^m</p> <p>39 cases of discontinuation due to adverse events caused if using INH for everyone; 17 of those caused by unnecessary treatment (for people with false positives)^m</p>	

^a Projected benefits and harms were determined for people in whom screening had not previously been performed and who would be eligible for and offered treatment for LTBI based on a positive screening test. When relevant, projected outcomes are shown as overall, and in parentheses for people with false positive tests and those with true positive tests, to illustrate how many people would undergo unnecessary intervention with resulting harm.

^b The prevalence of LTBI is 4.7 percent and 20.5 percent for the U.S. overall population and foreign-born U.S. population, respectively, based on 2011-2012 NHANES.²⁰ We use the foreign-born U.S. population as an example of outcomes among a higher risk population because available estimates of prevalence and progression are readily available for this population, unlike other high-risk populations.

^c Based on sensitivity (0.79) and specificity(0.97) for TST with 10-mm threshold for positive test, which is the threshold recommended for recent arrivals (within past 5 years) to the United States from high-risk areas based on current CDC recommendations.¹² A small proportion of those x-rayed will have findings suggestive of active TB disease and will go on to receive further diagnostic evaluation and treatment. A precise estimate of this proportion is not available and we have assumed it to be zero.

^d Estimates for benefits were based on the IUAT trial, which may have limited applicability to current clinical practice because the study population was composed of subjects with pulmonary fibrotic lesions. Rate of progression in the absence of treatment at 5 years was 1.39 percent in the placebo arm and 0.5 percent in the 24-week isoniazid treatment arm, for a relative risk reduction of 0.35. Patients with false positive or false negative screening results receive no benefit from treatment, thus progression to active TB is only relevant for true positives.

Table 6. Projected 5-Year Outcomes of Screening 100,000 Asymptomatic Adults for Latent Tuberculosis Infection^a

^cThis range is an estimate based on the lower and upper bounds of the 95% CI, for prevalence. That is, it provides the range of possible estimates given the precision of the LTBI prevalence estimates available.

^f Number needed to treat is calculated as 1/absolute risk reduction between treatment and control groups.

^g Because using the rate of progression to active TB from the IUAT may not reflect contemporary risk, we also used a more recent estimate, based on the rate of nonclustered TB cases, a proxy for reactivation. The rate of progression is estimated at 0.084/100 person-years for the overall U.S. population, and 0.098/100 person-years for the foreign-born U.S. population.⁸ We assumed the same relative risk reduction for treatment with isoniazid from the IUAT trial (0.35).

^h We used rate of hepatotoxicity from the IUAT trial: 0.1 percent in the placebo group and 0.46 percent in the 24-week INH treatment group.¹³⁵

ⁱ Number needed to harm is calculated as 1/absolute harm risk difference between treatment and control groups.

^j Estimates of hepatotoxicity in the studies evaluating isoniazid versus rifampin reported hepatotoxicity rates over a different time period, using different definitions, were conducted nearly 30 years after the IUAT trial, and the event rate in the INH treatment arm were much higher than the rates reported in IUAT likely because these studies were designed specifically to evaluate harms. For these reasons, these rates could not be directly used in this outcomes table to compare to the NNT calculated from the IUAT trial. Thus, we used the pooled RR for hepatotoxicity for INH versus RIF of 3.29 from our meta-analysis to adjust the IUAT trial rate for hepatotoxicity among the INH-treated arm to obtain an indirect estimate of a 5-year risk of hepatotoxicity from RIF as compared with placebo of 0.14 percent.

^k We used estimates of discontinuation of treatment due to adverse events from the IUAT.¹³⁵

^l This range includes estimate based on rates of LTBI progression in the absence of treatment from the IUAT trial and an estimate based on a more recent lower estimate of the rate of progression in the absence of treatment.

^m Unnecessary treatment is determined by the number of persons for whom a false positive test resulted in treatment that is unnecessary and for which they have no potential to benefit.

CI = confidence interval; INH = isoniazid; LTBI = latent tuberculosis infection; NA = not applicable; NNH = number needed to harm; NNT = number needed to treat; RIF = rifampin; TB = tuberculosis.

Appendix A Table 1. Prevalence of LTBI by High-Risk Category From Published Studies in English, French, or Spanish, 2009 Through 2014*

High-risk Description	Prevalence Based on TST \geq 5 mm Median (Range)	Prevalence Based on T-SPOT.TB Median (Range)	Prevalence Based on QFT-GIT Median (Range)	Incidence of Active TB Median Rate per 1,000 (Range)
High risk because of increased likelihood of TB exposure				
Prisoners	45.5 (23.1–87.6)	NR	NR	2.6 (0.03–9.8)
Health care workers	29.5 (1.4–97.6)	5.2 (3.5–28.7)	14.1 (0.9–76.7)	1.3 (0.4–4.1)
Adult contacts of active TB cases	26.3 (1.8–82.7)	48.0 (29.6–59.6)	21.1 (6.6–55.1)	0.6 ^a
Immigrants from high TB burden countries	39.7 (17.8–55.4)	17.0 (9.0–24.9)	30.2 (9.8–53.8)	3.6 (1.3–41.2)
Illicit drug-users	85.0 (0.3–86.7)	45.8 (34.1–57.5)	63.0 (1.4–66.4)	6.0 ^a
Homeless persons	45.6 (20.5–79.8)	NR	53.8 (18.6–75.9)	2.2 (0.1–4.3)
High risk because of underlying medical conditions				
HIV infection	19.2 (2.1–54.8)	11.3 (4.3–67.6)	14.5 (2.7–21.5)	16.2 (12.4–28.0)
Use of TNF alpha blockers	18.6 (11.3–68.2)	20.0 (12.9–25.0)	11.8 (4.0–22.3)	1.4 ^a
Silicosis	NR	61.0 ^a	46.6 ^a	32.1 ^a
Organ transplantation	7.7 (4.4–21.9)	29.5 (20.5–38.5)	21.9 (16.4–23.5)	5.1 ^a
Hemodialysis	21.9 (2.6–42.1)	43.6 (23.3–58.2)	33.4 (17.4–44.2)	26.6 (1.3–52.0)

*Adapted from Getahun et al., 2015⁹

^aSingle study

HIV = human immunodeficiency virus; LTBI = latent TB infection; mm = milligram; NR = not reported; QFT-GIT = QuantiFERON TB Gold-In-Tube[®] test (3rd generation test); TB = tuberculosis; TNF = tumor necrosis factor; TST = tuberculin skin test

Appendix A Table 2. Recommended LTBI Treatment Regimens^a

Drug(s)	Duration	Dose	Frequency	Total Doses
INH	9 months	5 mg/kg Maximum dose: 300 mg	Daily	270
		15 mg/kg Maximum dose: 900 mg	Twice weekly ^a	76
	6 months	5 mg/kg Maximum dose: 300 mg	Daily	180
		15 mg/kg Maximum dose: 900 mg	Twice weekly ^a	52
INH and RPT	3 months	INH: 15 mg/kg rounded up to the nearest 50 or 100 mg; 900 mg maximum RPT: 10.0–14.0 kg 300 mg 14.1–25.0 kg 450 mg 25.1–32.0 kg 600 mg 32.1–49.9 kg 750 mg ≥50.0 kg, 900 mg maximum	Once weekly ^a	12
RIF	4 months	10 mg/kg Maximum dose: 600 mg	Daily	120

^a Intermittent regimens must be provided via directly observed therapy (DOT) (i.e., health care worker observes the ingestion of medication).

INH = isoniazid; LTBI = latent TB infection; mg/kg = milligram/kilogram; RIF = rifampin; RPT = rifapentine

Appendix B. Detailed Methods

Search Strategies

Initial searches

PubMed (08/29/14)

Tuberculosis final searches, 8-29-14 – 4,288 citations saved in EndNote PubMed:
3,531 total English language citations saved in EndNote

Search Query	Items found
#1 Search ("Tuberculosis"[Mesh] OR "Latent Tuberculosis"[Mesh])	159183
#2 Search ("Interferon-gamma Release Tests"[Mesh] OR IGRA[All Fields] OR "Mantoux tuberculin skin test"[All Fields] OR "Tuberculin Test"[Mesh] OR "tuberculin skin test"[All Fields] OR TST[tiab] OR "T-SPOT"[All Fields] OR "T-SPOT.TB"[All Fields] OR QuantiFERON[All Fields] OR "QFT-GIT"[All Fields])	15573
#3 Search (#1 and #2)	9057
#4 Search (#1 and #2) Filters: Humans	7781
#5 Search (#1 and #2) Filters: Systematic Reviews; Humans	151
#6 Search ((randomized[title/abstract] AND controlled[title/abstract] AND trial[title/abstract]) OR (controlled[title/abstract] AND trial[title/abstract]) OR "controlled clinical trial"[publication type] OR "Randomized Controlled Trial"[Publication Type] OR "Single-Blind Method"[MeSH] OR "Double-Blind Method"[MeSH] OR "Random Allocation"[MeSH])	576514
#7 Search (#4 and #6)	176
#8 Search ("Cohort Studies"[MeSH] OR "Epidemiologic Studies"[MeSH] OR "Cross-Sectional Studies"[MeSH] OR "Follow-Up Studies"[MeSH] OR "Seroepidemiologic Studies"[MeSH] OR "Evaluation Studies"[Publication Type] OR "observational study" OR "observational studies")	1820788
#9 Search (#4 and #8)	1800
#10 Search ("Isoniazid"[Mesh] OR isoniazid[All Fields] OR "rifapentine"[Supplementary Concept] OR rifapentine[All Fields] OR "Rifampin"[Mesh] OR Rifampin[All Fields])	32885
#11 Search (#1 and #10)	12750
#12 Search (#1 and #10) Filters: Humans	10205
#13 Search (#1 and #10) Filters: Systematic Reviews; Humans	155
#14 Search (#12 and #6)	684
#15 Search ("Case-Control Studies"[MeSH] OR "Cohort Studies"[MeSH] OR "Epidemiologic Studies"[MeSH] OR "Follow-Up Studies"[MeSH] OR "Seroepidemiologic Studies"[MeSH] OR "Evaluation Studies"[Publication Type] OR "observational study" OR "observational studies")	1820788
#16 Search (#12 and #15)	1811
#17 Search (#5 or #13)	282
#18 Search (#5 or #13) Filters: English	243
#19 Search (#17 not #18)	39
#20 Search (#7 or #9 or #14 or #16)	4013
#21 Search (#7 or #9 or #14 or #16) Filters: English	3348
#22 Search (#20 not #21)	665
#23 Search (#18 or #21)	3531
#24 Search (#19 or #22)	703

Appendix B. Detailed Methods

Cochrane Library (08/29/14)

For the Cochrane Library search, we did not limit using study design terms, because Cochrane breaks out study results by the categories we seek. We searched the Cochrane Library of Reviews, Trials, Methods, and Technology Assessments. We did not save the Economic Evaluations.

Results

Screening:

All = 250 total results (179 without Economic Evaluations)

Cochrane Reviews = 12, all imported

Other reviews = 27, all imported

Trials = 136

Technology Assessments = 4

Economic Evaluations = 71 (not saved)

Drug therapy:

All = 714 total results (638 without Economic Evaluations)

Cochrane Reviews = 22, 11 imported

Other reviews = 36, 34 imported

Trials = 580, 533 imported

Economic Evaluations = 76 (not saved)

Importing to EndNote = 84 total reviews

673 Trials and Technology Assessments

Total in EndNote from the Cochrane Library = 757

ClinicalTrials.gov search 4-27-15

291 trials

Tuberculosis AND ("Interferon-gamma Release Tests" OR IGRA OR "Mantoux tuberculin skin test" OR "Tuberculin Test" OR "tuberculin skin test" OR TST or "T-SPOT" OR "T-SPOT.TB" or QuantiFERON or "QFT-GIT" OR isoniazid OR rifapentine OR Rifampin)

WHO ICTRP search and results (05/19/15)

185 records for 173 trials

Searched in Advanced search:

Condition box: Tuberculosis

Intervention box: Interferon-gamma Release Tests OR IGRA OR Mantoux tuberculin skin test OR Tuberculin Test OR tuberculin skin test OR TST or T-SPOT OR T-SPOT.TB or QuantiFERON or QFT-GIT OR isoniazid OR rifapentine OR Rifampin

(Recruitment status ALL)

185 records for 173 trials found

Appendix B. Detailed Methods

Bridge Searches

Tuberculosis update searches, July 30-31, 2015 and Aug 3, 2015

PubMed (07/30/15)

Search	Query	Items found
#1	Search ("Tuberculosis"[Mesh] OR "Latent Tuberculosis"[Mesh])	162959
#2	Search ("Interferon-gamma Release Tests"[Mesh] OR IGRA[All Fields] OR "Mantoux tuberculin skin test"[All Fields] OR "Tuberculin Test"[Mesh] OR "tuberculin skin test"[All Fields] OR TST[tiab] OR "T-SPOT"[All Fields] OR "T-SPOT.TB"[All Fields] OR QuantiFERON[All Fields] OR "QFT-GIT"[All Fields])	16219
#3	Search (#1 and #2)	9413
#4	Search (#1 and #2) Filters: Humans	8116
#5	Search (#1 and #2) Filters: Systematic Reviews; Humans	163
#6	Search ((randomized[title/abstract] AND controlled[title/abstract] AND trial[title/abstract]) OR (controlled[title/abstract] AND trial[title/abstract]) OR "controlled clinical trial"[publication type] OR "Randomized Controlled Trial"[Publication Type] OR "Single-Blind Method"[MeSH] OR "Double-Blind Method"[MeSH] OR "Random Allocation"[MeSH])	608085
#7	Search (#4 and #6)	184
#8	Search ("Cohort Studies"[MeSH] OR "Epidemiologic Studies"[MeSH] OR "Cross-Sectional Studies"[MeSH] OR "Follow-Up Studies"[MeSH] OR "Seroepidemiologic Studies"[MeSH] OR "Evaluation Studies"[Publication Type] OR "observational study" OR "observational studies")	1946601
#9	Search (#4 and #8)	1945
#10	Search ("Isoniazid"[Mesh] OR isoniazid[All Fields] OR "rifapentine"[Supplementary Concept] OR rifapentine[All Fields] OR "Rifampin"[Mesh] OR Rifampin[All Fields])	33825
#11	Search (#1 and #10)	13136
#12	Search (#1 and #10) Filters: Humans	10571
#13	Search (#1 and #10) Filters: Systematic Reviews; Humans	170
#14	Search (#12 and #6)	708
#15	Search ("Case-Control Studies"[MeSH] OR "Cohort Studies"[MeSH] OR "Epidemiologic Studies"[MeSH] OR "Follow-Up Studies"[MeSH] OR "Seroepidemiologic Studies"[MeSH] OR "Evaluation Studies"[Publication Type] OR "observational study" OR "observational studies")	1946601
#16	Search (#12 and #15)	1931
#17	Search (#5 or #13)	309
#18	Search (#5 or #13) Filters: English	270
#19	Search (#17 not #18)	39
#20	Search (#7 or #9 or #14 or #16)	4285
#21	Search (#7 or #9 or #14 or #16) Filters: English	3604
#22	Search (#20 not #21)	681
#23	Search (#18 or #21)	3813
#24	Search (#19 or #22)	719
#25	Search (child* OR children OR teen OR teens OR teenage OR teenaged OR adolescen* OR pediatric OR paediatric* OR boys OR girls OR youth OR youths)	3221593
#26	Search (#5 or #13) Filters: English; Child: birth-18 years	67
#27	Search (#18 and #25)	84
#28	Search (#27 or #26)	84
#29	Search (#5 or #13) Filters: English; Adult: 19+ years	65
#30	Search (#28 and #29)	39
#31	Search (#28 NOT #30)	45
#32	Search (#18 NOT #31)	225
#33	Search (#28 NOT #30) Filters: Publication date from 2014/03/29	4

Appendix B. Detailed Methods

Search	Query	Items found
#34	Search (#18 NOT #31) Filters: Publication date from 2014/03/29	11
#35	Search (#19 and #25)	12
#36	Search (#17 not #18) Filters: Child: birth-18 years	10
#37	Search (#35 or #36)	12
#38	Search (#17 not #18) Filters: Adult: 19+ years	10
#39	Search (#37 and #38)	8
#40	Search (#37 NOT #39)	4
#41	Search (#37 NOT #40)	8
#42	Search (#7 or #9 or #14 or #16) Filters: Publication date from 2014/03/29; English	200
#43	Search (#7 or #9 or #14 or #16) Filters: Publication date from 2014/03/29; English; Child: birth-18 years	84
#44	Search (#42 and #25)	85
#45	Search (#43 or #44)	87
#46	Search (#7 or #9 or #14 or #16) Filters: Publication date from 2014/03/29; English; Adult: 19+ years	143
#47	Search (#45 and #46)	58
#48	Search (#45 NOT #47)	29
#49	Search (#46 NOT #48)	143
#50	Search (#19 or #22) Filters: Publication date from 2014/03/29	12
#51	Search (#19 or #22) Filters: Publication date from 2014/03/29; Child: birth-18 years	4
#52	Search (#50 and #25)	4
#53	Search (#19 or #22) Filters: Publication date from 2014/03/29; Adult: 19+ years	6

Cochrane Library (08/03/15)

Adults+ = 10

4 Cochrane Reviews

6 Trials

Children = 21

8 Reviews

3 Cochrane Reviews

5 Other reviews

8 Trials

1 Technology Assessment

4 Econ Evaluations (not saved)

ID	Search	Hits
#1	[mh Tuberculosis] or [mh "Latent Tuberculosis"]	1766
#2	[mh "Interferon-gamma Release Tests"] or IGRA or "Mantoux tuberculin skin test" or [mh "Tuberculin Test"] or "tuberculin skin test" or TST or "T-SPOT" or "T-SPOT.TB" or QuantiFERON or "QFT-GIT"	699
#3	#1 and #2	260
#4	[mh Isoniazid] or isoniazid or rifapentine or [mh Rifampin] or Rifampin	1918
#5	#1 and #4	740
#6	#3 or #5 Publication Year from 2014 to 2015	31
#7	(child* or children or teen or teens or teenage or teenaged or adolescen* or pediatric or paediatric* or boys or girls or youth or youths)	172203
#8	#6 and #7	16
#9	#6 and (adult* or middle-age* or elderly)	19
#10	#9 and #8, Adults+	10
#11	#6 not #10, Children	21

Appendix B. Detailed Methods

WHO ICTRP search and results

(6/9/15 – 8/3/15)

0 results

Searched in Advanced search:

Condition box: Tuberculosis

Intervention box:

Interferon-gamma Release Tests OR IGRA OR Mantoux tuberculin skin test OR Tuberculin Test OR tuberculin skin test OR TST or T-SPOT OR T-SPOT.TB or QuantiFERON or QFT-GIT OR isoniazid OR rifapentine OR Rifampin

(Recruitment status ALL)

ClinicalTrials.gov search update (08/03/15)

Last Updated Date: 4/27/15-08/03/15

39 trials

11 child

28 adult

Tuberculosis AND ("Interferon-gamma Release Tests" OR IGRA OR "Mantoux tuberculin skin test" OR "Tuberculin Test" OR "tuberculin skin test" OR TST or "T-SPOT" OR "T-SPOT.TB" or QuantiFERON or "QFT-GIT" OR isoniazid OR rifapentine OR Rifampin)

Appendix B. Eligibility Criteria for Studies by Key Question

Key Question	Population	Intervention and Comparator	Setting	Outcomes	Study Design
KQ 1: Effect of screening for LTBI on morbidity, mortality, quality of life, and transmission	Asymptomatic adults belonging to populations at increased risk for LTBI ^a The following are excluded: children, symptomatic adults, close contacts of active TB patients, and populations at highest risk for progression from LTBI to active TB disease because of underlying immunosuppression or for whom LTBI screening and treatment would be part of standard disease management by specialty care providers. This includes people with HIV, head and neck cancer, leukemia or lymphoma, silicosis, history of or planned organ transplant, dialysis, planned or active use of TNF- α inhibitors, and planned or active use of chemotherapy. Mixed populations can be included if results are stratified for the included portion of the study population or the excluded portion does not exceed 25% of the study population.	Screening with TST or IGRA as compared with no screening. Studies with no comparator group are excluded.	Primary care settings in countries categorized as “Very High” on the Human Development Index (as defined by the United Nations Development Programme). Study settings considered to be applicable to primary care will also include homeless shelters, correctional facilities, college health settings, long-term care facilities, public health clinics, and workplaces. HIV and subspecialty care settings and workplace settings that screen for LTBI as part of a formal surveillance program for occupational exposure are excluded.	Active TB disease, reduction in transmission, improved quality of life, and reduction in mortality (disease-specific and overall). Other outcomes are excluded.	RCTs, prospective cohort studies. Other study designs are excluded.
KQ 2a: Accuracy and reliability of TST and IGRA screening tests KQ 2b: Accuracy and reliability of sequential screening strategies using TST	For sensitivity outcome: Patients with bacteriologically confirmed active TB who have not yet received treatment or who had received no more than a few weeks of treatment. Subjects with TB infection not confirmed by culture, AFB smear, or molecular tests are excluded. For specificity outcome: Healthy subjects with no history of TB exposure or risks. Subjects with known history of TB or TB exposure, subjects with HIV, and acutely ill subjects are excluded.	TST using Mantoux method with intermediate strength dose of PPD (i.e., 5 TU PPD-S, 2.5 TU RT-23) and standard thresholds for positive test (i.e., 5-mm, 10-mm, 15-mm). Commercially available, FDA-approved IGRA tests. T-SPOT.TB®; QuantiFERON® TB Gold (2nd generation); and QuantiFERON® TB Gold-	For sensitivity outcome: Studies in any country in any setting are included. For specificity outcome: Studies in intermediate or low TB-burden countries are included. Studies in high TB-burden countries are excluded. ^b	Sensitivity, specificity, and reliability (i.e., test-retest). Concordance rates among tests and other outcomes are excluded. Studies assessing two-step TST testing were excluded.	Systematic reviews, RCTs, cohort studies, cross-sectional studies. Other study designs are excluded.

Appendix B. Eligibility Criteria for Studies by Key Question

Key Question	Population	Intervention and Comparator	Setting	Outcomes	Study Design
and IGRA screening tests	Mixed populations of children and adults or studies with both HIV-negative and HIV-positive subjects (sensitivity outcome only) can be included if results are stratified for the includable portion of the study population or the excluded portion does not exceed 25% of the study population.	In-Tube (3rd generation). Other tests, such as nucleic acid amplification, are excluded.			
KQ 3: Effectiveness of treatment for LTBI	Asymptomatic adults with confirmed LTBI; otherwise, same criteria as for KQ 1 except that close contacts of active TB patients were eligible if LTBI was confirmed (e.g., with a positive TST).	Treatment with CDC-recommended regimen (isoniazid daily for 6 or 9 months, isoniazid twice weekly by directly observed therapy for 6 or 9 months, rifampin daily for 4 months, or isoniazid plus rifapentine weekly by directly observed therapy for 3 months) as compared to no treatment, delayed treatment, or another eligible treatment. Studies comparing other treatments or combinations are excluded.	Same as KQ 1, except that workplace settings were eligible.	Active TB disease (i.e., progression to active TB disease), reduction in transmission, improved quality of life, and reduction in mortality (disease-specific and overall).	RCTs
KQ 4: Harms of screening for LTBI	Same as KQ 1.	TST and IGRA tests as described in KQ 2.	Same as KQ 1	False-positive results leading to unnecessary testing or treatment, labeling, stigma, anxiety, and cellulitis	Systematic reviews; RCTs and prospective cohort studies.
KQ 5: Harms of treatment for LTBI	Same as KQ 3.	Same as KQ 3.	Same as KQ 1, except that workplace settings were eligible	Hepatotoxicity, mortality from hepatotoxicity, nausea, vomiting, peripheral neuropathy, development of	RCTs, prospective cohort studies, and case-control studies.

Appendix B. Eligibility Criteria for Studies by Key Question

Key Question	Population	Intervention and Comparator	Setting	Outcomes	Study Design
				drug-resistant TB, and other specific adverse effects of medications.	

^a Adult population subgroups at increased risk for developing active TB include (1) people who have immigrated from TB-endemic countries, (2) people who work or reside in facilities or institutions with high-risk individuals, such as homeless shelters, correctional facilities, nursing homes or residential facilities, and (3) people with increased risk for progression from LTBI to active TB due to underlying illness or use of medications, injection drug use, or radiographic evidence of prior healed TB.¹

^b High TB-burden countries include the following: Afghanistan, Bangladesh, Brazil, Cambodia, China, the Democratic Republic of the Congo, Ethiopia, India, Indonesia, Kenya, Mozambique, Myanmar, Nigeria, Pakistan, the Philippines, the Russian Federation, South Africa, Thailand, Uganda, the United Republic of Tanzania, Vietnam and Zimbabwe. This list is not exhaustive but represents the countries with the highest absolute burden (high rates and high population).³⁰

AFB = acid fast bacilli; CDC = Centers for Disease Control and Prevention; FDA = Food and Drug Administration; HIV = human immunodeficiency virus; IGRA = interferon gamma release assay; KQ = key question; LTBI = latent tuberculosis infection; mm = milligram; PPD = purified protein derivative; RCT = randomized, controlled trial; TB = tuberculosis; TNF- α = tumor necrosis factor- α ; TST = tuberculin skin test; TU = tuberculin units.

Appendix B. U.S. Preventive Services Task Force Quality Rating Criteria

Randomized Controlled Trials

Criteria

Initial assembly of comparable groups: Randomized controlled trials (RCTs)—adequate randomization, including concealment and whether potential confounders were distributed equally among groups; cohort studies—consideration of potential confounders with either restriction or measurement for adjustment in the analysis; consideration of inception cohorts

Maintenance of comparable groups (includes attrition, crossovers, adherence, and contamination)

Important differential loss to followup or overall high loss to followup

Measurements: Equal, reliable, and valid (includes masking of outcome assessment)

Clear definition of interventions

Important outcomes considered

Analysis: Adjustment for potential confounders for cohort studies or intention-to-treat analysis for RCTs; for cluster RCTs, correction for correlation coefficient

Definition of Ratings Based on Above Criteria

- Good: • Meets all criteria: Comparable groups are assembled initially and maintained throughout the study (followup $\geq 80\%$); reliable and valid measurement instruments are used and applied equally to the groups; interventions are spelled out clearly; important outcomes are considered; and appropriate attention is given to confounders in analysis.
- Fair: • Studies will be graded “fair” if any or all of the following problems occur, without the important limitations noted in the “poor” category below: Generally comparable groups are assembled initially but some question remains on whether some (although not major) differences occurred in followup; measurement instruments are acceptable (although not the best) and generally applied equally; some but not all important outcomes are considered; and some but not all potential confounders are accounted for.
- Poor: • Studies will be graded “poor” if any of the following major limitations exist: Groups assembled initially are not close to being comparable or maintained throughout the study; unreliable or invalid measurement instruments are used or not applied equally among groups (including not masking outcome assessment); and key confounders are given little or no attention.

Sources: U.S. Preventive Services Task Force, Procedure Manual, Appendix VII

<http://www.uspreventiveservicestaskforce.org/Page/Name/procedure-manual---appendix-vii>

Harris et al., 2001³¹

Studies of Diagnostic Tests

Criteria

Screening test relevant, available for primary care, adequately described. Although this is one of the USPSTF criteria in its procedures manual, this criterion was not relevant for studies of sensitivity because no reference standard for LTBI exists and the population for sensitivity outcomes are patients with bacteriologic-confirmed active TB.

Study uses a credible reference standard, performed regardless of test results.

Reference standard interpreted independently of screening test.

Handles indeterminate results in a reasonable manner.

Spectrum of patients included in study. Although a USPSTF criterion, this criterion was also not relevant for this topic, given the very specific nature of the population required to estimate sensitivity and specificity in the absence of a reference standard for LTBI.

Appendix B. U.S. Preventive Services Task Force Quality Rating Criteria

Sample size: Although this is one of the criteria listed in the current procedures manual, we did not consider sample size when assessing study quality, as sample size affects precision of the estimate.

Administration of reliable screening test: We also did not consider this criterion, as reliability was itself a separate outcome in this review.

- In addition to the criteria listed in the USPSTF procedures manual, we also considered whether patient selection criteria were clearly described, whether withdrawals were explained, and whether the methods for calculating outcomes were valid.

Definition of Ratings Based on Above Criteria

Good: Relevant and adequately described study populations for the outcome of interest (i.e., Sensitivity, Specificity), screening test well described in terms of test procedures followed and threshold used for a “positive” or “negative” test, credible reference standard used for outcome of interest (i.e., Sensitivity or Specificity), generally interprets reference standard independently of screening test, outcomes clearly reported and valid, handles indeterminate results in a reasonable manner.

Fair: Mostly includes a relevant and adequately described study population for the outcome of interest (i.e., Sensitivity, Specificity), screening test described although may include some ambiguity about test procedures followed or threshold for a “positive” or “negative” test, credible reference standard mostly used for outcome of interest (i.e., Sensitivity or specificity), interpretation of reference standard may or may not be independent of screening test, outcomes mostly clearly reported although may have some ambiguity regarding how indeterminate results were handled.

Poor: Has fatal flaw such as study population not appropriate for outcome of interest (i.e., Sensitivity, Specificity), screening test improperly administered or not at all described, use of noncredible reference standard, reference and screening test not independently assessed, outcomes not clearly or accurately reported with no information about how indeterminate tests were handled.

Criteria Adapted from: U.S. Preventive Services Task Force, Procedure Manual Appendix VII
<http://www.uspreventiveservicestaskforce.org/Page/Name/procedure-manual---appendix-vii>
Harris et al., 2001³¹

Appendix C. Excluded Studies

- X1. Not original research
 - X2. Ineligible Population
 - X3. Ineligible or No Screening/Intervention(s)
 - X4. Ineligible or No Comparator(s)
 - X5. Ineligible or No Outcome(s)
 - X6. Ineligible Setting
 - X7. Ineligible Study Design
 - X8. Could Not Obtain Full Text
 - X9. Poor Quality
 -
1. Sharma SK, Sharma A, Kadiravan T, et al. Rifamycins (rifampicin, rifabutin and rifapentine) compared to isoniazid for preventing tuberculosis in HIV-negative people at risk of active TB. *Cochrane Database Syst Rev*. 2013;7:Cd007545. PMID: 23828580. Exclusion Code: X7
 2. Longhi RM, Zembruski VM, Basta PC, et al. Genetic polymorphism and immune response to tuberculosis in indigenous populations: a brief review. *Braz J Infect Dis*. 2013 May-Jun;17(3):363-8. PMID: 23665009. Exclusion Code: X3
 3. Cohen D, Corbett E. Evidence supports TB test, so what now? *Cochrane Database Syst Rev*. 2013;2:Ed000051. PMID: 23450616. Exclusion Code: X1
 4. Steingart KR, Sohn H, Schiller I, et al. Xpert(R) MTB/RIF assay for pulmonary tuberculosis and rifampicin resistance in adults. *Cochrane Database Syst Rev*. 2013;1:Cd009593. PMID: 23440842. Exclusion Code: X3
 5. Munoz L, Santin M. Interferon-gamma release assays versus tuberculin skin test for targeting people for tuberculosis preventive treatment: an evidence-based review. *J Infect*. 2013 Apr;66(4):381-7. PMID: 23298892. Exclusion Code: X5
 6. Horne DJ, Pinto LM, Arentz M, et al. Diagnostic accuracy and reproducibility of WHO-endorsed phenotypic drug susceptibility testing methods for first-line and second-line antituberculosis drugs. *J Clin Microbiol*. 2013 Feb;51(2):393-401. PMID: 23152548. Exclusion Code: X3
 7. Rogerson TE, Chen S, Kok J, et al. Tests for latent tuberculosis in people with ESRD: a systematic review. *Am J Kidney Dis*. 2013 Jan;61(1):33-43. PMID: 23068425. Exclusion Code: X5
 8. Dai Y, Feng Y, Xu R, et al. Evaluation of interferon-gamma release assays for the diagnosis of tuberculosis: an updated meta-analysis. *Eur J Clin Microbiol Infect Dis*. 2012 Nov;31(11):3127-37. PMID: 22833244. Exclusion Code: X2
 9. Diel R, Loddenkemper R, Nienhaus A. Predictive value of interferon-gamma release assays and tuberculin skin testing for progression from latent TB infection to disease state: a meta-analysis. *Chest*. 2012 Jul;142(1):63-75. PMID: 22490872. Exclusion Code: X2
 10. Fan L, Chen Z, Hao XH, et al. Interferon-gamma release assays for the diagnosis of extrapulmonary tuberculosis: a systematic review and meta-analysis. *FEMS Immunol Med Microbiol*. 2012 Aug;65(3):456-66. PMID: 22487051. Exclusion Code: X2
 11. Chang K, Lu W, Wang J, et al. Rapid and effective diagnosis of tuberculosis and rifampicin resistance with Xpert MTB/RIF assay: a meta-analysis. *J Infect*. 2012 Jun;64(6):580-8. PMID: 22381459. Exclusion Code: X3
 12. Shahidi N, Fu YT, Qian H, et al. Performance of interferon-gamma release assays in patients with inflammatory bowel disease: a systematic review and meta-analysis. *Inflamm Bowel Dis*. 2012 Nov;18(11):2034-42. PMID: 22294550. Exclusion Code: X2
 13. Fenner L, Rieder HL. Isoniazid preventive therapy for all: are we ready? *Int J Tuberc Lung Dis*. 2011 Oct;15(10):1281-2. PMID: 22283884. Exclusion Code: X1
 14. Amerio P, Amoroso G, Bardazzi F, et al. Detection and management of latent tuberculosis infections before biologic therapy for psoriasis. *J Dermatolog Treat*. 2013 Aug;24(4):305-11. PMID: 22208431. Exclusion Code: X1
 15. Nienhaus A, Schablon A, Costa JT, et al. Systematic review of cost and cost-effectiveness of different TB-screening strategies. *BMC Health Serv Res*. 2011;11:247. PMID: 21961888. Exclusion Code: X5
 16. Mrozek N, Pereira B, Soubrier M, et al. Screening of tuberculosis before biologics. *Med Mal Infect*. 2012 Jan;42(1):1-4. PMID: 21907513. Exclusion Code: X7
 17. Rangaka MX, Wilkinson KA, Glynn JR, et al. Predictive value of interferon-gamma release assays for incident active tuberculosis: a systematic review and meta-analysis. *Lancet*

Appendix C. Excluded Studies

- Infect Dis. 2012 Jan;12(1):45-55. PMID: 21846592. Exclusion Code: X5
18. Zhou Q, Chen YQ, Qin SM, et al. Diagnostic accuracy of T-cell interferon-gamma release assays in tuberculous pleurisy: a meta-analysis. *Respirology*. 2011 Apr;16(3):473-80. PMID: 21299686. Exclusion Code: X4
19. Zwerling A, van den Hof S, Scholten J, et al. Interferon-gamma release assays for tuberculosis screening of healthcare workers: a systematic review. *Thorax*. 2012 Jan;67(1):62-70. PMID: 21228420. Exclusion Code: X2
20. Diel R, Goletti D, Ferrara G, et al. Interferon-gamma release assays for the diagnosis of latent *Mycobacterium tuberculosis* infection: a systematic review and meta-analysis. *Eur Respir J*. 2011 Jan;37(1):88-99. PMID: 21030451. Exclusion Code: X2
21. Kunst H, Khan KS. Age-related risk of hepatotoxicity in the treatment of latent tuberculosis infection: a systematic review. *Int J Tuberc Lung Dis*. 2010 Nov;14(11):1374-81. PMID: 20937175. Exclusion Code: X7
22. Erkens CG, Kamphorst M, Abubakar I, et al. Tuberculosis contact investigation in low prevalence countries: a European consensus. *Eur Respir J*. 2010 Oct;36(4):925-49. PMID: 20889463. Exclusion Code: X1
23. Freeman RJ, Mancuso JD, Riddle MS, et al. Systematic review and meta-analysis of TST conversion risk in deployed military and long-term civilian travelers. *J Travel Med*. 2010 Jul-Aug;17(4):233-42. PMID: 20636596. Exclusion Code: X2
24. Greenaway C, Sandoe A, Vissandjee B, et al. Tuberculosis: evidence review for newly arriving immigrants and refugees. *Cmaj*. 2011 Sep 6;183(12):E939-51. PMID: 20634392. Exclusion Code: X7
25. Mazurek GH, Jereb J, Vernon A, et al. Updated guidelines for using Interferon Gamma Release Assays to detect *Mycobacterium tuberculosis* infection - United States, 2010. *MMWR Recomm Rep*. 2010 Jun 25;59(RR-5):1-25. PMID: 20577159. Exclusion Code: X1
26. Chang KC, Leung CC. Systematic review of interferon-gamma release assays in tuberculosis: focus on likelihood ratios. *Thorax*. 2010 Mar;65(3):271-6. PMID: 20335301. Exclusion Code: X2
27. Diel R, Loddenkemper R, Nienhaus A. Evidence-based comparison of commercial interferon-gamma release assays for detecting active TB: a metaanalysis. *Chest*. 2010 Apr;137(4):952-68. PMID: 20022968. Exclusion Code: X2
28. Ziakas PD, Mylonakis E. 4 months of rifampin compared with 9 months of isoniazid for the management of latent tuberculosis infection: a meta-analysis and cost-effectiveness study that focuses on compliance and liver toxicity. *Clin Infect Dis*. 2009 Dec 15;49(12):1883-9. PMID: 19911936. Exclusion Code: X7
29. Bliven EE, Podewils LJ. The role of chronic hepatitis in isoniazid hepatotoxicity during treatment for latent tuberculosis infection. *Int J Tuberc Lung Dis*. 2009 Sep;13(9):1054-60. PMID: 19723392. Exclusion Code: X7
30. Sadatsafavi M, Shahidi N, Marra F, et al. A statistical method was used for the meta-analysis of tests for latent TB in the absence of a gold standard, combining random-effect and latent-class methods to estimate test accuracy. *J Clin Epidemiol*. 2010 Mar;63(3):257-69. PMID: 19692208. Exclusion Code: X5
31. Pai M, Ramsay A, O'Brien R. Evidence-based tuberculosis diagnosis. *PLoS Med*. 2008 Jul 22;5(7):e156. PMID: 18651788. Exclusion Code: X1
32. Pai M, Zwerling A, Menzies D. Systematic review: T-cell-based assays for the diagnosis of latent tuberculosis infection: an update. *Ann Intern Med*. 2008 Aug 5;149(3):177-84. PMID: 18593687. Exclusion Code: X2
33. Madariaga MG, Jalali Z, Swindells S. Clinical utility of interferon gamma assay in the diagnosis of tuberculosis. *J Am Board Fam Med*. 2007 Nov-Dec;20(6):540-7. PMID: 17954861. Exclusion Code: X7
34. Menzies D, Pai M, Comstock G. Meta-analysis: new tests for the diagnosis of latent tuberculosis infection: areas of uncertainty and recommendations for research. *Ann Intern Med*. 2007 Mar 6;146(5):340-54. PMID: 17339619. Exclusion Code: X2
35. Dinnes J, Deeks J, Kunst H, et al. A systematic review of rapid diagnostic tests for the detection of tuberculosis infection. *Health Technol Assess*. 2007 Jan;11(3):1-196. PMID: 17266837. Exclusion Code: X3
36. Farhat M, Greenaway C, Pai M, et al. False-positive tuberculin skin tests: what is the absolute effect of BCG and non-tuberculous mycobacteria? *Int J Tuberc Lung Dis*. 2006 Nov;10(11):1192-204. PMID: 17131776. Exclusion Code: X2
37. Gao XF, Wang L, Liu GJ, et al. Rifampicin plus pyrazinamide versus isoniazid for treating latent tuberculosis infection: a meta-analysis. *Int J Tuberc Lung Dis*. 2006 Oct;10(10):1080-90. PMID: 17044199. Exclusion Code: X7

Appendix C. Excluded Studies

38. Balcells ME, Thomas SL, Godfrey-Faussett P, et al. Isoniazid preventive therapy and risk for resistant tuberculosis. *Emerg Infect Dis*. 2006 May;12(5):744-51. PMID: 16704830. Exclusion Code: X2
39. Fraser A, Paul M, Attamna A, et al. Treatment of latent tuberculosis in persons at risk for multidrug-resistant tuberculosis: systematic review. *Int J Tuberc Lung Dis*. 2006 Jan;10(1):19-23. PMID: 16466032. Exclusion Code: X2
40. Ena J, Valls V. Short-course therapy with rifampin plus isoniazid, compared with standard therapy with isoniazid, for latent tuberculosis infection: a meta-analysis. *Clin Infect Dis*. 2005 Mar 1;40(5):670-6. PMID: 15714411. Exclusion Code: X4
41. Pai M, Riley LW, Colford JM, Jr. Interferon-gamma assays in the immunodiagnosis of tuberculosis: a systematic review. *Lancet Infect Dis*. 2004 Dec;4(12):761-76. PMID: 15567126. Exclusion Code: X2
42. Smieja MJ, Marchetti CA, Cook DJ, et al. Isoniazid for preventing tuberculosis in non-HIV infected persons. *Cochrane Database Syst Rev*. 2000(2):Cd001363. PMID: 10796642. Exclusion Code: X7
43. Temple ME, Nahata MC. Rifapentine: its role in the treatment of tuberculosis. *Ann Pharmacother*. 1999 Nov;33(11):1203-10. PMID: 10573321. Exclusion Code: X2
44. Steele MA, Burk RF, DesPrez RM. Toxic hepatitis with isoniazid and rifampin. A meta-analysis. *Chest*. 1991 Feb;99(2):465-71. PMID: 1824929. Exclusion Code: X7
45. O'Shea MK, Fletcher TE, Tupper D, et al. Screening for latent tuberculosis and gastrointestinal parasite infections in Gurkha recruits: research driving policy change. *J R Army Med Corps*. 2014 Jun;160(2):180-2. PMID: 24607484. Exclusion Code: X2
46. Ang M, Wong WL, Kiew SY, et al. Prospective head-to-head study comparing 2 commercial interferon gamma release assays for the diagnosis of tuberculous uveitis. *Am J Ophthalmol*. 2014 Jun;157(6):1306-14; 14.e1-4. PMID: 24508163. Exclusion Code: X2
47. Auld SC, Click ES, Heilig CM, et al. Tuberculin skin test result and risk of death among persons with active TB. *PLoS One*. 2013;8(11):e78779. PMID: 24244358. Exclusion Code: X5
48. Auld SC, Click ES, Heilig CM, et al. Association between tuberculin skin test result and clinical presentation of tuberculosis disease. *BMC Infect Dis*. 2013;13:460. PMID: 24093965. Exclusion Code: X2
49. Moran-Mendoza O, Tello-Zavala MC, Rivera-Camarillo M, et al. Comparison of different methods and times for reading the tuberculin skin test. *Int J Tuberc Lung Dis*. 2013 Oct;17(10):1273-8. PMID: 24025377. Exclusion Code: X5
50. Slater ML, Welland G, Pai M, et al. Challenges with QuantiFERON-TB Gold assay for large-scale, routine screening of U.S. healthcare workers. *Am J Respir Crit Care Med*. 2013 Oct 15;188(8):1005-10. PMID: 23978270. Exclusion Code: X2
51. Gaur RL, Pai M, Banaei N. Impact of blood volume, tube shaking, and incubation time on reproducibility of QuantiFERON-TB gold in-tube assay. *J Clin Microbiol*. 2013 Nov;51(11):3521-6. PMID: 23966505. Exclusion Code: X4
52. Lee YM, Park KH, Kim SM, et al. Risk factors for false-negative results of T-SPOT.TB and tuberculin skin test in extrapulmonary tuberculosis. *Infection*. 2013 Dec;41(6):1089-95. PMID: 23943073. Exclusion Code: X2
53. Shu CC, Wu VC, Yang FJ, et al. Dynamic changes in positive interferon-gamma release assay in a dialysis population: An observational cohort study. *J Infect*. 2013 Dec;67(6):529-35. PMID: 23933475. Exclusion Code: X2
54. Duque A, Lin SY, Desmond E, et al. Evaluation of the BD Bactec MGIT 320 system for detection of mycobacteria and drug susceptibility testing of *Mycobacterium tuberculosis*. *J Clin Microbiol*. 2013 Oct;51(10):3403-5. PMID: 23863564. Exclusion Code: X2
55. Dobler CC. What do we know about the outcomes of tuberculosis contact investigations in NSW? *N S W Public Health Bull*. 2013 Jul;24(1):34-7. PMID: 23849028. Exclusion Code: X2
56. Ang M, Wong WL, Li X, et al. Interferon gamma release assay for the diagnosis of uveitis associated with tuberculosis: a Bayesian evaluation in the absence of a gold standard. *Br J Ophthalmol*. 2013 Aug;97(8):1062-7. PMID: 23723411. Exclusion Code: X2
57. Guglielmetti L, Cazzadori A, Conti M, et al. Lymphocyte subpopulations in active tuberculosis: association with disease severity and the QFT-GIT assay. *Int J Tuberc Lung Dis*. 2013 Jun;17(6):825-8. PMID: 23676170. Exclusion Code: X3
58. Zwerling A, Benedetti A, Cojocariu M, et al. Repeat IGRA testing in Canadian health workers: conversions or unexplained variability? *PLoS One*. 2013;8(1):e54748. PMID: 23382955. Exclusion Code: X5

Appendix C. Excluded Studies

59. Aabye MG, Latorre I, Diaz J, et al. Dried plasma spots in the diagnosis of tuberculosis: IP-10 release assay on filter paper. *Eur Respir J*. 2013 Aug;42(2):495-503. PMID: 23349445. Exclusion Code: X2
60. Chawla H, Lobato MN, Sosa LE, et al. Predictors for a positive QuantiFERON-TB-Gold test in BCG-vaccinated adults with a positive tuberculin skin test. *J Infect Public Health*. 2012 Dec;5(6):369-73. PMID: 23287606. Exclusion Code: X2
61. Lytras T, Spala G, Bonovas S, et al. Evaluation of tuberculosis underreporting in Greece through comparison with anti-tuberculosis drug consumption. *PLoS One*. 2012;7(11):e50033. PMID: 23185524. Exclusion Code: X2
62. Tiernan JF, Gilhooley S, Jones ME, et al. Does an interferon-gamma release assay change practice in possible latent tuberculosis? *Qjm*. 2013 Feb;106(2):139-46. PMID: 23159840. Exclusion Code: X5
63. del Campo MT, Fouad H, Solis-Bravo MM, et al. Cost-effectiveness of different screening strategies (single or dual) for the diagnosis of tuberculosis infection in healthcare workers. *Infect Control Hosp Epidemiol*. 2012 Dec;33(12):1226-34. PMID: 23143360. Exclusion Code: X5
64. Li H, Yang L, Zheng CY, et al. Use of bronchoalveolar lavage enzyme-linked immunospot for diagnosis of smear-negative pulmonary tuberculosis. *Int J Tuberc Lung Dis*. 2012 Dec;16(12):1668-73. PMID: 23131267. Exclusion Code: X9
65. Kamiya H, Ikushima S, Kondo K, et al. Diagnostic performance of interferon-gamma release assays in elderly populations in comparison with younger populations. *J Infect Chemother*. 2013 Apr;19(2):217-22. PMID: 23108426. Exclusion Code: X9
66. Kim SY, Park MS, Kim YS, et al. Conversion rates of an interferon-gamma release assay and the tuberculin skin test in the serial monitoring of healthcare workers. *Infection*. 2013 Apr;41(2):511-6. PMID: 23104257. Exclusion Code: X5
67. Chan PC, Yang CH, Chang LY, et al. Lower prevalence of tuberculosis infection in BCG vaccinees: a cross-sectional study in adult prison inmates. *Thorax*. 2013 Mar;68(3):263-8. PMID: 23019256. Exclusion Code: X2
68. Shrestha R, Gyawali P, Yadav BK, et al. In-vitro assessment of cell-mediated immunity by demonstrating effector-t cells for diagnosis of tuberculosis in Nepalese subjects. *Nepal Med Coll J*. 2011 Dec;13(4):275-8. PMID: 23016479. Exclusion Code: X9
69. He GX, Wang LX, Chai SJ, et al. Risk factors associated with tuberculosis infection among health care workers in Inner Mongolia, China. *Int J Tuberc Lung Dis*. 2012 Nov;16(11):1485-91. PMID: 22964074. Exclusion Code: X5
70. Haldar P, Thuraisingam H, Patel H, et al. Single-step QuantiFERON screening of adult contacts: a prospective cohort study of tuberculosis risk. *Thorax*. 2013 Mar;68(3):240-6. PMID: 22956558. Exclusion Code: X2
71. Campainha S, Gomes T, Carvalho A, et al. Negative predictive value of TST and IGRA in anti-TNF treated patients. *Eur Respir J*. 2012 Sep;40(3):790-1. PMID: 22941549. Exclusion Code: X2
72. Zwerling A, Cojocariu M, McIntosh F, et al. TB screening in Canadian health care workers using interferon-gamma release assays. *PLoS One*. 2012;7(8):e43014. PMID: 22916197. Exclusion Code: X5
73. Shu CC, Wu VC, Yang FJ, et al. Predictors and prevalence of latent tuberculosis infection in patients receiving long-term hemodialysis and peritoneal dialysis. *PLoS One*. 2012;7(8):e42592. PMID: 22916137. Exclusion Code: X5
74. Rose MV, Kimaro G, Kroidl I, et al. Evaluation of QuantiFERON microtube, using 0.9 mL blood, for diagnosing tuberculosis infection. *Eur Respir J*. 2013 Apr;41(4):909-16. PMID: 22878880. Exclusion Code: X2
75. Jo KW, Jeon K, Kang YA, et al. Poor correlation between tuberculin skin tests and interferon-gamma assays in close contacts of patients with multidrug-resistant tuberculosis. *Respirology*. 2012 Oct;17(7):1125-30. PMID: 22758779. Exclusion Code: X2
76. Banfield S, Pascoe E, Thambiran A, et al. Factors associated with the performance of a blood-based interferon-gamma release assay in diagnosing tuberculosis. *PLoS One*. 2012;7(6):e38556. PMID: 22701664. Exclusion Code: X5
77. Pareek M, Bond M, Shorey J, et al. Community-based evaluation of immigrant tuberculosis screening using interferon gamma release assays and tuberculin skin testing: observational study and economic analysis. *Thorax*. 2013 Mar;68(3):230-9. PMID: 22693179. Exclusion Code: X2
78. Gautam M, Darroch J, Bassett P, et al. Tuberculosis infection in the indigenous elderly White UK population: a study of IGRAs. *Int J*

Appendix C. Excluded Studies

- Tuberc Lung Dis. 2012 Apr;16(4):564. PMID: 22640515. Exclusion Code: X7
79. Lee SW, Lee SH, Yim JJ. Serial interferon-gamma release assays after chemoprophylaxis in a tuberculosis outbreak cohort. *Infection*. 2012 Aug;40(4):431-5. PMID: 22585454. Exclusion Code: X5
 80. Turtle L, Kemp T, Davies GR, et al. In routine UK hospital practice T-SPOT.TB is useful in some patients with a modest pre-test probability of active tuberculosis. *Eur J Intern Med*. 2012 Jun;23(4):363-7. PMID: 22560387. Exclusion Code: X9
 81. Chan PC, Yang CH, Chang LY, et al. Latent tuberculosis infection treatment for prison inmates: a randomised controlled trial. *Int J Tuberc Lung Dis*. 2012 May;16(5):633-8. PMID: 22410137. Exclusion Code: X6
 82. Ang M, Wong W, Ngan CC, et al. Interferon-gamma release assay as a diagnostic test for tuberculosis-associated uveitis. *Eye (Lond)*. 2012 May;26(5):658-65. PMID: 22302066. Exclusion Code: X2
 83. Welch RJ, Lawless KM, Litwin CM. Antituberculosis IgG antibodies as a marker of active *Mycobacterium tuberculosis* disease. *Clin Vaccine Immunol*. 2012 Apr;19(4):522-6. PMID: 22301692. Exclusion Code: X2
 84. Leiro-Fernandez V, Valverde D, Vazquez-Gallardo R, et al. N-acetyltransferase 2 polymorphisms and risk of anti-tuberculosis drug-induced hepatotoxicity in Caucasians. *Int J Tuberc Lung Dis*. 2011 Oct;15(10):1403-8. PMID: 22283902. Exclusion Code: X7
 85. Altunoren O, Kahraman H, Sayarlioglu H, et al. The affecting factors and comparison of tuberculin skin test in peritoneal dialysis and hemodialysis patients. *Ren Fail*. 2012;34(3):304-7. PMID: 22260191. Exclusion Code: X5
 86. Noorbakhsh S, Mousavi J, Barati M, et al. Evaluation of an interferon-gamma release assay in young contacts of active tuberculosis cases. *East Mediterr Health J*. 2011 Sep;17(9):714-8. PMID: 22259925. Exclusion Code: X5
 87. Abu-Taleb AM, El-Sokkary RH, El Tarhouny SA. Interferon-gamma release assay for detection of latent tuberculosis infection in casual and close contacts of tuberculosis cases. *East Mediterr Health J*. 2011 Oct;17(10):749-53. PMID: 22256408. Exclusion Code: X2
 88. Nogueira PA, Abrahao RM, Galesi VM. Tuberculosis and latent tuberculosis in prison inmates. *Rev Saude Publica*. 2012 Feb;46(1):119-27. PMID: 22252791. Exclusion Code: X5
 89. Marco A, Sole N, Orcau A, et al. Prevalence of latent tuberculosis infection in inmates recently incarcerated in a men's prison in Barcelona. *Int J Tuberc Lung Dis*. 2012 Jan;16(1):60-4. PMID: 22236847. Exclusion Code: X2
 90. Wang JY, Shu CC, Lee CH, et al. Interferon-gamma release assay and Rifampicin therapy for household contacts of tuberculosis. *J Infect*. 2012 Mar;64(3):291-8. PMID: 22207002. Exclusion Code: X2
 91. Maden E, Bekci TT, Kesli R, et al. Evaluation of performance of quantiferon assay and tuberculin skin test in end stage renal disease patients receiving hemodialysis. *New Microbiol*. 2011 Oct;34(4):351-6. PMID: 22143808. Exclusion Code: X2
 92. Yilmaz N, Zehra Aydin S, Inanc N, et al. Comparison of QuantiFERON-TB Gold test and tuberculin skin test for the identification of latent *Mycobacterium tuberculosis* infection in lupus patients. *Lupus*. 2012 Apr;21(5):491-5. PMID: 22140142. Exclusion Code: X2
 93. Joshi R, Narang U, Zwerling A, et al. Predictive value of latent tuberculosis tests in Indian healthcare workers: a cohort study. *Eur Respir J*. 2011 Dec;38(6):1475-7. PMID: 22130764. Exclusion Code: X6
 94. Khalil EA, Elnour AA, Musa AM, et al. Tuberculous lymphadenitis: skin delayed-type hypersensitivity reaction and cellular immune responses. *West Afr J Med*. 2011 May-Jun;30(3):193-6. PMID: 22120485. Exclusion Code: X2
 95. Banach DB, Harris TG. Indeterminate QuantiFERON(R)-TB Gold results in a public health clinic setting. *Int J Tuberc Lung Dis*. 2011 Dec;15(12):1623-30. PMID: 22118169. Exclusion Code: X2
 96. Grinsdale JA, Ho CS, Banouvong H, et al. Programmatic impact of using QuantiFERON(R)-TB Gold in routine contact investigation activities. *Int J Tuberc Lung Dis*. 2011 Dec;15(12):1614-20. PMID: 22118167. Exclusion Code: X2
 97. Gupta D, Kumar S, Aggarwal AN, et al. Interferon gamma release assay (QuantiFERON-TB Gold In Tube) in patients of sarcoidosis from a population with high prevalence of tuberculosis infection. *Sarcoidosis Vasc Diffuse Lung Dis*. 2011 Oct;28(2):95-101. PMID: 22117500. Exclusion Code: X3
 98. Worjolah A, Kato-Maeda M, Osmond D, et al. Interferon gamma release assay compared with the tuberculin skin test for latent tuberculosis detection in pregnancy. *Obstet Gynecol*. 2011

Appendix C. Excluded Studies

- Dec;118(6):1363-70. PMID: 22105266. Exclusion Code: X2
99. Gonzalez-Salazar F, Vargas-Villarreal J, Garcialuna-Martinez FJ, et al. Snapshot of QuantiFERON TB gold testing in Northern Mexico. *Tuberculosis (Edinb)*. 2011 Dec;91 Suppl 1:S34-7. PMID: 22099419. Exclusion Code: X2
100. Friedrich SO, von Groote-Bidlingmaier F, Diacon AH. Xpert MTB/RIF assay for diagnosis of pleural tuberculosis. *J Clin Microbiol*. 2011 Dec;49(12):4341-2. PMID: 21998430. Exclusion Code: X3
101. Hussain R, Ansari A, Talat N, et al. CCL2/MCP-I genotype-phenotype relationship in latent tuberculosis infection. *PLoS One*. 2011;6(10):e25803. PMID: 21991356. Exclusion Code: X6
102. Soysal A, Toprak D, Koc M, et al. Diagnosing latent tuberculosis infection in haemodialysis patients: T-cell based assay (T-SPOT.TB) or tuberculin skin test? *Nephrol Dial Transplant*. 2012 Apr;27(4):1645-50. PMID: 21931124. Exclusion Code: X6
103. Schablon A, Diel R, Diner G, et al. Specificity of a whole blood IGRA in German nursing students. *BMC Infect Dis*. 2011;11:245. PMID: 21929799. Exclusion Code: X2
104. Fresard I, Bridevaux PO, Rochat T, et al. Adverse effects and adherence to treatment of rifampicin 4 months vs isoniazid 6 months for latent tuberculosis: a retrospective analysis. *Swiss Med Wkly*. 2011;141:w13240. PMID: 21842452. Exclusion Code: X7
105. Soares Pires F, Pinto C, Duarte R. Who misses the second step of evaluation in tuberculosis contact screening? *Eur Respir J*. 2011 Aug;38(2):474-6. PMID: 21804166. Exclusion Code: X5
106. Mancuso JD, Tribble D, Mazurek GH, et al. Impact of targeted testing for latent tuberculosis infection using commercially available diagnostics. *Clin Infect Dis*. 2011 Aug 1;53(3):234-44. PMID: 21765072. Exclusion Code: X5
107. Kim SH, Lee SO, Park JB, et al. A prospective longitudinal study evaluating the usefulness of a T-cell-based assay for latent tuberculosis infection in kidney transplant recipients. *Am J Transplant*. 2011 Sep;11(9):1927-35. PMID: 21749641. Exclusion Code: X2
108. Weinfurter P, Blumberg HM, Goldbaum G, et al. Predictors of discordant tuberculin skin test and QuantiFERON(R)-TB Gold In-Tube results in various high-risk groups. *Int J Tuberc Lung Dis*. 2011 Aug;15(8):1056-61. PMID: 21740668. Exclusion Code: X2
109. Doberne D, Gaur RL, Banaei N. Preanalytical delay reduces sensitivity of QuantiFERON-TB gold in-tube assay for detection of latent tuberculosis infection. *J Clin Microbiol*. 2011 Aug;49(8):3061-4. PMID: 21697332. Exclusion Code: X3
110. Bradshaw L, Davies E, Devine M, et al. The role of the interferon gamma release assay in assessing recent tuberculosis transmission in a hospital incident. *PLoS One*. 2011;6(6):e20770. PMID: 21695149. Exclusion Code: X4
111. Sauzullo I, Massetti AP, Mengoni F, et al. Influence of previous tuberculin skin test on serial IFN-gamma release assays. *Tuberculosis (Edinb)*. 2011 Jul;91(4):322-6. PMID: 21664872. Exclusion Code: X5
112. de Andrade Lima E, de Andrade Lima M, de Lorena VM, et al. Evaluation of an IFN-gamma assay in the diagnosis of latent tuberculosis in patients with psoriasis in a highly endemic setting. *Acta Derm Venereol*. 2011 Oct;91(6):694-7. PMID: 21629971. Exclusion Code: X5
113. Komiya K, Ariga H, Nagai H, et al. Reversion rates of QuantiFERON-TB Gold are related to pre-treatment IFN-gamma levels. *J Infect*. 2011 Jul;63(1):48-53. PMID: 21624664. Exclusion Code: X4
114. Milman N, Soborg B, Svendsen CB, et al. QuantiFERON test for tuberculosis screening in sarcoidosis patients. *Scand J Infect Dis*. 2011 Sep;43(9):728-35. PMID: 21619424. Exclusion Code: X2
115. Ringshausen FC, Nienhaus A, Torres Costa J, et al. Within-subject variability of Mycobacterium tuberculosis-specific gamma interferon responses in German health care workers. *Clin Vaccine Immunol*. 2011 Jul;18(7):1176-82. PMID: 21593237. Exclusion Code: X2
116. Kim YS, Kim YH, Kim WH, et al. Diagnostic utility of anti-Saccharomyces cerevisiae antibody (ASCA) and Interferon-gamma assay in the differential diagnosis of Crohn's disease and intestinal tuberculosis. *Clin Chim Acta*. 2011 Aug 17;412(17-18):1527-32. PMID: 21575618. Exclusion Code: X2
117. Risk of tuberculosis among contacts of isoniazid-resistant and isoniazid-susceptible cases. *Int J Tuberc Lung Dis*. 2011 Jun;15(6):782-8. PMID: 21575299. Exclusion Code: X2
118. Takeda N, Nojima T, Terao C, et al. Interferon-gamma release assay for diagnosing Mycobacterium tuberculosis infections in patients with systemic lupus erythematosus.

Appendix C. Excluded Studies

- Lupus. 2011 Jul;20(8):792-800. PMID: 21562022. Exclusion Code: X2
119. Caglayan V, Ak O, Dabak G, et al. Comparison of tuberculin skin testing and QuantiFERON-TB Gold-In Tube test in health care workers. *Tuberk Toraks*. 2011;59(1):43-7. PMID: 21554229. Exclusion Code: X2
 120. Pareek M, Watson JP, Ormerod LP, et al. Screening of immigrants in the UK for imported latent tuberculosis: a multicentre cohort study and cost-effectiveness analysis. *Lancet Infect Dis*. 2011 Jun;11(6):435-44. PMID: 21514236. Exclusion Code: X4
 121. Lee JE, Kim HJ, Lee SW. The clinical utility of tuberculin skin test and interferon-gamma release assay in the diagnosis of active tuberculosis among young adults: a prospective observational study. *BMC Infect Dis*. 2011;11:96. PMID: 21501477. Exclusion Code: X2
 122. Jafari C, Kessler P, Sotgiu G, et al. Impact of a *Mycobacterium tuberculosis*-specific interferon-gamma release assay in bronchoalveolar lavage fluid for a rapid diagnosis of tuberculosis. *J Intern Med*. 2011 Sep;270(3):254-62. PMID: 21418341. Exclusion Code: X5
 123. Kim HJ, Yoon HI, Park KU, et al. The impact of previous tuberculosis history on T-SPOT.TB(R) interferon-gamma release assay results. *Int J Tuberc Lung Dis*. 2011 Apr;15(4):510-6. PMID: 21396211. Exclusion Code: X2
 124. Ling DI, Pai M, Davids V, et al. Are interferon-gamma release assays useful for diagnosing active tuberculosis in a high-burden setting? *Eur Respir J*. 2011 Sep;38(3):649-56. PMID: 21349910. Exclusion Code: X2
 125. Smith BM, Schwartzman K, Bartlett G, et al. Adverse events associated with treatment of latent tuberculosis in the general population. *Cmaj*. 2011 Feb 22;183(3):E173-9. PMID: 21220436. Exclusion Code: X7
 126. Anibarro L, Trigo M, Villaverde C, et al. Interferon-gamma release assays in tuberculosis contacts: is there a window period? *Eur Respir J*. 2011 Jan;37(1):215-7. PMID: 21205718. Exclusion Code: X2
 127. Gilham L, France J, Stirling A, et al. Tuberculosis screening before biologics -- T-SPOT for all? *J Rheumatol*. 2011 Jan;38(1):179. PMID: 21196587. Exclusion Code: X2
 128. Leung EC, Leung CC, Leung WW, et al. Role of whole-blood interferon-gamma release assay in the diagnosis of smear-negative tuberculosis. *Int J Tuberc Lung Dis*. 2010 Dec;14(12):1564-70. PMID: 21144241. Exclusion Code: X2
 129. Zhang M, Wang H, Liao M, et al. Diagnosis of latent tuberculosis infection in bacille Calmette-Guerin vaccinated subjects in China by interferon-gamma ELISpot assay. *Int J Tuberc Lung Dis*. 2010 Dec;14(12):1556-63. PMID: 21144240. Exclusion Code: X2
 130. Kim BJ, Choi YS, Jang BI, et al. Prospective evaluation of the clinical utility of interferon-gamma assay in the differential diagnosis of intestinal tuberculosis and Crohn's disease. *Inflamm Bowel Dis*. 2011 Jun;17(6):1308-13. PMID: 21053248. Exclusion Code: X2
 131. Gandra S, Scott WS, Somaraju V, et al. Questionable effectiveness of the QuantiFERON-TB Gold Test (Cellestis) as a screening tool in healthcare workers. *Infect Control Hosp Epidemiol*. 2010 Dec;31(12):1279-85. PMID: 20979495. Exclusion Code: X2
 132. Lee SW, Lee CT, Yim JJ. Serial interferon-gamma release assays during treatment of active tuberculosis in young adults. *BMC Infect Dis*. 2010;10:300. PMID: 20950477. Exclusion Code: X2
 133. Connell DW, Singanayagam A, Charif R, et al. A comparison between interferon gamma release assays and the tuberculin skin test in the contact tracing of patients with chronic kidney disease. *Thorax*. 2011 Aug;66(8):729-30; author reply 30. PMID: 20947898. Exclusion Code: X2
 134. Ball PM, Pernollet M, Bouillet L, et al. Usefulness of an in-vitro tuberculosis interferon-gamma release assay (T-SPOT.TB) in the first-line check-up of uveitis patients. *Ann Med*. 2010 Oct;42(7):546-54. PMID: 20868342. Exclusion Code: X2
 135. Baboolal S, Ramoutar D, Akpaka PE. Comparison of the QuantiFERON(R)-TB Gold assay and tuberculin skin test to detect latent tuberculosis infection among target groups in Trinidad & Tobago. *Rev Panam Salud Publica*. 2010 Jul;28(1):36-42. PMID: 20857019. Exclusion Code: X2
 136. Chigbu LN, Iroegbu CU. Incidence and spread of *Mycobacterium tuberculosis*-associated infection among Aba Federal prison inmates in Nigeria. *J Health Popul Nutr*. 2010 Aug;28(4):327-32. PMID: 20824975. Exclusion Code: X2
 137. Petrescu L, Stancu S, Tardei G, et al. Tuberculin skin test, interferon-gamma assay, and T cells subpopulations in hemodialysis patients. *J Ren Nutr*. 2010 Sep;20(5 Suppl):S109-17. PMID: 20797557. Exclusion Code: X2
 138. Harstad I, Winje BA, Heldal E, et al. Predictive values of QuantiFERON-TB Gold testing in screening for tuberculosis disease in asylum seekers. *Int J Tuberc Lung Dis*. 2010 Sep;14(9):1209-11. PMID: 20819271. Exclusion Code: X2

Appendix C. Excluded Studies

139. Dilektasli AG, Erdem E, Durukan E, et al. Feasibility of the T-SPOT.TB assay for the immunodiagnosis of smear-positive pulmonary tuberculosis using induced sputum. *Int J Tuberc Lung Dis*. 2010 Sep;14(9):1205-8. PMID: 20819270. Exclusion Code: X2
140. Diel R, Loddenkemper R, Niemann S, et al. Negative and positive predictive value of a whole-blood interferon-gamma release assay for developing active tuberculosis: an update. *Am J Respir Crit Care Med*. 2011 Jan 1;183(1):88-95. PMID: 20802162. Exclusion Code: X2
141. Person AK, Goswami ND, Bisette DJ, et al. Pairing QuantiFERON gold in-tube with opt-out HIV testing in a tuberculosis contact investigation in the Southeastern United States. *AIDS Patient Care STDS*. 2010 Sep;24(9):539-43. PMID: 20731612. Exclusion Code: X2
142. Spence MR, Sibert D, Fleischer L, et al. A field trial evaluating the Quantiferon Gold tuberculosis tube test in an endemic population. *Am J Infect Control*. 2011 Feb;39(1):79-80. PMID: 20673599. Exclusion Code: X2
143. Ringshausen FC, Nienhaus A, Schablon A, et al. Predictors of persistently positive Mycobacterium-tuberculosis-specific interferon-gamma responses in the serial testing of health care workers. *BMC Infect Dis*. 2010;10:220. PMID: 20653946. Exclusion Code: X2
144. Aspler A, Long R, Trajman A, et al. Impact of treatment completion, intolerance and adverse events on health system costs in a randomised trial of 4 months rifampin or 9 months isoniazid for latent TB. *Thorax*. 2010 Jul;65(7):582-7. PMID: 20627913. Exclusion Code: X5
145. Gallant CJ, Cobat A, Simkin L, et al. Impact of age and sex on mycobacterial immunity in an area of high tuberculosis incidence. *Int J Tuberc Lung Dis*. 2010 Aug;14(8):952-9. PMID: 20626938. Exclusion Code: X2
146. Wang SH, Powell DA, Nagaraja HN, et al. Evaluation of a modified interferon-gamma release assay for the diagnosis of latent tuberculosis infection in adult and paediatric populations that enables delayed processing. *Scand J Infect Dis*. 2010 Dec;42(11-12):845-50. PMID: 20608764. Exclusion Code: X2
147. Wu X, Li Q, Liang Y, et al. Clinical evaluation of a homemade enzyme-linked immunospot assay for the diagnosis of active tuberculosis in China. *Mol Biotechnol*. 2011 Jan;47(1):18-25. PMID: 20596901. Exclusion Code: X3
148. Guo N, Marra F, Fitzgerald JM, et al. Impact of adverse drug reaction and predictivity of quality of life status in tuberculosis. *Eur Respir J*. 2010 Jul;36(1):206-8. PMID: 20595167. Exclusion Code: X3
149. Abdalhamid B, Hinrichs SH, Garrett JL, et al. Utilization of the QuantiFERON-TB Gold test in a two-step process with the tuberculin skin test to evaluate health care workers for latent tuberculosis. *J Clin Microbiol*. 2010 Aug;48(8):2955-6. PMID: 20573876. Exclusion Code: X2
150. Yoshiyama T, Harada N, Higuchi K, et al. Use of the QuantiFERON-TB Gold test for screening tuberculosis contacts and predicting active disease. *Int J Tuberc Lung Dis*. 2010 Jul;14(7):819-27. PMID: 20550763. Exclusion Code: X2
151. Lee SS, Chou KJ, Dou HY, et al. High prevalence of latent tuberculosis infection in dialysis patients using the interferon-gamma release assay and tuberculin skin test. *Clin J Am Soc Nephrol*. 2010 Aug;5(8):1451-7. PMID: 20538837. Exclusion Code: X2
152. Jong Lee K, Ae Kang Y, Mi Kim Y, et al. Screening for latent tuberculosis infection in South Korean healthcare workers using a tuberculin skin test and whole blood interferon-gamma assay. *Scand J Infect Dis*. 2010 Sep;42(9):672-8. PMID: 20482459. Exclusion Code: X2
153. Goletti D, Raja A, Ahamed Kabeer BS, et al. IFN-gamma, but not IP-10, MCP-2 or IL-2 response to RD1 selected peptides associates to active tuberculosis. *J Infect*. 2010 Jul;61(2):133-43. PMID: 20470822. Exclusion Code: X2
154. Lienhardt C, Fielding K, Hane AA, et al. Evaluation of the prognostic value of IFN-gamma release assay and tuberculin skin test in household contacts of infectious tuberculosis cases in Senegal. *PLoS One*. 2010;5(5):e10508. PMID: 20463900. Exclusion Code: X2
155. Cashmore TJ, Peter JG, van Zyl-Smit RN, et al. Feasibility and diagnostic utility of antigen-specific interferon-gamma responses for rapid immunodiagnosis of tuberculosis using induced sputum. *PLoS One*. 2010;5(4):e10389. PMID: 20442850. Exclusion Code: X3
156. Patel VB, Singh R, Connolly C, et al. Cerebrospinal T-cell responses aid in the diagnosis of tuberculous meningitis in a human immunodeficiency virus- and tuberculosis-endemic population. *Am J Respir Crit Care Med*. 2010 Aug 15;182(4):569-77. PMID: 20442433. Exclusion Code: X2
157. Chung WK, Zheng ZL, Kim HS, et al. Serial testing of interferon-gamma-release assays for the diagnosis of latent tuberculosis in hemodialysis patients. *J Infect*. 2010

Appendix C. Excluded Studies

- Jul;61(2):144-9. PMID: 20435059. Exclusion Code: X2
158. Orlando G, Merli S, Cordier L, et al. Interferon-gamma releasing assay versus tuberculin skin testing for latent tuberculosis infection in targeted screening programs for high risk immigrants. *Infection*. 2010 Jun;38(3):195-204. PMID: 20411295. Exclusion Code: X2
 159. Simsek H, Alpar S, Ucar N, et al. Comparison of tuberculin skin testing and T-SPOT.TB for diagnosis of latent and active tuberculosis. *Jpn J Infect Dis*. 2010 Mar;63(2):99-102. PMID: 20332570. Exclusion Code: X2
 160. Dyrhol-Riise AM, Gran G, Wentzel-Larsen T, et al. Diagnosis and follow-up of treatment of latent tuberculosis; the utility of the QuantiFERON-TB Gold In-tube assay in outpatients from a tuberculosis low-endemic country. *BMC Infect Dis*. 2010;10:57. PMID: 20210999. Exclusion Code: X2
 161. Sheriff FG, Manji KP, Manji MP, et al. Latent tuberculosis among pregnant mothers in a resource poor setting in Northern Tanzania: a cross-sectional study. *BMC Infect Dis*. 2010;10:52. PMID: 20205938. Exclusion Code: X2
 162. Cordova J, Shiloh R, Gilman RH, et al. Evaluation of molecular tools for detection and drug susceptibility testing of *Mycobacterium tuberculosis* in stool specimens from patients with pulmonary tuberculosis. *J Clin Microbiol*. 2010 May;48(5):1820-6. PMID: 20200293. Exclusion Code: X3
 163. Kleinert S, Kurzai O, Elias J, et al. Comparison of two interferon-gamma release assays and tuberculin skin test for detecting latent tuberculosis in patients with immune-mediated inflammatory diseases. *Ann Rheum Dis*. 2010 Apr;69(4):782-4. PMID: 20185504. Exclusion Code: X2
 164. Saracino A, Scotto G, Fornabaio C, et al. QuantiFERON-TB Gold In-Tube test (QFT-GIT) for the screening of latent tuberculosis in recent immigrants to Italy. *New Microbiol*. 2009 Oct;32(4):369-76. PMID: 20128443. Exclusion Code: X2
 165. Bua A, Mollicotti P, Cannas S, et al. Usefulness of the QuantiFERON-TB-Gold in tube in a population at risk of bovine tubercular infection. *Acta Microbiol Immunol Hung*. 2009 Dec;56(4):369-73. PMID: 20038488. Exclusion Code: X2
 166. Nakamura H, Tateyama M, Tasato D, et al. Active tuberculosis in patients undergoing hemodialysis for end-stage renal disease: a 9-year retrospective analysis in a single center. *Intern Med*. 2009;48(24):2061-7. PMID: 20009393. Exclusion Code: X2
 167. Kim EY, Lim JE, Jung JY, et al. Performance of the tuberculin skin test and interferon-gamma release assay for detection of tuberculosis infection in immunocompromised patients in a BCG-vaccinated population. *BMC Infect Dis*. 2009;9:207. PMID: 20003535. Exclusion Code: X2
 168. Kifai EJ, Bakari M. Mantoux skin test reactivity among household contacts of HIV-infected and HIV un-infected patients with sputum smear positive TB in Dar es Salaam, Tanzania. *East Afr J Public Health*. 2009 Aug;6(2):211-8. PMID: 20000032. Exclusion Code: X2
 169. Kang JS, Cherian A, Gan SH, et al. Strong purified protein derivative responses are associated with poor mycobacterium inhibition in latent TB. *Eur Respir J*. 2010 Aug;36(2):348-54. PMID: 19996195. Exclusion Code: X2
 170. Gogus F, Gunendi Z, Karakus R, et al. Comparison of tuberculin skin test and QuantiFERON-TB gold in tube test in patients with chronic inflammatory diseases living in a tuberculosis endemic population. *Clin Exp Med*. 2010 Sep;10(3):173-7. PMID: 19949831. Exclusion Code: X2
 171. Babu S, Bhat SQ, Kumar NP, et al. Regulatory T cells modulate Th17 responses in patients with positive tuberculin skin test results. *J Infect Dis*. 2010 Jan 1;201(1):20-31. PMID: 19929695. Exclusion Code: X2
 172. Li L, Mahan CS, Palaci M, et al. Sputum *Mycobacterium tuberculosis* mRNA as a marker of bacteriologic clearance in response to antituberculosis therapy. *J Clin Microbiol*. 2010 Jan;48(1):46-51. PMID: 19923475. Exclusion Code: X3
 173. Seyhan EC, Sokucu S, Altin S, et al. Comparison of the QuantiFERON-TB Gold In-Tube test with the tuberculin skin test for detecting latent tuberculosis infection in hemodialysis patients. *Transpl Infect Dis*. 2010 Apr;12(2):98-105. PMID: 19903322. Exclusion Code: X2
 174. Lee SW, Jang YS, Park CM, et al. The role of chest CT scanning in TB outbreak investigation. *Chest*. 2010 May;137(5):1057-64. PMID: 19880906. Exclusion Code: X2
 175. Babu K, Satish V, Satish S, et al. Utility of QuantiFERON TB gold test in a south Indian patient population of ocular inflammation. *Indian J Ophthalmol*. 2009 Nov-Dec;57(6):427-30. PMID: 19861743. Exclusion Code: X2
 176. Shovman O, Anouk M, Vinnitsky N, et al. QuantiFERON-TB Gold in the identification of latent tuberculosis infection in rheumatoid

Appendix C. Excluded Studies

- arthritis: a pilot study. *Int J Tuberc Lung Dis*. 2009 Nov;13(11):1427-32. PMID: 19861018. Exclusion Code: X2
177. Kobashi Y, Mouri K, Yagi S, et al. Clinical evaluation of the QuantiFERON-TB Gold test in patients with non-tuberculous mycobacterial disease. *Int J Tuberc Lung Dis*. 2009 Nov;13(11):1422-6. PMID: 19861017. Exclusion Code: X2
178. Zhao X, Mazlagic D, Flynn EA, et al. Is the QuantiFERON-TB blood assay a good replacement for the tuberculin skin test in tuberculosis screening? a pilot study at Berkshire Medical Center. *Am J Clin Pathol*. 2009 Nov;132(5):678-86. PMID: 19846807. Exclusion Code: X2
179. Kik SV, Franken WP, Mensen M, et al. Predictive value for progression to tuberculosis by IGRA and TST in immigrant contacts. *Eur Respir J*. 2010 Jun;35(6):1346-53. PMID: 19840963. Exclusion Code: X5
180. Hussain R, Talat N, Shahid F, et al. Biomarker changes associated with Tuberculin Skin Test (TST) conversion: a two-year longitudinal follow-up study in exposed household contacts. *PLoS One*. 2009;4(10):e7444. PMID: 19826490. Exclusion Code: X2
181. Ferrara G, Losi M, D'Amico R, et al. Interferon-gamma-release assays detect recent tuberculosis re-infection in elderly contacts. *Int J Immunopathol Pharmacol*. 2009 Jul-Sep;22(3):669-77. PMID: 19822083. Exclusion Code: X2
182. Cummings KJ, Smith TS, Shogren ES, et al. Prospective comparison of tuberculin skin test and QuantiFERON-TB Gold In-Tube assay for the detection of latent tuberculosis infection among healthcare workers in a low-incidence setting. *Infect Control Hosp Epidemiol*. 2009 Nov;30(11):1123-6. PMID: 19803719. Exclusion Code: X9
183. Shalabi NM, Houssen ME. Discrepancy between the tuberculin skin test and the levels of serum interferon-gamma in the diagnosis of tubercular infection in contacts. *Clin Biochem*. 2009 Nov;42(16-17):1596-601. PMID: 19732759. Exclusion Code: X9
184. Casas I, Latorre I, Esteve M, et al. Evaluation of interferon-gamma release assays in the diagnosis of recent tuberculosis infection in health care workers. *PLoS One*. 2009;4(8):e6686. PMID: 19701460. Exclusion Code: X2
185. Swindells JE, Aliyu SH, Enoch DA, et al. Role of interferon-gamma release assays in healthcare workers. *J Hosp Infect*. 2009 Oct;73(2):101-8. PMID: 19699551. Exclusion Code: X2
186. Park SH, Yang SK, Yang DH, et al. Prospective randomized trial of six-month versus nine-month therapy for intestinal tuberculosis. *Antimicrob Agents Chemother*. 2009 Oct;53(10):4167-71. PMID: 19667282. Exclusion Code: X2
187. Li J, Munsiff SS, Tarantino T, et al. Adherence to treatment of latent tuberculosis infection in a clinical population in New York City. *Int J Infect Dis*. 2010 Apr;14(4):e292-7. PMID: 19656705. Exclusion Code: X5
188. Chee CB, Lim LK, Barkham TM, et al. Use of a T cell interferon-gamma release assay to evaluate tuberculosis risk in newly qualified physicians in Singapore healthcare institutions. *Infect Control Hosp Epidemiol*. 2009 Sep;30(9):870-5. PMID: 19637958. Exclusion Code: X2
189. Kalantri Y, Hemvani N, Chitnis DS. Evaluation of whole blood IFN-gamma test using PPD and recombinant antigen challenge for diagnosis of pulmonary and extra-pulmonary tuberculosis. *Indian J Exp Biol*. 2009 Jun;47(6):463-8. PMID: 19634712. Exclusion Code: X9
190. Kobashi Y, Shimizu H, Ohue Y, et al. False negative results of QuantiFERON TB-2G test in patients with active tuberculosis. *Jpn J Infect Dis*. 2009 Jul;62(4):300-2. PMID: 19628910. Exclusion Code: X9
191. Alvarez-Leon EE, Espinosa-Vega E, Santana-Rodriguez E, et al. Screening for tuberculosis infection in spanish healthcare workers: Comparison of the QuantiFERON-TB gold in-tube test with the tuberculin skin test. *Infect Control Hosp Epidemiol*. 2009 Sep;30(9):876-83. PMID: 19614541. Exclusion Code: X2
192. Dheda K, van Zyl-Smit RN, Meldau R, et al. Quantitative lung T cell responses aid the rapid diagnosis of pulmonary tuberculosis. *Thorax*. 2009 Oct;64(10):847-53. PMID: 19592392. Exclusion Code: X2
193. Ang M, Htoon HM, Chee SP. Diagnosis of tuberculous uveitis: clinical application of an interferon-gamma release assay. *Ophthalmology*. 2009 Jul;116(7):1391-6. PMID: 19576501. Exclusion Code: X4
194. Detjen AK, Loeberberg L, Grewal HM, et al. Short-term reproducibility of a commercial interferon gamma release assay. *Clin Vaccine Immunol*. 2009 Aug;16(8):1170-5. PMID: 19535542. Exclusion Code: X2
195. Tsiouri G, Gaitanis G, Kiorpelidou D, et al. Tuberculin skin test overestimates tuberculosis hypersensitivity in adult patients with psoriasis. *Dermatology*. 2009;219(2):119-25. PMID: 19478478. Exclusion Code: X2

Appendix C. Excluded Studies

196. Bartacek A, Schutt D, Panosch B, et al. Comparison of a four-drug fixed-dose combination regimen with a single tablet regimen in smear-positive pulmonary tuberculosis. *Int J Tuberc Lung Dis*. 2009 Jun;13(6):760-6. PMID: 19460254. Exclusion Code: X2
197. Khanna P, Nikolayevskyy V, Warburton F, et al. Rate of latent tuberculosis infection detected by occupational health screening of nurses new to a london teaching hospital. *Infect Control Hosp Epidemiol*. 2009 Jun;30(6):581-4. PMID: 19415970. Exclusion Code: X2
198. Dheda K, van Zyl-Smit RN, Sechi LA, et al. Utility of quantitative T-cell responses versus unstimulated interferon- γ for the diagnosis of pleural tuberculosis. *Eur Respir J*. 2009 Nov;34(5):1118-26. PMID: 19386693. Exclusion Code: X2
199. Baker CA, Thomas W, Stauffer WM, et al. Serial testing of refugees for latent tuberculosis using the QuantiFERON-gold in-tube: effects of an antecedent tuberculin skin test. *Am J Trop Med Hyg*. 2009 Apr;80(4):628-33. PMID: 19346390. Exclusion Code: X5
200. Fu R, Wang C, Shi C, et al. An improved whole-blood gamma interferon assay based on the CFP21-MPT64 fusion protein. *Clin Vaccine Immunol*. 2009 May;16(5):686-91. PMID: 19279170. Exclusion Code: X2
201. Hess K, Goad J, Wu J, et al. Isoniazid completion rates for latent tuberculosis infection among college students managed by a community pharmacist. *J Am Coll Health*. 2009 Mar-Apr;57(5):553-5. PMID: 19254898. Exclusion Code: X7
202. Meintjes G, Rangaka MX, Maartens G, et al. Novel relationship between tuberculosis immune reconstitution inflammatory syndrome and antitubercular drug resistance. *Clin Infect Dis*. 2009 Mar 1;48(5):667-76. PMID: 19191655. Exclusion Code: X2
203. Lee SS, Chou KJ, Su IJ, et al. High prevalence of latent tuberculosis infection in patients in end-stage renal disease on hemodialysis: Comparison of QuantiFERON-TB GOLD, ELISPOT, and tuberculin skin test. *Infection*. 2009 Apr;37(2):96-102. PMID: 19139810. Exclusion Code: X2
204. Kobashi Y, Sugiu T, Mouri K, et al. Indeterminate results of QuantiFERON TB-2G test performed in routine clinical practice. *Eur Respir J*. 2009 Apr;33(4):812-5. PMID: 19129287. Exclusion Code: X2
205. Sahni R, Miranda C, Yen-Lieberman B, et al. Does the implementation of an interferon-gamma release assay in lieu of a tuberculin skin test increase acceptance of preventive therapy for latent tuberculosis among healthcare workers? *Infect Control Hosp Epidemiol*. 2009 Feb;30(2):197-9. PMID: 19119939. Exclusion Code: X5
206. Sagheb MM, Goodarzi M, Roozbeh J. The booster phenomenon of tuberculin skin testing in patients receiving hemodialysis. *Iran J Immunol*. 2008 Dec;5(4):212-6. PMID: 19098365. Exclusion Code: X5
207. Araujo Z, Giampietro F, Cancado LC, et al. Comparison of serological responses in two different populations with pulmonary tuberculosis. *Mem Inst Oswaldo Cruz*. 2008 Nov;103(7):661-7. PMID: 19057815. Exclusion Code: X3
208. Diel R, Loddenkemper R, Meywald-Walter K, et al. Comparative performance of tuberculin skin test, QuantiFERON-TB-Gold In Tube assay, and T-SPOT.TB test in contact investigations for tuberculosis. *Chest*. 2009 Apr;135(4):1010-8. PMID: 19017873. Exclusion Code: X2
209. Golub JE, Astemborski J, Ahmed M, et al. Long-term effectiveness of diagnosing and treating latent tuberculosis infection in a cohort of HIV-infected and at-risk injection drug users. *J Acquir Immune Defic Syndr*. 2008 Dec 15;49(5):532-7. PMID: 18989223. Exclusion Code: X7
210. Janssens JP, Roux-Lombard P, Perneger T, et al. Contribution of a IFN- γ assay in contact tracing for tuberculosis in a low-incidence, high immigration area. *Swiss Med Wkly*. 2008 Oct 4;138(39-40):585-93. PMID: 18853288. Exclusion Code: X5
211. Anderson BL, Welch RJ, Litwin CM. Assessment of three commercially available serologic assays for detection of antibodies to *Mycobacterium tuberculosis* and identification of active tuberculosis. *Clin Vaccine Immunol*. 2008 Nov;15(11):1644-9. PMID: 18827190. Exclusion Code: X3
212. Ruhwald M, Bodmer T, Maier C, et al. Evaluating the potential of IP-10 and MCP-2 as biomarkers for the diagnosis of tuberculosis. *Eur Respir J*. 2008 Dec;32(6):1607-15. PMID: 18684849. Exclusion Code: X2
213. Chegou NN, Walzl G, Bolliger CT, et al. Evaluation of adapted whole-blood interferon-gamma release assays for the diagnosis of pleural tuberculosis. *Respiration*. 2008;76(2):131-8. PMID: 18434705. Exclusion Code: X2
214. Chang KC, Leung CC, Yew WW, et al. Hepatotoxicity of pyrazinamide: cohort and case-control analyses. *Am J Respir Crit Care Med*.

Appendix C. Excluded Studies

- 2008 Jun 15;177(12):1391-6. PMID: 18388355. Exclusion Code: X2
215. Carvalho AC, Crotti N, Crippa M, et al. QuantiFERON-TB Gold test for healthcare workers. *J Hosp Infect.* 2008 May;69(1):91-2. PMID: 18358562. Exclusion Code: X5
216. Dosanjh DP, Hinks TS, Innes JA, et al. Improved diagnostic evaluation of suspected tuberculosis. *Ann Intern Med.* 2008 Mar 4;148(5):325-36. PMID: 18316751. Exclusion Code: X2
217. Eum SY, Lee YJ, Kwak HK, et al. Evaluation of the diagnostic utility of a whole-blood interferon-gamma assay for determining the risk of exposure to *Mycobacterium tuberculosis* in Bacille Calmette-Guerin (BCG)-vaccinated individuals. *Diagn Microbiol Infect Dis.* 2008 Jun;61(2):181-6. PMID: 18296002. Exclusion Code: X9
218. Diel R, Loddenkemper R, Meywald-Walter K, et al. Predictive value of a whole blood IFN-gamma assay for the development of active tuberculosis disease after recent infection with *Mycobacterium tuberculosis*. *Am J Respir Crit Care Med.* 2008 May 15;177(10):1164-70. PMID: 18276940. Exclusion Code: X2
219. Adams LV, Waddell RD, Von Reyn CF. T-SPOT.TB Test(R) results in adults with *Mycobacterium avium* complex pulmonary disease. *Scand J Infect Dis.* 2008;40(3):196-203. PMID: 18274952. Exclusion Code: X2
220. Haley CA, Stephan S, Vossell LF, et al. Successful use of rifampicin for Hispanic foreign-born patients with latent tuberculosis infection. *Int J Tuberc Lung Dis.* 2008 Feb;12(2):160-7. PMID: 18230248. Exclusion Code: X4
221. Perry S, Sanchez L, Yang S, et al. Reproducibility of QuantiFERON-TB gold in-tube assay. *Clin Vaccine Immunol.* 2008 Mar;15(3):425-32. PMID: 18199741. Exclusion Code: X2
222. Beffa P, Zellweger A, Janssens JP, et al. Indeterminate test results of T-SPOT.TB performed under routine field conditions. *Eur Respir J.* 2008 Apr;31(4):842-6. PMID: 18057053. Exclusion Code: X2
223. Palazzo R, Spensieri F, Massari M, et al. Use of whole-blood samples in in-house bulk and single-cell antigen-specific gamma interferon assays for surveillance of *Mycobacterium tuberculosis* infections. *Clin Vaccine Immunol.* 2008 Feb;15(2):327-37. PMID: 18032595. Exclusion Code: X9
224. Choi CM, Hwang SS, Lee CH, et al. Latent tuberculosis infection in a military setting diagnosed by whole-blood interferon-gamma assay. *Respirology.* 2007 Nov;12(6):898-901. PMID: 17986121. Exclusion Code: X2
225. Dominguez J, Ruiz-Manzano J, De Souza-Galvao M, et al. Comparison of two commercially available gamma interferon blood tests for immunodiagnosis of tuberculosis. *Clin Vaccine Immunol.* 2008 Jan;15(1):168-71. PMID: 17978008. Exclusion Code: X2
226. Veesser PI, Smith PK, Handy B, et al. Tuberculosis screening on a health science campus: use of QuantiFERON-TB Gold Test for students and employees. *J Am Coll Health.* 2007 Sep-Oct;56(2):175-80. PMID: 17967764. Exclusion Code: X4
227. Smith-Rohrberg D, Sharma SK. Tuberculin skin test among pulmonary sarcoidosis patients with and without tuberculosis: its utility for the screening of the two conditions in tuberculosis-endemic regions. *Sarcoidosis Vasc Diffuse Lung Dis.* 2006 Jun;23(2):130-4. PMID: 17937109. Exclusion Code: X2
228. Falagas ME, Voidonikola PT, Angelousi AG. Tuberculosis in patients with systemic rheumatic or pulmonary diseases treated with glucocorticosteroids and the preventive role of isoniazid: a review of the available evidence. *Int J Antimicrob Agents.* 2007 Dec;30(6):477-86. PMID: 17913470. Exclusion Code: X2
229. Markov M, Patel K, Raeesy A, et al. Liver and pancreatic injury induced by antituberculous therapy. *Dig Dis Sci.* 2007 Nov;52(11):3275-81. PMID: 17909976. Exclusion Code: X7
230. Chan-Yeung M, Dai DL, Cheung AH, et al. Tuberculin skin test reaction and body mass index in old age home residents in Hong Kong. *J Am Geriatr Soc.* 2007 Oct;55(10):1592-7. PMID: 17908061. Exclusion Code: X4
231. Kobashi Y, Obase Y, Fukuda M, et al. Usefulness of QuantiFERON TB-2G, a diagnostic method for latent tuberculosis infection, in a contact investigation of health care workers. *Intern Med.* 2007;46(18):1543-9. PMID: 17878640. Exclusion Code: X2
232. Levy MH, Butler TG, Zhou J. Prevalence of Mantoux positivity and annual risk of infection for tuberculosis in New South Wales prisoners, 1996 and 2001. *N S W Public Health Bull.* 2007 Jul-Aug;18(7-8):119-24. PMID: 17854540. Exclusion Code: X3
233. Mori T, Harada N, Higuchi K, et al. Waning of the specific interferon-gamma response after years of tuberculosis infection. *Int J Tuberc Lung Dis.* 2007 Sep;11(9):1021-5. PMID: 17705982. Exclusion Code: X2
234. Moran-Mendoza O, Marion SA, Elwood K, et al. Tuberculin skin test size and risk of tuberculosis

Appendix C. Excluded Studies

- development: a large population-based study in contacts. *Int J Tuberc Lung Dis*. 2007 Sep;11(9):1014-20. PMID: 17705981. Exclusion Code: X2
235. Passalent L, Khan K, Richardson R, et al. Detecting latent tuberculosis infection in hemodialysis patients: a head-to-head comparison of the T-SPOT.TB test, tuberculin skin test, and an expert physician panel. *Clin J Am Soc Nephrol*. 2007 Jan;2(1):68-73. PMID: 17699389. Exclusion Code: X4
 236. Kobashi Y, Mouri K, Obase Y, et al. Clinical evaluation of QuantiFERON TB-2G test for immunocompromised patients. *Eur Respir J*. 2007 Nov;30(5):945-50. PMID: 17652312. Exclusion Code: X2
 237. Matulis G, Juni P, Villiger PM, et al. Detection of latent tuberculosis in immunosuppressed patients with autoimmune diseases: performance of a Mycobacterium tuberculosis antigen-specific interferon gamma assay. *Ann Rheum Dis*. 2008 Jan;67(1):84-90. PMID: 17644549. Exclusion Code: X2
 238. Igari H, Watanabe A, Sato T. Booster phenomenon of QuantiFERON-TB Gold after prior intradermal PPD injection. *Int J Tuberc Lung Dis*. 2007 Jul;11(7):788-91. PMID: 17609055. Exclusion Code: X2
 239. Nienhaus A, Schablon A, Bacle CL, et al. Evaluation of the interferon-gamma release assay in healthcare workers. *Int Arch Occup Environ Health*. 2008 Jan;81(3):295-300. PMID: 17605033. Exclusion Code: X2
 240. Pai M, O'Brien R. Serial testing for tuberculosis: can we make sense of T cell assay conversions and reversions? *PLoS Med*. 2007 Jun;4(6):e208. PMID: 17564491. Exclusion Code: X1
 241. Hill PC, Brookes RH, Fox A, et al. Longitudinal assessment of an ELISPOT test for Mycobacterium tuberculosis infection. *PLoS Med*. 2007 Jun;4(6):e192. PMID: 17564487. Exclusion Code: X2
 242. Gomez LM, Sanchez E, Ruiz-Narvaez EA, et al. Macrophage migration inhibitory factor gene influences the risk of developing tuberculosis in northwestern Colombian population. *Tissue Antigens*. 2007 Jul;70(1):28-33. PMID: 17559578. Exclusion Code: X5
 243. Wang JY, Chou CH, Lee LN, et al. Diagnosis of tuberculosis by an enzyme-linked immunospot assay for interferon-gamma. *Emerg Infect Dis*. 2007 Apr;13(4):553-8. PMID: 17553269. Exclusion Code: X2
 244. Cho YJ, Lee SM, Yoo CG, et al. Clinical characteristics of tuberculosis in patients with liver cirrhosis. *Respirology*. 2007 May;12(3):401-5. PMID: 17539845. Exclusion Code: X3
 245. Ozdemir D, Annakkaya AN, Tarhan G, et al. Comparison of the tuberculin skin test and the quantiferon test for latent Mycobacterium tuberculosis infections in health care workers in Turkey. *Jpn J Infect Dis*. 2007 May;60(2-3):102-5. PMID: 17515641. Exclusion Code: X2
 246. Leyten EM, Arend SM, Prins C, et al. Discrepancy between Mycobacterium tuberculosis-specific gamma interferon release assays using short and prolonged in vitro incubation. *Clin Vaccine Immunol*. 2007 Jul;14(7):880-5. PMID: 17507543. Exclusion Code: X2
 247. Kang YA, Lee HW, Hwang SS, et al. Usefulness of whole-blood interferon-gamma assay and interferon-gamma enzyme-linked immunospot assay in the diagnosis of active pulmonary tuberculosis. *Chest*. 2007 Sep;132(3):959-65. PMID: 17505029. Exclusion Code: X9
 248. Tuuminen T, Sorva S, Liippo K, et al. Feasibility of commercial interferon-gamma-based methods for the diagnosis of latent Mycobacterium tuberculosis infection in Finland, a country of low incidence and high bacille Calmette-Guerin vaccination coverage. *Clin Microbiol Infect*. 2007 Aug;13(8):836-8. PMID: 17501976. Exclusion Code: X2
 249. Porsa E, Cheng L, Graviss EA. Comparison of an ESAT-6/CFP-10 peptide-based enzyme-linked immunospot assay to a tuberculin skin test for screening of a population at moderate risk of contracting tuberculosis. *Clin Vaccine Immunol*. 2007 Jun;14(6):714-9. PMID: 17442844. Exclusion Code: X2
 250. Gazetta CE, Ruffino-Netto A, Pinto Neto JM, et al. Investigation of tuberculosis contacts in the tuberculosis control program of a medium-sized municipality in the southeast of Brazil in 2002. *J Bras Pneumol*. 2006 Nov-Dec;32(6):559-65. PMID: 17435907. Exclusion Code: X4
 251. Gustafson P, Lisse I, Gomes V, et al. Risk factors for positive tuberculin skin test in Guinea-Bissau. *Epidemiology*. 2007 May;18(3):340-7. PMID: 17435442. Exclusion Code: X2
 252. Salles CG, Ruffino-Netto A, Lapa-e-Silva JR, et al. The presence of a booster phenomenon among contacts of active pulmonary tuberculosis cases: a retrospective cohort. *BMC Public Health*. 2007;7:38. PMID: 17371600. Exclusion Code: X2
 253. Kwon YS, Koh WJ, Suh GY, et al. Hepatitis C virus infection and hepatotoxicity during antituberculosis chemotherapy. *Chest*. 2007

Appendix C. Excluded Studies

- Mar;131(3):803-8. PMID: 17356096. Exclusion Code: X2
254. Franken WP, Timmermans JF, Prins C, et al. Comparison of Mantoux and QuantiFERON TB Gold tests for diagnosis of latent tuberculosis infection in Army personnel. *Clin Vaccine Immunol.* 2007 Apr;14(4):477-80. PMID: 17301213. Exclusion Code: X9
255. Jung SW, Jeon SW, Do BH, et al. Clinical aspects of rifampicin-associated pseudomembranous colitis. *J Clin Gastroenterol.* 2007 Jan;41(1):38-40. PMID: 17198063. Exclusion Code: X2
256. Gooding S, Chowdhury O, Hinks T, et al. Impact of a T cell-based blood test for tuberculosis infection on clinical decision-making in routine practice. *J Infect.* 2007 Mar;54(3):e169-74. PMID: 17188363. Exclusion Code: X2
257. Hill PC, Brookes RH, Fox A, et al. Surprisingly high specificity of the PPD skin test for M. tuberculosis infection from recent exposure in The Gambia. *PLoS One.* 2006;1:e68. PMID: 17183699. Exclusion Code: X2
258. Arend SM, Thijsen SF, Leyten EM, et al. Comparison of two interferon-gamma assays and tuberculin skin test for tracing tuberculosis contacts. *Am J Respir Crit Care Med.* 2007 Mar 15;175(6):618-27. PMID: 17170386. Exclusion Code: X2
259. Choi CM, Kang CI, Kim DH, et al. The role of TST in the diagnosis of latent tuberculosis infection among military personnel in South Korea. *Int J Tuberc Lung Dis.* 2006 Dec;10(12):1342-6. PMID: 17167950. Exclusion Code: X2
260. Lardizabal A, Passannante M, Kojakali F, et al. Enhancement of treatment completion for latent tuberculosis infection with 4 months of rifampin. *Chest.* 2006 Dec;130(6):1712-7. PMID: 17166986. Exclusion Code: X7
261. Panickar JR, Hoskyns W. Treatment failure in tuberculosis. *Eur Respir J.* 2007 Mar;29(3):561-4. PMID: 17135229. Exclusion Code: X2
262. Page KR, Sifakis F, Montes de Oca R, et al. Improved adherence and less toxicity with rifampin vs isoniazid for treatment of latent tuberculosis: a retrospective study. *Arch Intern Med.* 2006 Sep 25;166(17):1863-70. PMID: 17000943. Exclusion Code: X7
263. Huizinga TW, Arend SM. Is the tuberculin skin test an accurate method of detecting tuberculosis in patients with rheumatoid arthritis? *Nat Clin Pract Rheumatol.* 2006 Apr;2(4):188-9. PMID: 16932683. Exclusion Code: X1
264. Tuberculin skin testing of close contacts: recent or long-standing infection? *Can Commun Dis Rep.* 2006 Jun 15;32(12):133-40. PMID: 16805063. Exclusion Code: X2
265. Cook PP, Maldonado RA, Yarnell CT, et al. Safety and completion rate of short-course therapy for treatment of latent tuberculosis infection. *Clin Infect Dis.* 2006 Aug 1;43(3):271-5. PMID: 16804838. Exclusion Code: X4
266. Pai M, Joshi R, Dogra S, et al. Serial testing of health care workers for tuberculosis using interferon-gamma assay. *Am J Respir Crit Care Med.* 2006 Aug 1;174(3):349-55. PMID: 16690977. Exclusion Code: X2
267. Gomez LM, Anaya JM, Martin J. Genetic influence of PTPN22 R620W polymorphism in tuberculosis. *Hum Immunol.* 2005 Dec;66(12):1242-7. PMID: 16690411. Exclusion Code: X5
268. Frenzel EC, Thomas GA, Hanna HA. The importance of two-step tuberculin skin testing for newly employed healthcare workers. *Infect Control Hosp Epidemiol.* 2006 May;27(5):512-4. PMID: 16671035. Exclusion Code: X2
269. Harada N, Nakajima Y, Higuchi K, et al. Screening for tuberculosis infection using whole-blood interferon-gamma and Mantoux testing among Japanese healthcare workers. *Infect Control Hosp Epidemiol.* 2006 May;27(5):442-8. PMID: 16671023. Exclusion Code: X2
270. Shah SS, McGowan JP, Klein RS, et al. Agreement between Mantoux skin testing and QuantiFERON-TB assay using dual mycobacterial antigens in current and former injection drug users. *Med Sci Monit.* 2006 Apr;12(4):Mt11-6. PMID: 16572060. Exclusion Code: X2
271. Loh LC, Chan SK, Ch'ng KI, et al. Influence of co-morbidity in the interpretation of tuberculin skin reactivity in multi-ethnic adult patients with tuberculosis. *Med J Malaysia.* 2005 Oct;60(4):426-31. PMID: 16570703. Exclusion Code: X2
272. Mahomed H, Hughes EJ, Hawkridge T, et al. Comparison of mantoux skin test with three generations of a whole blood IFN-gamma assay for tuberculosis infection. *Int J Tuberc Lung Dis.* 2006 Mar;10(3):310-6. PMID: 16562712. Exclusion Code: X2
273. Dewan PK, Grinsdale J, Liska S, et al. Feasibility, acceptability, and cost of tuberculosis testing by whole-blood interferon-gamma assay. *BMC Infect Dis.* 2006;6:47. PMID: 16539718. Exclusion Code: X4
274. Abrahao RM, Nogueira PA, Malucelli MI. Tuberculosis in county jail prisoners in the western sector of the city of Sao Paulo, Brazil. *Int*

Appendix C. Excluded Studies

- J Tuberc Lung Dis. 2006 Feb;10(2):203-8. PMID: 16499262. Exclusion Code: X5
275. Jeffries DJ, Hill PC, Fox A, et al. Identifying ELISPOT and skin test cut-offs for diagnosis of Mycobacterium tuberculosis infection in The Gambia. *Int J Tuberc Lung Dis*. 2006 Feb;10(2):192-8. PMID: 16499260. Exclusion Code: X2
 276. Schechter M, Zajdenverg R, Falco G, et al. Weekly rifapentine/isoniazid or daily rifampin/pyrazinamide for latent tuberculosis in household contacts. *Am J Respir Crit Care Med*. 2006 Apr 15;173(8):922-6. PMID: 16474028. Exclusion Code: X4
 277. Anlar FY, Kabasakal E, Karsi R. Tuberculosis and atopy: a study in an endemic area. *Respir Med*. 2006 Sep;100(9):1647-50. PMID: 16469489. Exclusion Code: X3
 278. Menzies D, Dion MJ, Francis D, et al. In closely monitored patients, adherence in the first month predicts completion of therapy for latent tuberculosis infection. *Int J Tuberc Lung Dis*. 2005 Dec;9(12):1343-8. PMID: 16466056. Exclusion Code: X5
 279. Roth VR, Garrett DO, Laserson KF, et al. A multicenter evaluation of tuberculin skin test positivity and conversion among health care workers in Brazilian hospitals. *Int J Tuberc Lung Dis*. 2005 Dec;9(12):1335-42. PMID: 16466055. Exclusion Code: X5
 280. Jacono KM, O'Riordan MA, Furman L. Can we improve the return rate for tuberculin skin test readings? *Arch Pediatr Adolesc Med*. 2006 Jan;160(1):106. PMID: 16389222. Exclusion Code: X2
 281. Wilkinson KA, Kon OM, Newton SM, et al. Effect of treatment of latent tuberculosis infection on the T cell response to Mycobacterium tuberculosis antigens. *J Infect Dis*. 2006 Feb 1;193(3):354-9. PMID: 16388482. Exclusion Code: X5
 282. Schlesinger N, Lardizabal A, Rao J, et al. Tuberculosis of the spine: experience in an inner city hospital. *J Clin Rheumatol*. 2005 Feb;11(1):17-20. PMID: 16357692. Exclusion Code: X7
 283. Codecasa LR, Ferrarese M, Penati V, et al. Comparison of tuberculin skin test and Quantiferon immunological assay for latent tuberculosis infection. *Monaldi Arch Chest Dis*. 2005 Sep;63(3):158-62. PMID: 16312206. Exclusion Code: X2
 284. Jawahar MS, Rajaram K, Sivasubramanian S, et al. Treatment of lymph node tuberculosis--a randomized clinical trial of two 6-month regimens. *Trop Med Int Health*. 2005 Nov;10(11):1090-8. PMID: 16262733. Exclusion Code: X2
 285. Radhakrishna S, Frieden TR, Subramani R, et al. Value of dual testing for identifying tuberculous infection. *Tuberculosis (Edinb)*. 2006 Jan;86(1):47-53. PMID: 16256435. Exclusion Code: X2
 286. White MC, Tulskey JP, Menendez E, et al. Incidence of TB in inmates with latent TB infection: 5-year follow-up. *Am J Prev Med*. 2005 Nov;29(4):295-301. PMID: 16242592. Exclusion Code: X3
 287. Eleftheriadis T, Tsiaga P, Antoniadis G, et al. The value of serum antilipoarabinomannan antibody detection in the diagnosis of latent tuberculosis in hemodialysis patients. *Am J Kidney Dis*. 2005 Oct;46(4):706-12. PMID: 16183426. Exclusion Code: X2
 288. Richards B, Kozak R, Brassard P, et al. Tuberculosis surveillance among new immigrants in Montreal. *Int J Tuberc Lung Dis*. 2005 Aug;9(8):858-64. PMID: 16104631. Exclusion Code: X5
 289. Kimura M, Comstock GW, Mori T. Comparison of erythema and induration as results of tuberculin tests. *Int J Tuberc Lung Dis*. 2005 Aug;9(8):853-7. PMID: 16104630. Exclusion Code: X3
 290. Ponce de Leon D, Acevedo-Vasquez E, Sanchez-Torres A, et al. Attenuated response to purified protein derivative in patients with rheumatoid arthritis: study in a population with a high prevalence of tuberculosis. *Ann Rheum Dis*. 2005 Sep;64(9):1360-1. PMID: 16100342. Exclusion Code: X6
 291. Fountain FF, Tolley E, Chrisman CR, et al. Isoniazid hepatotoxicity associated with treatment of latent tuberculosis infection: a 7-year evaluation from a public health tuberculosis clinic. *Chest*. 2005 Jul;128(1):116-23. PMID: 16002924. Exclusion Code: X4
 292. Britton WJ, Gilbert GL, Wheatley J, et al. Sensitivity of human gamma interferon assay and tuberculin skin testing for detecting infection with Mycobacterium tuberculosis in patients with culture positive tuberculosis. *Tuberculosis (Edinb)*. 2005 May;85(3):137-45. PMID: 15850752. Exclusion Code: X2
 293. Lobato MN, Reves RR, Jasmer RM, et al. Adverse events and treatment completion for latent tuberculosis in jail inmates and homeless persons. *Chest*. 2005 Apr;127(4):1296-303. PMID: 15821208. Exclusion Code: X3
 294. Dheda K, Udawadia ZF, Huggett JF, et al. Utility of the antigen-specific interferon-gamma assay for the management of tuberculosis. *Curr Opin*

Appendix C. Excluded Studies

- Pulm Med. 2005 May;11(3):195-202. PMID: 15818179. Exclusion Code: X1
295. Deniz O, Tozkoparan E, Yonem A, et al. Low parathormone levels and hypercalcaemia in patients with pulmonary tuberculosis: relation to radiological extent of disease and tuberculin skin test. *Int J Tuberc Lung Dis*. 2005 Mar;9(3):317-21. PMID: 15786897. Exclusion Code: X5
 296. Tortajada C, Martinez-Lacasa J, Sanchez F, et al. Is the combination of pyrazinamide plus rifampicin safe for treating latent tuberculosis infection in persons not infected by the human immunodeficiency virus? *Int J Tuberc Lung Dis*. 2005 Mar;9(3):276-81. PMID: 15786890. Exclusion Code: X4
 297. Priest DH, Vossell LF, Jr., Sherfy EA, et al. Use of intermittent rifampin and pyrazinamide therapy for latent tuberculosis infection in a targeted tuberculin testing program. *Clin Infect Dis*. 2004 Dec 15;39(12):1764-71. PMID: 15578397. Exclusion Code: X3
 298. van Hest R, Baars H, Kik S, et al. Hepatotoxicity of rifampin-pyrazinamide and isoniazid preventive therapy and tuberculosis treatment. *Clin Infect Dis*. 2004 Aug 15;39(4):488-96. PMID: 15356811. Exclusion Code: X4
 299. Wauters A, Peetermans WE, Van den Brande P, et al. The value of tuberculin skin testing in haemodialysis patients. *Nephrol Dial Transplant*. 2004 Feb;19(2):433-8. PMID: 14736970. Exclusion Code: X5
 300. Chau CH, Yew WW, Chan CK. Familial clustering of rifampin-induced acute renal failure. *Int J Tuberc Lung Dis*. 2003 Dec;7(12):1210. PMID: 14677899. Exclusion Code: X5
 301. Leung CC, Law WS, Chang KC, et al. Initial experience on rifampin and pyrazinamide vs isoniazid in the treatment of latent tuberculosis infection among patients with silicosis in Hong Kong. *Chest*. 2003 Dec;124(6):2112-8. PMID: 14665488. Exclusion Code: X4
 302. Al-Moamary MS, Al-Baz S, Alothman A, et al. Does tuberculin skin test predict tuberculosis in patients with end-stage liver disease? *Saudi Med J*. 2003 Nov;24(11):1269-70. PMID: 14647571. Exclusion Code: X3
 303. Mitchison DA. Role of isoniazid in once-weekly rifapentine treatment of pulmonary tuberculosis. *Am J Respir Crit Care Med*. 2003 May 15;167(10):1298-9. PMID: 12738593. Exclusion Code: X1
 304. Bugiani M, Borraccino A, Migliore E, et al. Tuberculin reactivity in adult BCG-vaccinated subjects: a cross-sectional study. *Int J Tuberc Lung Dis*. 2003 Apr;7(4):320-6. PMID: 12729336. Exclusion Code: X2
 305. McNeill L, Allen M, Estrada C, et al. Pyrazinamide and rifampin vs isoniazid for the treatment of latent tuberculosis: improved completion rates but more hepatotoxicity. *Chest*. 2003 Jan;123(1):102-6. PMID: 12527609. Exclusion Code: X4
 306. Lee AM, Mennone JZ, Jones RC, et al. Risk factors for hepatotoxicity associated with rifampin and pyrazinamide for the treatment of latent tuberculosis infection: experience from three public health tuberculosis clinics. *Int J Tuberc Lung Dis*. 2002 Nov;6(11):995-1000. PMID: 12475146. Exclusion Code: X4
 307. Benito N, Sued O, Moreno A, et al. Diagnosis and treatment of latent tuberculosis infection in liver transplant recipients in an endemic area. *Transplantation*. 2002 Nov 27;74(10):1381-6. PMID: 12451235. Exclusion Code: X3
 308. Stout JE, Engemann JJ, Cheng AC, et al. Safety of 2 months of rifampin and pyrazinamide for treatment of latent tuberculosis. *Am J Respir Crit Care Med*. 2003 Mar 15;167(6):824-7. PMID: 12446275. Exclusion Code: X3
 309. Howard AA, Klein RS, Schoenbaum EE, et al. Crack cocaine use and other risk factors for tuberculin positivity in drug users. *Clin Infect Dis*. 2002 Nov 15;35(10):1183-90. PMID: 12410478. Exclusion Code: X5
 310. Quaglio G, Lugoboni F, Talamini G, et al. Prevalence of tuberculosis infection and comparison of multiple-puncture liquid tuberculin test and Mantoux test among drug users. *Scand J Infect Dis*. 2002;34(8):574-6. PMID: 12238571. Exclusion Code: X2
 311. Panlilio AL, Burwen DR, Curtis AB, et al. Tuberculin skin testing surveillance of health care personnel. *Clin Infect Dis*. 2002 Aug 1;35(3):219-27. PMID: 12115085. Exclusion Code: X2
 312. Batki SL, Gruber VA, Bradley JM, et al. A controlled trial of methadone treatment combined with directly observed isoniazid for tuberculosis prevention in injection drug users. *Drug Alcohol Depend*. 2002 May 1;66(3):283-93. PMID: 12062463. Exclusion Code: X3
 313. Bock NN, Sterling TR, Hamilton CD, et al. A prospective, randomized, double-blind study of the tolerability of rifapentine 600, 900, and 1,200 mg plus isoniazid in the continuation phase of tuberculosis treatment. *Am J Respir Crit Care Med*. 2002 Jun 1;165(11):1526-30. PMID: 12045127. Exclusion Code: X2
 314. Noertjojo K, Tam CM, Chan SL, et al. Contact examination for tuberculosis in Hong Kong is

Appendix C. Excluded Studies

- useful. *Int J Tuberc Lung Dis*. 2002 Jan;6(1):19-24. PMID: 11931396. Exclusion Code: X5
315. Steinmann RA, Rickel MK. A 23-year-old with refractory seizures following an isoniazid overdose. *J Emerg Nurs*. 2002 Feb;28(1):7-10. PMID: 11830727. Exclusion Code: X7
 316. Rangel-Frausto MS, Ponce-De-Leon-Rosales S, Martinez-Abaroa C, et al. Tuberculosis and tuberculin quality: best intentions, misleading results. *Infect Control Hosp Epidemiol*. 2001 Aug;22(8):481-4. PMID: 11700874. Exclusion Code: X3
 317. Plitt SS, Soskolne CL, Fanning EA, et al. Prevalence and determinants of tuberculin reactivity among physicians in Edmonton, Canada: 1996-1997. *Int J Epidemiol*. 2001 Oct;30(5):1022-8. PMID: 11689515. Exclusion Code: X2
 318. Saigal S, Agarwal SR, Nandeesh HP, et al. Safety of an ofloxacin-based antitubercular regimen for the treatment of tuberculosis in patients with underlying chronic liver disease: a preliminary report. *J Gastroenterol Hepatol*. 2001 Sep;16(9):1028-32. PMID: 11595068. Exclusion Code: X2
 319. Al Zahrani K, Al Jahdali H, Poirier L, et al. Yield of smear, culture and amplification tests from repeated sputum induction for the diagnosis of pulmonary tuberculosis. *Int J Tuberc Lung Dis*. 2001 Sep;5(9):855-60. PMID: 11573898. Exclusion Code: X5
 320. Kanaya AM, Glidden DV, Chambers HF. Identifying pulmonary tuberculosis in patients with negative sputum smear results. *Chest*. 2001 Aug;120(2):349-55. PMID: 11502628. Exclusion Code: X2
 321. Klein RS, Gourevitch MN, Teeter R, et al. The incidence of tuberculosis in drug users with small tuberculin reaction sizes. *Int J Tuberc Lung Dis*. 2001 Aug;5(8):707-11. PMID: 11495260. Exclusion Code: X2
 322. Moreno S, Blazquez R, Novoa A, et al. The effect of BCG vaccination on tuberculin reactivity and the booster effect among hospital employees. *Arch Intern Med*. 2001 Jul 23;161(14):1760-5. PMID: 11485509. Exclusion Code: X4
 323. Fine PE, Floyd S, Stanford JL, et al. Environmental mycobacteria in northern Malawi: implications for the epidemiology of tuberculosis and leprosy. *Epidemiol Infect*. 2001 Jun;126(3):379-87. PMID: 11467795. Exclusion Code: X2
 324. Lalvani A, Pathan AA, Durkan H, et al. Enhanced contact tracing and spatial tracking of *Mycobacterium tuberculosis* infection by enumeration of antigen-specific T cells. *Lancet*. 2001 Jun 23;357(9273):2017-21. PMID: 11438135. Exclusion Code: X3
 325. Datta M, Radhamani MP, Sadacharam K, et al. Survey for tuberculosis in a tribal population in North Arcot District. *Int J Tuberc Lung Dis*. 2001 Mar;5(3):240-9. PMID: 11326823. Exclusion Code: X5
 326. Taskapan H, Oymak O, Utas C. Tuberculin and anergy testing in CAPD patients. *Perit Dial Int*. 2000 Nov-Dec;20(6):807-9. PMID: 11216589. Exclusion Code: X2
 327. El-Sadr WM, Perlman DC, Denning E, et al. A review of efficacy studies of 6-month short-course therapy for tuberculosis among patients infected with human immunodeficiency virus: differences in study outcomes. *Clin Infect Dis*. 2001 Feb 15;32(4):623-32. PMID: 11181127. Exclusion Code: X2
 328. Sherman RA, Shimoda KJ. Tuberculosis tracking: determining the frequency of the booster effect in patients and staff. *Am J Infect Control*. 2001 Feb;29(1):7-12. PMID: 11172312. Exclusion Code: X2
 329. Memish ZA, Mah MW, Mahmood SA, et al. Clinico-diagnostic experience with tuberculous lymphadenitis in Saudi Arabia. *Clin Microbiol Infect*. 2000 Mar;6(3):137-41. PMID: 11168089. Exclusion Code: X9
 330. Raut VS, Mahashur AA, Sheth SS. The Mantoux test in the diagnosis of genital tuberculosis in women. *Int J Gynaecol Obstet*. 2001 Feb;72(2):165-9. PMID: 11166750. Exclusion Code: X2
 331. Roselle GA, Danko LH, Kralovic SM, et al. Tuberculosis in the veterans healthcare system: a six-year review and evaluation of programme effectiveness. *Epidemiol Infect*. 2000 Oct;125(2):315-23. PMID: 11117955. Exclusion Code: X7
 332. Uyan AP, Baskin E, Buyukbese E, et al. Evaluating Bacillus-Calmette-Guerin vaccination by tuberculin skin test response. *Indian Pediatr*. 2000 Oct;37(10):1106-10. PMID: 11042711. Exclusion Code: X2
 333. Boggess KA, Myers ER, Hamilton CD. Antepartum or postpartum isoniazid treatment of latent tuberculosis infection. *Obstet Gynecol*. 2000 Nov;96(5 Pt 1):757-62. PMID: 11042314. Exclusion Code: X7
 334. Ramasamy R, Reginald A, Ganesan E. The use of high-dose isomazid in intermittent regime TB treatment--some preliminary findings. *Trop Doct*. 2000 Jan;30(1):56. PMID: 10842542. Exclusion Code: X3

Appendix C. Excluded Studies

335. Stuart RL, Wilson J, Grayson ML. Isoniazid toxicity in health care workers. *Clin Infect Dis*. 1999 Apr;28(4):895-7. PMID: 10825056. Exclusion Code: X4
336. Tam CM, Chan SL, Kam KM, et al. Rifapentine and isoniazid in the continuation phase of a 6-month regimen. Interim report: no activity of isoniazid in the continuation phase. *Int J Tuberc Lung Dis*. 2000 Mar;4(3):262-7. PMID: 10751074. Exclusion Code: X4
337. Rasolofo V, Rasolonalavona T, Ramarokoto H, et al. Predictive values of the ICT Tuberculosis test for the routine diagnosis of tuberculosis in Madagascar. *Int J Tuberc Lung Dis*. 2000 Feb;4(2):184-5. PMID: 10694099. Exclusion Code: X3
338. Streeton JA, Desem N, Jones SL. Sensitivity and specificity of a gamma interferon blood test for tuberculosis infection. *Int J Tuberc Lung Dis*. 1998 Jun;2(6):443-50. PMID: 9626600. Exclusion Code: X3
339. Kendig EL, Jr., Kirkpatrick BV, Carter WH, et al. Underreading of the tuberculin skin test reaction. *Chest*. 1998 May;113(5):1175-7. PMID: 9596290. Exclusion Code: X5
340. Sonmez E, Yakinci C, Aladag M, et al. Diagnosis of tuberculosis: PPD or BCG test. *J Trop Pediatr*. 1998 Feb;44(1):40-2. PMID: 9538605. Exclusion Code: X3
341. McCurdy SA, Arretz DS, Bates RO. Tuberculin reactivity among California Hispanic migrant farm workers. *Am J Ind Med*. 1997 Dec;32(6):600-5. PMID: 9358916. Exclusion Code: X5
342. Duchin JS, Jereb JA, Nolan CM, et al. Comparison of sensitivities to two commercially available tuberculin skin test reagents in persons with recent tuberculosis. *Clin Infect Dis*. 1997 Sep;25(3):661-3. PMID: 9314456. Exclusion Code: X2
343. Wells CD, Zuber PL, Nolan CM, et al. Tuberculosis prevention among foreign-born persons in Seattle--King County, Washington. *Am J Respir Crit Care Med*. 1997 Aug;156(2 Pt 1):573-7. PMID: 9279242. Exclusion Code: X4
344. Woeltje KF, Kilo CM, Johnson K, et al. Tuberculin skin testing of hospitalized patients. *Infect Control Hosp Epidemiol*. 1997 Aug;18(8):561-5. PMID: 9276237. Exclusion Code: X2
345. Thomas RE. Mantoux (tuberculosis) testing. Evaluation of guidelines for testing in Canadian institutions. *Can Fam Physician*. 1997 May;43:933-8. PMID: 9154365. Exclusion Code: X1
346. Lifson AR, Grant SM, Lorvick J, et al. Two-step tuberculin skin testing of injection drug users recruited from community-based settings. *Int J Tuberc Lung Dis*. 1997 Apr;1(2):128-34. PMID: 9441076. Exclusion Code: X2
347. Sorresso DJ, Mehta JB, Harvill LM, et al. Underutilization of isoniazid chemoprophylaxis in tuberculosis contacts 50 years of age and older. A prospective analysis. *Chest*. 1995 Sep;108(3):706-11. PMID: 7656620. Exclusion Code: X4
348. Cieslak TJ, Irwin RG, Dougherty PA, et al. A pseudoepidemic of tuberculin skin test conversions caused by a particular lot of purified protein derivative of tuberculin test solution. *Pediatr Infect Dis J*. 1995 May;14(5):392-3. PMID: 7638016. Exclusion Code: X2
349. John GT, Thomas PP, Thomas M, et al. A double-blind randomized controlled trial of primary isoniazid prophylaxis in dialysis and transplant patients. *Transplantation*. 1994 Jun 15;57(11):1683-4. PMID: 8009608. Exclusion Code: X6
350. Bellin E, Fletcher D, Safyer S. Abnormal chest x-rays in intravenous drug users: implications for tuberculosis screening programs. *Am J Public Health*. 1993 May;83(5):698-700. PMID: 8387246. Exclusion Code: X4
351. Farmakis M, Travlou O, Aroni S, et al. Fibrinogen and antithrombin III blood levels fluctuations during isoniazid or isoniazid plus rifampicin administration. *Arzneimittelforschung*. 1992 Aug;42(8):1041-4. PMID: 1418078. Exclusion Code: X5
352. Graham NM, Nelson KE, Solomon L, et al. Prevalence of tuberculin positivity and skin test anergy in HIV-1-seropositive and -seronegative intravenous drug users. *Jama*. 1992 Jan 15;267(3):369-73. PMID: 1727959. Exclusion Code: X2
353. Lee KC, Tami TA, Lalwani AK, et al. Contemporary management of cervical tuberculosis. *Laryngoscope*. 1992 Jan;102(1):60-4. PMID: 1731159. Exclusion Code: X3
354. Martin-Casabona N, Ocana Rivera I, Vidal Pla R, et al. Diagnosis of mycobacterial infection in acquired immunodeficiency syndrome (AIDS) patients and HIV carriers. *J Hyg Epidemiol Microbiol Immunol*. 1992;36(3):293-302. PMID: 1293212. Exclusion Code: X2
355. Gordin FM, Perez-Stable EJ, Reid M, et al. Stability of positive tuberculin tests: are boosted reactions valid? *Am Rev Respir Dis*. 1991 Sep;144(3 Pt 1):560-3. PMID: 1892295. Exclusion Code: X5

Appendix C. Excluded Studies

356. Havlir DV, van der Kuyp F, Duffy E, et al. A 19-year follow-up of tuberculin reactors. Assessment of skin test reactivity and in vitro lymphocyte responses. *Chest*. 1991 May;99(5):1172-6. PMID: 2019174. Exclusion Code: X4
357. Lau SK, Wei WI, Kwan S, et al. Combined use of fine-needle aspiration cytologic examination and tuberculin skin test in the diagnosis of cervical tuberculous lymphadenitis. A prospective study. *Arch Otolaryngol Head Neck Surg*. 1991 Jan;117(1):87-90. PMID: 1986768. Exclusion Code: X3
358. Theuer CP, Hopewell PC, Elias D, et al. Human immunodeficiency virus infection in tuberculosis patients. *J Infect Dis*. 1990 Jul;162(1):8-12. PMID: 1972384. Exclusion Code: X5
359. De March-Ayuela P. Choosing an appropriate criterion for true or false conversion in serial tuberculin testing. *Am Rev Respir Dis*. 1990 Apr;141(4 Pt 1):815-20. PMID: 2327645. Exclusion Code: X5
360. Aziz S, Alam SE. Criteria for diagnosis in pulmonary tuberculosis. *J Pak Med Assoc*. 1990 Jan;40(1):14-6. PMID: 2109125. Exclusion Code: X5
361. Dutt AK, Moers D, Stead WW. Smear- and culture-negative pulmonary tuberculosis: four-month short-course chemotherapy. *Am Rev Respir Dis*. 1989 Apr;139(4):867-70. PMID: 2930066. Exclusion Code: X2
362. Franks AL, Binkin NJ, Snider DE, Jr., et al. Isoniazid hepatitis among pregnant and postpartum Hispanic patients. *Public Health Rep*. 1989 Mar-Apr;104(2):151-5. PMID: 2495549. Exclusion Code: X4
363. Onwubalili JK, Scott GM. Immune status in tuberculosis and response to treatment. *Tubercle*. 1988 Jun;69(2):81-94. PMID: 3188236. Exclusion Code: X3
364. Biggs B, Connor H, Dwyer BW, et al. Comparison of a multiple puncture tuberculin test, 'Imotest', and the Mantoux test in an Australian population. *Tubercle*. 1987 Dec;68(4):285-90. PMID: 3455568. Exclusion Code: X3
365. Stanford CF, Bell A. Rifampicin does not suppress the tuberculin reaction in man. *Eur J Respir Dis*. 1987 Nov;71(5):339-40. PMID: 3443155. Exclusion Code: X5
366. Aitken ML, Anderson KM, Albert RK. Is the tuberculosis screening program of hospital employees still required? *Am Rev Respir Dis*. 1987 Oct;136(4):805-7. PMID: 3116895. Exclusion Code: X4
367. Dorken E, Grzybowski S, Allen EA. Significance of the tuberculin test in the elderly. *Chest*. 1987 Aug;92(2):237-40. PMID: 3111795. Exclusion Code: X5
368. Pitchenik AE, Burr J, Suarez M, et al. Human T-cell lymphotropic virus-III (HTLV-III) seropositivity and related disease among 71 consecutive patients in whom tuberculosis was diagnosed. A prospective study. *Am Rev Respir Dis*. 1987 Apr;135(4):875-9. PMID: 3645999. Exclusion Code: X2
369. Cowie RL, Langton ME, Escreet BC. Diagnosis of sputum smear- and sputum culture-negative pulmonary tuberculosis. *S Afr Med J*. 1985 Dec 7;68(12):878. PMID: 3934771. Exclusion Code: X6
370. van Assendelft AH. Leucopenia in rifampicin chemotherapy. *J Antimicrob Chemother*. 1985 Sep;16(3):407-8. PMID: 4055546. Exclusion Code: X3
371. Welty C, Burstin S, Muspratt S, et al. Epidemiology of tuberculous infection in a chronic care population. *Am Rev Respir Dis*. 1985 Jul;132(1):133-6. PMID: 4014857. Exclusion Code: X4
372. Mukerjee CM, McKenzie DK. Safety of thrice-weekly rifampicin for tuberculosis in South-East Asian refugees. *Aust N Z J Med*. 1985 Apr;15(2):226-9. PMID: 3861165. Exclusion Code: X3
373. Dorken E, Grzybowski S, Enarson DA. Ten year evaluation of a trial of chemoprophylaxis against tuberculosis in Frobisher Bay, Canada. *Tubercle*. 1984 Jun;65(2):93-9. PMID: 6380067. Exclusion Code: X3
374. Isoniazid prevention of tuberculosis. *Lancet*. 1983 Feb 19;1(8321):395-6. PMID: 6130384. Exclusion Code: X1
375. Rudd RM, Gellert AR, Venning M. Comparison of Mantoux, tine, and 'Imotest' tuberculin tests. *Lancet*. 1982 Sep 4;2(8297):515-8. PMID: 6125678. Exclusion Code: X3
376. Ellard GA, Girling DJ, Nunn AJ. The hepatotoxicity of isoniazid among the three acetylator phenotypes. *Am Rev Respir Dis*. 1981 May;123(5):568-70. PMID: 7235381. Exclusion Code: X3
377. Dash LA, Comstock GW, Flynn JP. Isoniazid preventive therapy: Retrospect and prospect. *Am Rev Respir Dis*. 1980 Jun;121(6):1039-44. PMID: 7416591. Exclusion Code: X4
378. Costello HD, Snider DE, Jr. The incidence of cancer among participants in controlled, randomized isoniazid preventive therapy trial. *Am J Epidemiol*. 1980 Jan;111(1):67-74. PMID: 6986082. Exclusion Code: X6

Appendix C. Excluded Studies

379. Howe GR, Lindsay J, Coppock E, et al. Isoniazid exposure in relation to cancer incidence and mortality in a cohort of tuberculosis patients. *Int J Epidemiol.* 1979 Dec;8(4):305-12. PMID: 541154. Exclusion Code: X3
380. Falk A, Fuchs GF. Prophylaxis with isoniazid in inactive tuberculosis. A Veterans Administration Cooperative Study XII. *Chest.* 1978 Jan;73(1):44-8. PMID: 340155. Exclusion Code: X3
381. Glassroth JL, White MC, Snider DE, Jr. An assessment of the possible association of isoniazid with human cancer deaths. *Am Rev Respir Dis.* 1977 Dec;116(6):1065-74. PMID: 337865. Exclusion Code: X2
382. . Controlled trial of intermittent regimens of rifampin plus isoniazid for pulmonary tuberculosis in Singapore. The results up to 30 months. *Am Rev Respir Dis.* 1977 Nov;116(5):807-20. PMID: 411405. Exclusion Code: X3
383. Byrd RB, Horn BR, Griggs GA, et al. Isoniazid chemoprophylaxis. Association with detection and incidence of liver toxicity. *Arch Intern Med.* 1977 Sep;137(9):1130-3. PMID: 332099. Exclusion Code: X3
384. Atuk NO, Hart AD, Hunt EH. Close monitoring is essential during isoniazid prophylaxis. *South Med J.* 1977 Feb;70(2):156-9. PMID: 841390. Exclusion Code: X4
385. Grzybowski S, Ashley MJ, Pinkus G. Chemoprophylaxis in inactive tuberculosis: long-term evaluation of a Canadian trial. *Can Med Assoc J.* 1976 Apr 3;114(7):607-11. PMID: 816447. Exclusion Code: X7
386. Dubra FA. Short course treatment with two drugs, isoniazid plus rifampicin for six months: a controlled clinical trial with two treatment regimens. *Bull Int Union Tuberc.* 1976;51(1):61-4. PMID: 1030314. Exclusion Code: X3
387. Falk A, Fuchs G. Isoniazid (INH) prophylaxis in inactive pulmonary tuberculosis: report of a Veterans Administration Cooperative Study. *Bull Int Union Tuberc.* 1976;51(1):219-23. PMID: 1030286. Exclusion Code: X3
388. Eule H, Iwainsky H, Kaernbach E, et al. Intermittent treatment using isoniazid and rifampicin once a week: final results of a controlled trial. *Bull Int Union Tuberc.* 1976;51(1):115-9. PMID: 1030270. Exclusion Code: X3
389. Krebs A. The IUAT trial on isoniazid preventive treatment in persons with fibrotic lung lesions. *Bull Int Union Tuberc.* 1976;51(1):193-201. PMID: 801115. Exclusion Code: X1
390. Furcolow ML. Tuberculosis case finding by tuberculin testing a complete population, with follow-up. *Chest.* 1975 Sep;68(3 Suppl):443-5. PMID: 1157559. Exclusion Code: X3
391. Somner AR. Proceedings: Short course chemotherapy in pulmonary tuberculosis: a controlled trial by the British Thoracic and Tuberculosis Association. (A report from one of the three clinical co-ordinators). *Tubercle.* 1975 Jun;56(2):165. PMID: 1103397. Exclusion Code: X2
392. Lewis JE, Mello P, Knauer CM. Isoniazid-associated hepatitis--serum enzyme determinations and histologic features. *West J Med.* 1975 May;122(5):371-6. PMID: 1130028. Exclusion Code: X4
393. . A controlled clinical trial of small daily doses of rifampicin in the prevention of adverse reactions to the drug in a once-weekly regimen of chemotherapy in Hong Kong: second report:- the results at 12 months. *Tubercle.* 1974 Sep;55(3):193-210. PMID: 4620298. Exclusion Code: X2
394. Bailey WC, Weill H, DeRouen TA, et al. The effect of isoniazid on transaminase levels. *Ann Intern Med.* 1974 Aug;81(2):200-2. PMID: 4843577. Exclusion Code: X3
395. Comstock GW, Woolpert SF, Baum C. Isoniazid prophylaxis among Alaskan Eskimos: a progress report. *Am Rev Respir Dis.* 1974 Aug;110(2):195-7. PMID: 4414015. Exclusion Code: X3
396. Horwitz O. Long-range evaluation of a mass screening program. *Am J Epidemiol.* 1974 Jul;100(1):20-8. PMID: 4841817. Exclusion Code: X2
397. Horwitz O, Magnus K. Epidemiologic evaluation of chemoprophylaxis against tuberculosis. *Am J Epidemiol.* 1974 May;99(5):333-42. PMID: 4596645. Exclusion Code: X3
398. Erdtmann FJ, Dixon KE, Llewellyn CH. Skin testing for tuberculosis. Antigen and observer variability. *Jama.* 1974 Apr 22;228(4):479-81. PMID: 4594690. Exclusion Code: X2
399. . A Hong Kong Tuberculosis Treatment Services/Brompton Hospital/British Medical Research Council Investigation. *Tubercle.* 1974 Mar;55(1):1-27. PMID: 4620294. Exclusion Code: X2
400. Ludford J, Doster B, Woolpert SF. Effect of isoniazid on reproduction. *Am Rev Respir Dis.* 1973 Nov;108(5):1170-4. PMID: 4583539. Exclusion Code: X3
401. Ville de Goyet C, Kleeberg HH. Tuberculin testing by multiple puncture and by intradermal injection. *S Afr Med J.* 1973 Sep

Appendix C. Excluded Studies

- 15;47(36):1648-52. PMID: 4795768. Exclusion Code: X2
402. Bethlem N, Gerhardt FG, Magarao S. Comparison of two regimens of simplified two-stage chemotherapy in previously untreated pulmonary tuberculosis. *Tubercle*. 1973 Sep;54(3):180-4. PMID: 4204111. Exclusion Code: X2
403. Stead WW, Jurgens GH. Productivity of prolonged follow-up after chemotherapy for tuberculosis. *Am Rev Respir Dis*. 1973 Aug;108(2):314-20. PMID: 4198349. Exclusion Code: X2
404. Cramer PG. Tuberculosis: why our concepts have changed. *Mich Med*. 1973 Jul;72(21):473-4. PMID: 4732674. Exclusion Code: X7
405. Maddrey WC, Boitnott JK. Isoniazid hepatitis. *Ann Intern Med*. 1973 Jul;79(1):1-12. PMID: 4721174. Exclusion Code: X7
406. Horwitz O, Edwards PQ, Lowell AM. National tuberculosis control program in Denmark and the United States. *Health Serv Rep*. 1973 Jun-Jul;88(6):493-8. PMID: 4715710. Exclusion Code: X1
407. B.C.G. tested. *Br Med J*. 1973 Feb 24;1(5851):435. PMID: 4734730. Exclusion Code: X1
408. Furcolow ML, Deuschle KW. Modern tuberculosis control. A six-year follow-up in an Appalachian community. *Am Rev Respir Dis*. 1973 Feb;107(2):253-66. PMID: 4539600. Exclusion Code: X2
409. Ville de Goyet CD. The intradermal tuberculin test as a research tool. Its planning, execution, analysis and interpretation. *S Afr Med J*. 1973 Jan 27;47(4):142-5. PMID: 4595134. Exclusion Code: X1
410. Hudson LD, Sbarbaro JA. Twice weekly tuberculosis chemotherapy. *Jama*. 1973 Jan 8;223(2):139-43. PMID: 4347487. Exclusion Code: X3
411. Nitti V. Controlled clinical evaluation of three intermittent regimens employing low-dosage schedules of rifampicin in original treatment of pulmonary tuberculosis. Preliminary data on effectiveness and side effects. *Scand J Respir Dis Suppl*. 1973;84:180-5. PMID: 4593266. Exclusion Code: X2
412. Eule H, Karnbach E, Kaluza P, et al. Preliminary results of a controlled therapeutic trial administering INH-RMP once-weekly, after--or without--an initial period of continuous treatment. *Scand J Respir Dis Suppl*. 1973;84:153-9. PMID: 4593264. Exclusion Code: X2
413. Gomi J, Aoyagi T. Therapeutic effects and side effects of rifampicin administered daily or twice-weekly. *Scand J Respir Dis Suppl*. 1973;84:145-52. PMID: 4522070. Exclusion Code: X2
414. Mattson K. Side effects of rifampicin. A clinical study. *Scand J Respir Dis Suppl*. 1973;82:1-52. PMID: 4518740. Exclusion Code: X2
415. Debre R, Perdrizet S, Lotte A, et al. Isoniazid chemoprophylaxis of latent primary tuberculosis: in five trial centres in France from 1959 to 1969. *Int J Epidemiol*. 1973 Summer;2(2):153-60. PMID: 4204799. Exclusion Code: X7
416. Proust A, Evans C. The Australian rifampicin trial. *Med J Aust*. 1972 Oct 14;2(16):861-7. PMID: 4647286. Exclusion Code: X2
417. Schachter EN. Tuberculin negative tuberculosis. *Am Rev Respir Dis*. 1972 Oct;106(4):587-93. PMID: 4627830. Exclusion Code: X2
418. Byrd CB, Nelson R, Elliott RC. Isoniazid toxicity. A prospective study in secondary chemoprophylaxis. *Jama*. 1972 Jun 12;220(11):1471-3. PMID: 5067581. Exclusion Code: X3
419. Nitti V. Antituberculosis activity of rifampin. Report of studies performed and in progress (1966-1971). *Chest*. 1972 Jun;61(6):589-98. PMID: 4624281. Exclusion Code: X7
420. Verbist L, Mbete S, Van Landuyt H, et al. Intermittent therapy with rifampin once a week in advanced pulmonary tuberculosis. *Chest*. 1972 Jun;61(6):555-63. PMID: 4624278. Exclusion Code: X2
421. Controlled clinical trial of short-course (6-month) regimens of chemotherapy for treatment of pulmonary tuberculosis. *Lancet*. 1972 May 20;1(7760):1079-85. PMID: 4112569. Exclusion Code: X2
422. Narain R, Naganna K, Lal P. Nonspecific sensitivity and its influence on incidence of pulmonary tuberculosis. *Am Rev Respir Dis*. 1972 Apr;105(4):578-85. PMID: 5017881. Exclusion Code: X3
423. Doto EL, Furcolow ML, MacInnis FE. Size of the tuberculin reaction. Relationship to subsequent development of tuberculosis in Kansas-Missouri area. *Arch Environ Health*. 1971 Nov;23(5):392-6. PMID: 5133801. Exclusion Code: X4
424. Devine CM, Tall R, Nunn AJ, et al. Computer-aided procedures in a multi-centre co-operative controlled clinical trial in pulmonary tuberculosis. *Tubercle*. 1971 Sep;52(3):199-218. PMID: 4938217. Exclusion Code: X3
425. Polascik M, Golden B. Rifampicin. *Ann Ophthalmol*. 1971 Aug;3(8):877-82. PMID: 5005929. Exclusion Code: X1

Appendix C. Excluded Studies

426. Chusid EL, Shah R, Siltzbach LE. Tuberculin tests during the course of sarcoidosis in 350 patients. *Am Rev Respir Dis*. 1971 Jul;104(1):13-21. PMID: 5556229. Exclusion Code: X2
427. Moulding T. Chemoprophylaxis of tuberculosis: when is the benefit worth the risk and cost? *Ann Intern Med*. 1971 May;74(5):761-70. PMID: 5314806. Exclusion Code: X7
428. Citron KM, De Silva DJ. The results of tuberculosis chemotherapy in chest clinic practice. *Tubercle*. 1971 Mar;52(1):31-6. PMID: 5560200. Exclusion Code: X2
429. Flemming J, Virchow C. Late results following rifampicin therapy and tolerance of rifampicin given on a long-term basis. *Respiration*. 1971;28:Suppl:107-15. PMID: 5150790. Exclusion Code: X4
430. Trendelenburg F. Controlled clinical research on the therapeutic effect of rifampicin. *Respiration*. 1971;28:Suppl:123-9. PMID: 4948627. Exclusion Code: X5
431. Reid LR. Twenty-seven months of chemoprophylaxis for prevention of tuberculosis in Mississippi. *J Miss State Med Assoc*. 1970 Sep;11(9):485-92. PMID: 5457043. Exclusion Code: X7
432. Comstock GW, Ferebee SH. How much isoniazid is needed for prophylaxis? *Am Rev Respir Dis*. 1970 May;101(5):780-2. PMID: 4910642. Exclusion Code: X3
433. Narain R, Bagga AS, Naganna K, et al. Influence of isoniazid on naturally acquired tuberculin allergy and on induction of allergy by BCG vaccination. *Bull World Health Organ*. 1970;43(1):53-64. PMID: 5312322. Exclusion Code: X3
434. . A controlled comparison of a twice-weekly and three once-weekly regimens in the initial treatment of pulmonary tuberculosis. *Bull World Health Organ*. 1970;43(1):143-206. PMID: 5312317. Exclusion Code: X2
435. Horwitz O. The risk of tuberculosis in different groups of the general population. *Scand J Respir Dis Suppl*. 1970;72:59-60. PMID: 5273220. Exclusion Code: X1
436. Comstock GW, Hammes LM, Pio A. Isoniazid prophylaxis in Alaskan Boarding schools. A comparison of two doses. *Am Rev Respir Dis*. 1969 Dec;100(6):773-9. PMID: 5353852. Exclusion Code: X2
437. Grzybowski S, Ashley MJ, McKinnon NE, et al. In Canada: a trial of chemoprophylaxis in inactive tuberculosis. *Can Med Assoc J*. 1969 Nov 1;101(9):81-6. PMID: 5348494. Exclusion Code: X7
438. Schlaegel TF, Jr., Weber JC. Double-blind therapeutic trial of isoniazid in 344 patients with uveitis. *Br J Ophthalmol*. 1969 Jun;53(6):425-7. PMID: 4893874. Exclusion Code: X3
439. Ramakrishnan CV, Devadatta S, Evans C, et al. A four-year follow-up of patients with quiescent pulmonary tuberculosis at the end of a year of chemotherapy with twice-weekly isoniazid plus streptomycin or daily isoniazid plus pas. *Tubercle*. 1969 Jun;50(2):115-24. PMID: 4892322. Exclusion Code: X2
440. Evans C, Devadatta S, Fox W, et al. A 5-year study of patients with pulmonary tuberculosis treated at home in a controlled comparison of isoniazid plus PAS with 3 regimens of isoniazid alone. *Bull World Health Organ*. 1969;41(1):1-16. PMID: 5309083. Exclusion Code: X2
441. Johnson FK, Thompson R. A 35 year study of tuberculosis detection and treatment. *J Sch Health*. 1969 Jan;39(1):8-14. PMID: 5189363. Exclusion Code: X2
442. Riska N. Hospital-based ambulatory treatment of tuberculosis with capreomycin, ethambutol, and/or rifampicin. *Scand J Respir Dis Suppl*. 1969;69:75-9. PMID: 4906378. Exclusion Code: X2
443. Radenbach KL. Results of clinical studies with capreomycin, ethambutol and rifampicin in the Heckeshorn Hospital, Berlin. *Scand J Respir Dis Suppl*. 1969;69:43-53. PMID: 4906373. Exclusion Code: X2
444. Meissner G. Rifampicin in newly detected, untreated cases of pulmonary tuberculosis. *Acta Tuberc Pneumol Belg*. 1969;60(3):554-6. PMID: 4906103. Exclusion Code: X2
445. Tani P. The requirement of capreomycin, ethambutol and rifampicin in chronic pulmonary tuberculosis in Helsinki on the basis of resistance indications. *Scand J Respir Dis Suppl*. 1969;69:15-6. PMID: 4313500. Exclusion Code: X7
446. Veening GJ. Long term isoniazid prophylaxis. Controlled trial on INH prophylaxis after recent tuberculin conversion in young adults. *Bull Int Union Tuberc*. 1968 Dec;41:169-71. PMID: 4885378. Exclusion Code: X3
447. Horwitz O. Long-term results of the chemoprophylactic trial in Greenland. *Bull Int Union Tuberc*. 1968 Dec;41:167-8. PMID: 4885377. Exclusion Code: X3
448. Ferebee SH. Long-term effects of isoniazid prophylaxis. *Bull Int Union Tuberc*. 1968 Dec;41:161-6. PMID: 4885376. Exclusion Code: X7
449. Corpe RF, Blalock FA. A continuing study of patients with "open negative" status at Battey

Appendix C. Excluded Studies

- State Hospital. *Am Rev Respir Dis.* 1968 Dec;98(6):954-64. PMID: 4301382. Exclusion Code: X3
450. Myers JA, Bearman JE, Botkins AC. The natural history of tuberculosis in the human body. X. Prognosis among students with tuberculin reaction conversion before, during and after school of nursing. *Dis Chest.* 1968 Jun;53(6):687-98. PMID: 5653743. Exclusion Code: X2
 451. Stradling P. Clinical experience of intermittent treatment in the United Kingdom. *Tubercle.* 1968 Mar;49:Suppl:83-5. PMID: 4872962. Exclusion Code: X2
 452. Leskiewicz H. Permanent residual pulmonary lesions following primary tuberculosis in relation to antituberculous chemotherapy. *Pol Med J.* 1968;7(1):127-38. PMID: 5300914. Exclusion Code: X2
 453. Thomassen OK. The prognosis of pulmonary tuberculosis under modern treatment. *Scand J Respir Dis Suppl.* 1968;63:29-32. PMID: 5188461. Exclusion Code: X2
 454. Olsen PZ, Torning K. Isoniazid and loss of memory. *Scand J Respir Dis.* 1968;49(1):1-8. PMID: 5187246. Exclusion Code: X2
 455. Boszormenyi M, Schweiger O. A controlled clinical trial of the drug treatment of new cases of pulmonary tuberculosis. *Scand J Respir Dis Suppl.* 1968;65:189-93. PMID: 4978460. Exclusion Code: X2
 456. Bridge EV. Chemoprophylaxis: a major adjunct in the prevention of tuberculosis. *Mich Med.* 1967 Dec;66(24):1553-5. PMID: 6080307. Exclusion Code: X7
 457. Bobrowitz ID, Robins DE. Ethambutol-isoniazid versus PAS-isoniazid in original treatment of pulmonary tuberculosis. *Am Rev Respir Dis.* 1967 Sep;96(3):428-38. PMID: 6039097. Exclusion Code: X3
 458. Hyde L. Comparison of single and divided daily doses of isoniazid in original treatment of minimal and noncavitary moderately advanced pulmonary tuberculosis. XVIII. A report of the Veterans Administration-Armed Forces Cooperative Study on the chemotherapy of tuberculosis. *Am Rev Respir Dis.* 1967 Aug;96(2):204-8. PMID: 4951906. Exclusion Code: X3
 459. Comstock GW, Ferebee SH, Hammes LM. A controlled trial of community-wide isoniazid prophylaxis in Alaska. *Am Rev Respir Dis.* 1967 Jun;95(6):935-43. PMID: 6026165. Exclusion Code: X4
 460. Moodie AS. Mass ambulatory chemotherapy in the treatment of tuberculosis in a predominantly urban community. *Am Rev Respir Dis.* 1967 Mar;95(3):384-97. PMID: 6018699. Exclusion Code: X3
 461. Lees AW. Ethionamide, 500 mg. daily, plus isoniazid, 500 mg. or 300 mg. daily in previously untreated patients with pulmonary tuberculosis. *Am Rev Respir Dis.* 1967 Jan;95(1):109-11. PMID: 5333774. Exclusion Code: X3
 462. Shimamura K. Results of the cooperative study on chemotherapy of tuberculosis in the National Sanatoria. *Jpn J Tuberc.* 1966 Dec;13:Suppl:105-18. PMID: 5298476. Exclusion Code: X2
 463. Reisner D. Comparison of single and divided daily dosages of isoniazid and PAS in the treatment of pulmonary tuberculosis. XV. A report of the Veterans Administration-Armed Forces Cooperative. Study on the chemotherapy of tuberculosis. *Am Rev Respir Dis.* 1966 Dec;94(6):849-57. PMID: 4958797. Exclusion Code: X3
 464. Myers JA, Bearman JE, Botkins AC. Natural history of tuberculosis in the human body. IX. Prognosis among students with tuberculin reaction conversion before, during and after medical school. *Dis Chest.* 1966 Aug;50(2):120-32. PMID: 5923663. Exclusion Code: X5
 465. Nazareth O, Devadatta S, Evans C, et al. A two-year follow-up of patients with quiescent pulmonary tuberculosis following a year of chemotherapy with an intermittent (twice-weekly) regimen of isoniazid plus streptomycin or a daily regimen of isoniazid plus PAS. *Tubercle.* 1966 Jun;47(2):178-89. PMID: 5963586. Exclusion Code: X2
 466. Horwitz O, Payne PG, Wilbek E. Epidemiological basis of tuberculosis eradication. 4. The isoniazid trial in Greenland. *Bull World Health Organ.* 1966;35(4):509-26. PMID: 5335457. Exclusion Code: X3
 467. Egsmose T, Ang'awa JO, Poti SJ. The use of isoniazid among household contacts of open cases of pulmonary tuberculosis. *Bull World Health Organ.* 1965;33(3):419-33. PMID: 5321762. Exclusion Code: X3
 468. Fox Gregory J, Dobler Claudia C, Marks Guy B. Active case finding in contacts of people with tuberculosis. *Cochrane Database of Systematic Reviews: John Wiley & Sons, Ltd; 2011.* Exclusion Code: X3
 469. Song GG, Bae SC, Lee YH. Interferon-gamma release assays versus tuberculin skin testing in patients with rheumatoid arthritis (Provisional abstract). *International Journal of Rheumatic Diseases; 2013.* p. 279-83. Exclusion Code: X2
 470. Sester M, Sotgiu G, Lange C, et al. Interferon-release assays for the diagnosis of active

Appendix C. Excluded Studies

- tuberculosis: a systematic review and meta-analysis (Provisional abstract). *European Respiratory Journal*; 2011. p. 100-11. Exclusion Code: X3
471. Han T. Effectiveness of standard short-course chemotherapy for treating tuberculosis and the impact of drug resistance on its outcome. *International Journal of Evidence-Based Healthcare*; 2006. p. 101-17. Exclusion Code: X5
 472. Cox HS, Morrow M, Deutschmann PW. Long term efficacy of DOTS regimens for tuberculosis: systematic review (Structured abstract). *Bmj*; 2008. p. 484. Exclusion Code: X2
 473. Huebner RE, Schein MF, Cauthen GM, et al. Evaluation of the clinical usefulness of mycobacterial skin test antigens in adults with pulmonary mycobacterioses. *American review of respiratory disease*; 1992. p. 1160-6. Exclusion Code: X3
 474. Prognostic value of interferon gamma release assays in predicting active tuberculosis among individuals with, or at risk of, latent tuberculosis infection (Project record). *Health Technology Assessment Database: Health Technology Assessment*; 2010. Exclusion Code: X1
 475. Interferon-gamma release assays testing versus tuberculosis skin testing for tuberculosis: a review of the clinical effectiveness and guidelines (Structured abstract). *Health Technology Assessment Database: Canadian Agency for Drugs and Technologies in Health (CADTH)*; 2011. Exclusion Code: X7
 476. ?IGRA in Diagnostic Evaluation of Active TB (IDEA) (Project record). *Health Technology Assessment Database: Health Technology Assessment*; 2012. Exclusion Code: X1
 477. Hayes, Inc. Interferon-gamma release assays for tuberculosis (Structured abstract). *Health Technology Assessment Database: HAYES, Inc*; 2012. Exclusion Code: X8
 478. Jiménez-Fuentes MA, Souza-Galvao ML, Mila Augé C, et al. Rifampicin plus isoniazid for the prevention of tuberculosis in an immigrant population. *International journal of tuberculosis and lung disease*; 2013. p. 326-32. Exclusion Code: X4
 479. . Acceptability, compliance, and adverse reactions when isoniazid, rifampin, and pyrazinamide are given as a combined formulation or separately during three-times-weekly antituberculosis chemotherapy. *Am Rev Respir Dis*. 1989;140(6):1618-22. PMID: CN-00064618. Exclusion Code: X3
 480. Donomae I, Fujita S, Gomi J, et al. [Clinical evaluation of Rifampicin in pulmonary tuberculosis. 2. A controlled trial on the clinical effects of chemotherapy with Rifampicin-INH-PAS and SM-INH-PAS for treatment of original cases of pulmonary tuberculosis]. *Kekkaku : [Tuberculosis]*; 1970. p. 257-62. Exclusion Code: X2
 481. Kalankhodzhaev AA, Gamzaeva NF. [Complex treatment of patients with destructive pulmonary tuberculosis including intramuscular administration of isoniazid]. *Problemy tuberkuleza*; 1991. p. 21-2. Exclusion Code: X2
 482. Michele G, Iodice F, Lappa B. [Preliminary observations on the therapeutic effect of rifampycin associated with streptomycin or isoniazid]. *Archivio di fisiologia e delle malattie dell'apparato respiratorio*; 1968. p. 377-86. Exclusion Code: X2
 483. Weill G, Lévy A, Guignon JP, et al. [Reduction of fresh forms of pulmonary tuberculosis by the classical treatment and by combined antibiotics including rifampicin (comparative study of a personal survey)]. *Revue de tuberculose et de pneumologie*; 1969. p. 1119-25. Exclusion Code: X2
 484. Kraan JK, Mulder RJ, Dijk B. Controlled study on rifampicin in first treatment of fresh cases of pulmonary tuberculosis. *Acta tuberculosea et pneumologica Belgica*; 1969. p. 557-62. Exclusion Code: X3
 485. Adetifa IM, Ota MO, Jeffries DJ, et al. Interferon-? ELISPOT as a biomarker of treatment efficacy in latent tuberculosis infection: a clinical trial. *American journal of respiratory and critical care medicine*; 2013. p. 439-45. Exclusion Code: X5
 486. Loos U, Musch E, Mackes KG, et al. [Intravenous rifampicin therapy in open tuberculosis]. *Die Medizinische Welt*; 1983. p. 701-3. Exclusion Code: X2
 487. Kuntz HD, Rausch V. [Hepatotoxic side-effects of rifampicin; a comparative clinical study (author's transl)]. *Praxis und Klinik der Pneumologie*; 1977. p. 925-32. Exclusion Code: X2
 488. Fraga H, Gomes O, Paz de Almeida A, et al. [Comparative study (in a controlled therapeutic trial) of 3 intermittent regimens, after an initial period of daily administration, in the repeated treatment of pulmonary tuberculosis (results)]. *Bulletin of the International Union against Tuberculosis*; 1973. p. 125-38. Exclusion Code: X3
 489. Iodice F. [Long-term results of treatment with rifampicin in early pulmonary tuberculosis].

Appendix C. Excluded Studies

- Archivio Monaldi per la tisiologia e le malattie dell'apparato respiratorio; 1972. p. 51-6.
Exclusion Code: X2
490. Poppe de Figueiredo F, Brito AA, Laborne Valle JH, et al. [Short-term chemotherapy of pulmonary tuberculosis. Pilot study]. Bulletin of the International Union against Tuberculosis; 1974. p. 411-7. Exclusion Code: X3
491. Tonceanu S, Bazaciu E, Carp G, et al. [Quantitative evaluation of the dynamics of tubercular mediastinal adenopathy treated with rifampicin and ethambutol]. Revista de igien?, bacteriologie, virusologie, parazitologie, epidemiologie, pneumoftiziologie. Pneumoftiziologia; 1977. p. 143-50. Exclusion Code: X2
492. González Montaner LJ, Palma Beltran O, Abbate E, et al. [A comparison of the therapeutic efficacy of two drug combinations in cases of previously untreated open tuberculosis (author's transl)]. Praxis und Klinik der Pneumologie; 1978. p. 717-20. Exclusion Code: X2
493. Schütz I, Bartmann K. Sputum excretion of tubercle bacilli as early criterion in the evaluation of the therapeutic effectiveness of antituberculous drugs, illustrated by its use in a controlled trial of single-drug regimens. Antibiotica et chemotherapia. Fortschritte. Advances. Progrès; 1970. p. 490-500. Exclusion Code: X3
494. Bignall JR. [Controlled therapeutic study of 5 intermittent chemotherapy regimens for pulmonary tuberculosis. 4th international study of the UICT]. Bulletin of the International Union against Tuberculosis; 1974. p. 422-6. Exclusion Code: X3
495. Portilla J, Jordá P, Esteban J, et al. [Directly observed treatment of latent tuberculosis infection: comparative study of two isoniazid regimens]. Enfermedades infecciosas y microbiología clínica; 2003. p. 293-5. Exclusion Code: X3
496. [Evaluation of high dose INH twice weekly regimen in the original treatment for moderately advanced pulmonary tuberculosis. A report of the 12th series B study of the controlled trials]. Kekkaku : [Tuberculosis]; 1972. p. 159-66. Exclusion Code: X2
497. Ohmori M, Wada M, Nishii K, et al. Preventive therapy in middle-aged and elderly persons selected from the population-based screening by mass miniature radiography - Methodological aspect and adverse reactions. [Japanese]. Kekkaku : [Tuberculosis]; 2002. p. 647-58. Exclusion Code: X2
498. Favez G, Maillard JM, Vouilloz M. [Comparative results of medium and increased doses of isoniazid in the triple treatment of overt pulmonary tuberculosis]. Schweizerische medizinische Wochenschrift; 1968. p. 1392-5. Exclusion Code: X2
499. Carratù L, Sonaglioni F, Natali P. [Clinical tolerance of rifampycin administered alone or in combination with other antitubercular drugs]. Archivio di tisiologia e delle malattie dell'apparato respiratorio; 1968. p. 399-410. Exclusion Code: X3
500. Nitti V, Delli Veneri F, Marra A. [Rifomycin in treatment of chronic tuberculosis with bacilli resistant to standard medications]. Acta tuberculosea et pneumologica Belgica; 1969. p. 455-65. Exclusion Code: X2
501. Böszörményi M, Fauszt I, Károlyi A, et al. [Our initial clinical experience with rifomycin]. Acta tuberculosea et pneumologica Belgica; 1969. p. 471-7. Exclusion Code: X7
502. Baba H, Shinkai A, Azuma Y. [Controlled clinical trial of three 6 month regimens of chemotherapy for pulmonary tuberculosis. (Report 2)--Results at one year after completing chemotherapy--(author's transl)]. Kekkaku : [Tuberculosis]; 1979. p. 29-36. Exclusion Code: X2
503. Yamamoto K. [Controlled comparison of daily and intermittent treatment regimens with rifampicin]. Bulletin of the International Union against Tuberculosis; 1974. p. 442-6. Exclusion Code: X3
504. Nitti V, Delli Veneri F, Ninni A, et al. [Behavior of blood levels of rifampicin during the course of treatment]. Archivio Monaldi per la tisiologia e le malattie dell'apparato respiratorio; 1970. p. 310-20. Exclusion Code: X5
505. Eule H, Iwainsky H. [The influence of therapeutical and biological parameters on the appearance of side effects and complaints during intermittent chemotherapy of pulmonary tuberculosis (author's transl)]. Praxis und Klinik der Pneumologie; 1981. p. 347-53. Exclusion Code: X2
506. Loos U, Musch E, Mackes KG, et al. [Comparison of oral and intravenous rifampicin administration in the treatment of open pulmonary tuberculosis]. Praxis und Klinik der Pneumologie; 1983. p. 482-4. Exclusion Code: X2
507. Comstock GW. Isoniazid prophylaxis in an undeveloped area. Am Rev Respir Dis. 1962 Dec;86:810-22. PMID: 14022524. Exclusion Code: X3

Appendix C. Excluded Studies

508. Comstock GW, Baum C, Snider DE, Jr. Isoniazid prophylaxis among Alaskan Eskimos: a final report of the bethel isoniazid studies. *Am Rev Respir Dis.* 1979 May;119(5):827-30. PMID: 453704. Exclusion Code: X2
509. Bush OB, Jr., Sugimoto M, Fujii Y, et al. Isoniazid prophylaxis in contacts of persons with known tuberculosis. Second report. *Am Rev Respir Dis.* 1965 Nov;92(5):732-40. PMID: 5321147. Exclusion Code: X2
510. Del Castillo H, Bautista LD, Jacinto CP, et al. Chemoprophylaxis in the Philippines: a controlled pilot study among household contacts of tuberculosis cases. *Bulletin of the Quezon Institute.* 1965;7:277-90. Exclusion Code: X2
511. Ferebee SH, Mount FW. Tuberculosis morbidity in a controlled trial of the prophylactic use of isoniazid among household contacts. *Am Rev Respir Dis.* 1962 Apr;85:490-510. PMID: 13892318. Exclusion Code: X2
512. Ferebee SH, Mount FW, Murray FJ, et al. A controlled trial of isoniazid prophylaxis in mental institutions. *Am Rev Respir Dis.* 1963 Aug;88:161-75. PMID: 14045220. Exclusion Code: X2
513. Mount FW, Ferebee SH. The effect of isoniazid prophylaxis on tuberculosis morbidity among household contacts of previously known cases of tuberculosis. *Am Rev Respir Dis.* 1962 Jun;85:821-7. PMID: 14476668. Exclusion Code: X2
514. Meier T, Eulenbruch HP, Wrighton-Smith P, et al. Sensitivity of a new commercial enzyme-linked immunospot assay (T SPOT-TB) for diagnosis of tuberculosis in clinical practice. *Eur J Clin Microbiol Infect Dis.* 2005 Aug;24(8):529-36. PMID: 16133410. Exclusion Code: X8
515. Mori T, Sakatani M, Yamagishi F, et al. Specific detection of tuberculosis infection: an interferon-gamma-based assay using new antigens. *Am J Respir Crit Care Med.* 2004 Jul 1;170(1):59-64. PMID: 15059788. Exclusion Code: X2
516. Brock I, Munk ME, Kok-Jensen A, et al. Performance of whole blood IFN-gamma test for tuberculosis diagnosis based on PPD or the specific antigens ESAT-6 and CFP-10. *Int J Tuberc Lung Dis.* 2001 May;5(5):462-7. PMID: 11336278. Exclusion Code: X3
517. Johnson PD, Stuart RL, Grayson ML, et al. Tuberculin-purified protein derivative-, MPT-64-, and ESAT-6-stimulated gamma interferon responses in medical students before and after *Mycobacterium bovis* BCG vaccination and in patients with tuberculosis. *Clin Diagn Lab Immunol.* 1999 Nov;6(6):934-7. PMID: 10548589. Exclusion Code: X3
518. Lee JY, Choi HJ, Park IN, et al. Comparison of two commercial interferon-gamma assays for diagnosing *Mycobacterium tuberculosis* infection. *Eur Respir J.* 2006 Jul;28(1):24-30. PMID: 16611658. Exclusion Code: X2
519. Zhang S, Shao L, Mo L, et al. Evaluation of gamma interferon release assays using *Mycobacterium tuberculosis* antigens for diagnosis of latent and active tuberculosis in *Mycobacterium bovis* BCG-vaccinated populations. *Clin Vaccine Immunol.* 2010 Dec;17(12):1985-90. PMID: 20943878. Exclusion Code: X2
520. Lee HM, Shin JW, Kim JY, et al. HRCT and whole-blood interferon-gamma assay for the rapid diagnosis of smear-negative pulmonary tuberculosis. *Respiration.* 2010;79(6):454-60. PMID: 20110640. Exclusion Code: X2
521. Winqvist N, Bjorkman P, Noren A, et al. Use of a T cell interferon gamma release assay in the investigation for suspected active tuberculosis in a low prevalence area. *BMC Infect Dis.* 2009;9:105. PMID: 19575781. Exclusion Code: X2
522. Warier A, Gunawathi S, Venkatesh, et al. T-cell assay as a diagnostic tool for tuberculosis. *Indian Pediatrics.* 2010 2010/01/01;47(1):90-2. Exclusion Code: X2
523. Ferrara G, Losi M, Meacci M, et al. Routine hospital use of a new commercial whole blood interferon-gamma assay for the diagnosis of tuberculosis infection. *Am J Respir Crit Care Med.* 2005 Sep 1;172(5):631-5. PMID: 15961696. Exclusion Code: X2
524. Ferrara G, Losi M, D'Amico R, et al. Use in routine clinical practice of two commercial blood tests for diagnosis of infection with *Mycobacterium tuberculosis*: a prospective study. *Lancet.* 2006 Apr 22;367(9519):1328-34. PMID: 16631911. Exclusion Code: X4
525. Nishimura T, Hasegawa N, Mori M, et al. Accuracy of an interferon-gamma release assay to detect active pulmonary and extra-pulmonary tuberculosis. *Int J Tuberc Lung Dis.* 2008 Mar;12(3):269-74. PMID: 18284831. Exclusion Code: X4
526. Ozekinci T, Ozbek E, Celik Y. Comparison of tuberculin skin test and a specific T-cell-based test, T-SPOT.TB, for the diagnosis of latent tuberculosis infection. *J Int Med Res.* 2007 Sep-Oct;35(5):696-703. PMID: 17944056. Exclusion Code: X9
527. Brock I, Weldingh K, Leyten EM, et al. Specific T-cell epitopes for immunoassay-based diagnosis

Appendix C. Excluded Studies

- of *Mycobacterium tuberculosis* infection. *J Clin Microbiol.* 2004 Jun;42(6):2379-87. PMID: 15184408. Exclusion Code: X2
528. Taggart EW, Hill HR, Ruegner RG, et al. Evaluation of an in vitro assay for interferon gamma production in response to the *Mycobacterium tuberculosis*-synthesized peptide antigens ESAT-6 and CFP-10 and the PPD skin test. *Am J Clin Pathol.* 2006 Mar;125(3):467-73. PMID: 16613353. Exclusion Code: X5
529. Soborg B, Andersen AB, Larsen HK, et al. Detecting a low prevalence of latent tuberculosis among health care workers in Denmark detected by *M. tuberculosis* specific IFN-gamma whole-blood test. *Scand J Infect Dis.* 2007;39(6-7):554-9. PMID: 17577817. Exclusion Code: X2
530. Lalvani A, Nagvenkar P, Udwadia Z, et al. Enumeration of T cells specific for RD1-encoded antigens suggests a high prevalence of latent *Mycobacterium tuberculosis* infection in healthy urban Indians. *J Infect Dis.* 2001 Feb 1;183(3):469-77. PMID: 11133379. Exclusion Code: X3
531. Pathan AA, Wilkinson KA, Klenerman P, et al. Direct ex vivo analysis of antigen-specific IFN-gamma-secreting CD4 T cells in *Mycobacterium tuberculosis*-infected individuals: associations with clinical disease state and effect of treatment. *J Immunol.* 2001 Nov 1;167(9):5217-25. PMID: 11673535. Exclusion Code: X3
532. Stennis NL, Trieu L, Ahuja SD, et al. Estimated Prevalence of Tuberculosis Infection Among a New York City Clinic Population Using Interferon-gamma Release Assays. *Open Forum Infectious Diseases.* 2014 September 1, 2014;1(2). Exclusion Code: X2
533. LoBue PA, Moser KS. Use of isoniazid for latent tuberculosis infection in a public health clinic. *Am J Respir Crit Care Med.* 2003 Aug 15;168(4):443-7. PMID: 12746255. Exclusion Code: X4
534. Tagmouti S, Slater M, Benedetti A, et al. Reproducibility of interferon gamma (IFN-gamma) release Assays. A systematic review. *Ann Am Thorac Soc.* 2014 Oct;11(8):1267-76. PMID: 25188809. Exclusion Code: X2
535. Kruczak K, Duplaga M, Sanak M, et al. Comparison of IGRA tests and TST in the diagnosis of latent tuberculosis infection and predicting tuberculosis in risk groups in Krakow, Poland. *Scand J Infect Dis.* 2014 Sep;46(9):649-55. PMID: 25073535. Exclusion Code: X2
536. Leow MK, Dalan R, Chee CB, et al. Latent tuberculosis in patients with diabetes mellitus: prevalence, progression and public health implications. *Exp Clin Endocrinol Diabetes.* 2014 Oct;122(9):528-32. PMID: 25003362. Exclusion Code: X5
537. Lagrange PH, Thangaraj SK, Dayal R, et al. A toolbox for tuberculosis (TB) diagnosis: an Indian multi-centric study (2006-2008); evaluation of serological assays based on PGL-Tb1 and ESAT-6/CFP10 antigens for TB diagnosis. *PLoS One.* 2014;9(5):e96367. PMID: 24797271. Exclusion Code: X3
538. Park H, Shin JA, Kim HJ, et al. Whole blood interferon-gamma release assay is insufficient for the diagnosis of sputum smear negative pulmonary tuberculosis. *Yonsei Med J.* 2014 May;55(3):725-31. PMID: 24719140. Exclusion Code: X2
539. Erkens CG, Dinmohamed AG, Kamphorst M, et al. Added value of interferon-gamma release assays in screening for tuberculous infection in the Netherlands. *Int J Tuberc Lung Dis.* 2014 Apr;18(4):413-20. PMID: 24670695. Exclusion Code: X5
540. Faurholt-Jepsen D, Aabye MG, Jensen AV, et al. Diabetes is associated with lower tuberculosis antigen-specific interferon gamma release in Tanzanian tuberculosis patients and non-tuberculosis controls. *Scand J Infect Dis.* 2014 May;46(5):384-91. PMID: 24621055. Exclusion Code: X3
541. Ferguson TW, Tangri N, Macdonald K, et al. The diagnostic accuracy of tests for latent tuberculosis infection in hemodialysis patients: a systematic review and meta-analysis (Provisional abstract). *Database of Abstracts of Reviews of Effects*; 2014. p. epub. Exclusion Code: X2

Appendix D Table 1. Studies of Sensitivity of TST Screening Tests for Tuberculosis (KQ 2)

First Author, Year	Country (TB Burden ^a)	% Male	Mean Age in Years (SD)	% HIV	% BCG	Other Study Population Comments	TST 5 mm Sensitivity (95% CI, Interval) (N)	TST 10 mm Sensitivity (95% CI, Interval) (N)	TST 15 mm Sensitivity (95% CI, Interval) (N)	Quality Rating
Ak, 2009 ⁷⁰	Turkey (I)	47.7 ^b	34.4 ^b (17.9)	0	100.0	Data extracted for subjects with culture confirmation. Testing completed before treatment started for 90% of participants, and within 7 days of starting treatment for the remainder.	-	0.61 (0.45 to 0.75) (36)	-	Good
Berkel, 2005 ⁵⁴	Netherlands (L)	NR	NR	0	39.0 ^b	Data extracted for culture-confirmed patients; 19% were immunocompromised. Among sample, 86% were older than 45 years of age. BCG status reported for portion of study group. No information available on timing of testing with respect to treatment.	0.99 (0.97 to 1.00) (312)	0.96 (0.93 to 0.97) (312)	0.80 (0.75 to 0.84) 312	Fair
Bocchino, 2010 ⁷⁴	Italy (L)	60.0	39.2 (14.3)	0	43.3	Data extracted for subjects tested at baseline with culture confirmation or positive AFB smear. Study excluded subjects receiving previous TB treatment.	0.75 (0.63 to 0.84) (60)	-	-	Fair
Dilektasli, 2010 ⁷⁵	Turkey (I)	NR ^b	36.7 ^b (13.7)	NR	84.0	Data extracted for subjects with culture confirmation who had received treatment for less than 4 weeks.	0.87 (0.71 to 0.95) (31)	0.84 (0.67 to 0.93) (31)	0.26 (0.14 to 0.43) (31)	Fair
Fietta, 2003 ⁵³	Italy (L)	73.7	48.5 (NR)	0	NR	Study subjects had culture confirmation. Testing completed prior to treatment initiation.	0.65 (0.52 to 0.76) (57)	-	-	Fair
Kang, 2005 ⁵⁵	South Korea (I)	59.0	Median 43 Range 17 to 84	0	56.0	Study subjects had pathological or culture confirmation. Demographic data exclude indeterminates. No information available on	-	0.78 (0.65 to 0.87) (54)	0.70 (0.57 to 0.81) (54)	Fair

Appendix D Table 1. Studies of Sensitivity of TST Screening Tests for Tuberculosis (KQ 2)

First Author, Year	Country (TB Burden ^a)	% Male	Mean Age in Years (SD)	% HIV	% BCG	Other Study Population Comments	TST 5 mm Sensitivity (95% CI, Interval) (N)	TST 10 mm Sensitivity (95% CI, Interval) (N)	TST 15 mm Sensitivity (95% CI, Interval) (N)	Quality Rating
						timing of testing with respect to treatment.				
Mazurek, 2007 ⁶²	United States (L)	56.8 ^b	46.6 ^b Median 46.4 Range 16 to 87.1	0	33.8 ^b	Data extracted for subjects with mycobacterial confirmation and known negative HIV status. Subjects receiving treatment for longer than 7 days were not included.	0.74 (0.62 to 0.83) (69)	0.71 (0.59 to 0.80) (69)	0.62 (0.51 to 0.73) (69)	Good
Painter, 2013 ⁵¹	Vietnam (H)	68.9 ^b	37.3 ^b Range 15 to 65 and older	0.1 ^b	100.0	Data extracted for subjects with culture confirmation. No information available on timing of testing with respect to treatment.	0.89 (0.83 to 0.94) (132)	0.81 (0.74 to 0.87) (132)	0.52 (0.44 to 0.61) (132)	Fair
Park, 2009 ⁷³	South Korea (I)	54.0	52.2 (16.5)	0	NR	Data extracted for subjects with culture confirmation. No information available on timing of testing with respect to treatment.	-	0.76 (0.68 to 0.82) (153)	-	Fair
Seibert, 1991 ⁵²	United States (L)	67.0 ^b	47 ^b (18.4)	NR	NR	Data extracted for subjects with extrapulmonary TB culture-confirmed from sputum, pleural fluid, or pleural biopsy with demonstrated clinical evidence for TB. No information available on timing of testing with respect to treatment.	-	0.93 (0.81 to 0.98) (43)	-	Fair
Soysal, 2008 ⁶⁹	Turkey (I)	56.0	35 (16)	0	78.0	Data extracted for subjects with culture confirmation. All subjects had been untreated or treated for less than 7 days at the time of testing.	0.81 (0.72 to 0.87) (99)	0.70 (0.60 to 0.78) (99)	0.41 (0.32 to 0.51) (99)	Fair

Appendix D Table 1. Studies of Sensitivity of TST Screening Tests for Tuberculosis (KQ 2)

First Author, Year	Country (TB Burden ^a)	% Male	Mean Age in Years (SD)	% HIV	% BCG	Other Study Population Comments	TST 5 mm Sensitivity (95% CI, Interval) (N)	TST 10 mm Sensitivity (95% CI, Interval) (N)	TST 15 mm Sensitivity (95% CI, Interval) (N)	Quality Rating
Tsiouris, 2006 ⁵⁷	South Africa (H)	62.3 ^b	Male ^b 38 Female: 36.5 (NR)	0	65.7 ^b	Study subjects had culture confirmation. Data extracted for HIV-negative subjects.	-	0.94 (0.72 to 0.99) (16)	-	Good
Włodarczyk, 2014 ⁹⁸	Poland (I)	51.2	48.6 (18.2)	0	100	Data extracted for subjects with culture confirmation. Timing of treatment in relation to testing unstated.	0.58 (0.43 to 0.72) (43)	0.56 (0.41 to 0.70) (43)	0.26 (0.15 to 0.40) (43)	Good

^a TB burden according to World Health Organization classification. (L) Low <10 cases/100,000; (I) Intermediate 10–99 cases/100,000; (H) High >100 cases/100,000.

^b Represents demographics of the overall study population; demographics for subjects eligible for inclusion in analysis were not reported.

AFB = acid fast bacilli; BCG = bacillus Calmette-Guerin; CI = confidence interval; HIV = human immunodeficiency virus; SD = standard deviation; TB = tuberculosis; TST = tuberculin skin test.

Appendix D Table 2. Studies of Sensitivity of IGRA Screening Tests for Tuberculosis (KQ 2)

First Author, Year	Country (TB Burden ^a)	% Male	Mean Age in Years (SD)	% HIV	% BCG	Other Study Population Comments	T-SPOT.TB Sensitivity (95% CI, Interval) (N)	QFT-G Sensitivity (95% CI, Interval) (N)	QFT-GIT Sensitivity (95% CI, Interval) (N)	Quality Rating
Adetifa, 2007 ⁵⁸	Gambia (H)	63.8	31.2 IQR 23 to 36	8.8	23.8	Data extracted for subjects with smear and culture confirmation. No information available on timing of testing with respect to treatment.	-	-	0.64 (0.53 to 0.74) (75)	Fair
Ak, 2009 ⁷⁰	Turkey (I)	47.7 ^b	34.4 ^b (17.9)	0	100.0	Data extracted for subjects with culture confirmation. Testing completed before treatment started for 90% of participants, and within 7 days of starting treatment for the remainder.	-	0.78 (0.62 to 0.88) (36)	-	Good
Bocchino, 2010 ⁷⁴	Italy (L)	60.0	39.2 (14.3)	0	43.3	Data extracted for subjects tested at baseline with culture confirmation or positive AFB smear. Study excluded subjects receiving previous TB treatment.	-	-	0.88 (0.78 to 0.94) (60)	Fair
Boyd, 2011 ⁷⁹	United Kingdom (I)	57.0 ^b	NR	7.0 ^b	NR	Data extracted for subjects with positive AFB sputum, culture, or molecular confirmation. No information available on timing of testing with respect to treatment.	0.76 (0.59 to 0.87) (33)	-	-	Good
Chee, 2008 ⁶⁴	Singapore (I)	74.1	Median 48.6 Range 17 to 77	0	NR	Data extracted for HIV-negative subjects with culture confirmation. Study population recruited up to 14 days after starting treatment but 79% tested within 7 days of receiving treatment.	0.94 (0.90 to 0.96) (263)	-	0.79 (0.74 to 0.83) (283)	Good

Appendix D Table 2. Studies of Sensitivity of IGRA Screening Tests for Tuberculosis (KQ 2)

First Author, Year	Country (TB Burden ^a)	% Male	Mean Age in Years (SD)	% HIV	% BCG	Other Study Population Comments	T-SPOT.TB Sensitivity (95% CI, Interval) (N)	QFT-G Sensitivity (95% CI, Interval) (N)	QFT-GIT Sensitivity (95% CI, Interval) (N)	Quality Rating
Cho, 2011 ⁸⁰	South Korea (I)	41.1 ^b	48.3 ^b (16.1)	0	NR	Data extracted for immunocompetent subjects with culture or PCR confirmation. No information available on timing of testing with respect to treatment.	0.88 (0.80 to 0.92) (120)	-	-	Good
Dewan, 2007 ⁵⁹	United States (L)	NR	Range ^b 0 to 76	9.0 ^b	NR	Data extracted for group including 3 HIV-positive subjects.	-	0.62 (0.46 to 0.76) (37)	-	Fair
Dilektasli, 2010 ⁷⁵	Turkey (I)	36.7 ^b	13.4 ^b NR	NR	84.0	Data extracted for subjects with culture confirmation who had received treatment for less than 4 weeks.	0.74 (0.57 to 0.86) (31)	-	-	Fair
Erdem, 2014 ⁹⁹	Multiple (L and I)	52.6	39.7 (18.4)	NR	NR	Patient population culture confirmed tuberculous meningitis. Timing of test with respect to treatment not reported.	-	-	0.90 (0.77 to 0.96) (41)	Fair
Feng, 2013 ⁹¹	Taiwan (I)	67.5	63.6 (19.7)	0	47.6	Data extracted for subjects with pathology or culture confirmation. Timing of testing with respect to treatment unclear.	-	-	0.88 (0.81 to 0.92) (130)	Fair
Goletti, 2006 ⁵⁶	Italy(L)	65.2	33 (SE ± 2)	0	78.3	Study subjects had positive AFB smear or culture confirmation. Testing completed before treatment initiation.	0.91 (0.73 to 0.98) (23)	0.83 (0.63 to 0.93) (23)	-	Fair
Harada, 2008 ⁶⁵	Japan (I)	73.0	53.3 (NR)	1.0	37.0	Study subjects had positive culture or positive nucleic acid amplification. All subjects received less than 7 days of treatment prior to testing.	-	0.79 (0.70 to 0.86) (100)	0.87 (0.79 to 0.92) (100)	Good

Appendix D Table 2. Studies of Sensitivity of IGRA Screening Tests for Tuberculosis (KQ 2)

First Author, Year	Country (TB Burden ^a)	% Male	Mean Age in Years (SD)	% HIV	% BCG	Other Study Population Comments	T-SPOT.TB Sensitivity (95% CI, Interval) (N)	QFT-G Sensitivity (95% CI, Interval) (N)	QFT-GIT Sensitivity (95% CI, Interval) (N)	Quality Rating
Higuchi, 2009 ⁷¹	Japan (I)	78.7	52.7 Range 17 to 91	NR	100.0	Study subjects had culture, PCR, or positive smear confirmation before treatment or within 1 week after the start of treatment.	0.96 (0.86 to 0.99) (49)	0.87 (0.75 to 0.94) (47)	-	Fair
Janssens, 2007 ⁶⁰	Switzerland (L)	51.7	37 (17)	0	NR	Study subjects had smear or culture confirmation. Foreign-born represented 86% of the study group. Testing completed within 2 weeks of initiating treatment.	0.98 (0.91 to 1.00) (58)	-	-	Fair
Jeon, 2013 ⁹²	South Korea (I)	60.7	54.8 (20.1)	0	NR	Data extracted for subjects with PCR or culture confirmation. In this group, 13.7% were non-HIV immunosuppressed due to medications or advanced cancer. Subjects taking TB medication prior to exam were excluded.	-	-	0.65 (0.57 to 0.72) (168)	Fair
Kang, 2005 ⁵⁵	South Korea (I)	59.0	Median 43 Range 17 to 84	0	56.0	Study subjects had pathologic or culture confirmation. Demographic data excludes indeterminates. No information available on timing of testing with respect to treatment.	-	0.76 (0.63 to 0.85) (58)	-	Fair
Kim, 2011 ⁸¹	South Korea (I)	54.4	Median 49 Range 16 to 94	0	NR	Data extracted for subjects with culture confirmation. QFT testing completed before treatment initiation.	-	-	0.86 (0.82 to 0.89) (362)	Good (QFT-G) Poor (TST)
Kim, 2013 ⁹³	South Korea (I)	56.5	Median 48 Range	NR	67.4	Data extracted for subjects with positive sputum culture, or molecular	-	-	0.89 (0.77 to 0.95) (46)	Fair

Appendix D Table 2. Studies of Sensitivity of IGRA Screening Tests for Tuberculosis (KQ 2)

First Author, Year	Country (TB Burden ^a)	% Male	Mean Age in Years (SD)	% HIV	% BCG	Other Study Population Comments	T-SPOT.TB Sensitivity (95% CI, Interval) (N)	QFT-G Sensitivity (95% CI, Interval) (N)	QFT-GIT Sensitivity (95% CI, Interval) (N)	Quality Rating
			28 to 86			confirmation, though 2 subjects had clinical confirmation. No information available on the timing of testing with respect to treatment.				
Kim, 2014 ¹⁰⁰	South Korea (I)	39.0	64.0 (19)	5.0	NR	Study population limited to those with miliary TB. Timing of testing with respect to treatment not specifically reported, but testing was done within 5 days of hospital presentation, so likely no treatment for longer than 7 days prior to testing.	-	-	0.68 (0.53 to 0.80) (44)	Good
Kobashi, 2008 ⁶⁶	Japan (I)	64.3	62.8 (10.8)	0	NR	Study subjects had microbiological confirmation. No information on timing of testing with respect to treatment available, although study excluded 10 patients due to previous TB treatment.	-	0.81 (0.68 to 0.90) (48)	-	Fair
Kobashi, 2008 ⁶⁸	Japan (I)	77.0	NR	0	60.8	Study subjects had culture-confirmed pulmonary or extrapulmonary TB. No information available on the timing of testing with respect to treatment.	-	0.81 (0.68 to 0.90) (48)	-	Fair
Kobashi, 2008 ⁶⁷	Japan (I)	75.0	59.6 (10.6)	0	58.0	Data extracted for subjects with culture confirmation. Testing completed prior to treatment initiation.	0.88 (0.75 to 0.94) (48)	0.85 (0.77 to 0.90) (130)	-	Good

Appendix D Table 2. Studies of Sensitivity of IGRA Screening Tests for Tuberculosis (KQ 2)

First Author, Year	Country (TB Burden ^a)	% Male	Mean Age in Years (SD)	% HIV	% BCG	Other Study Population Comments	T-SPOT.TB Sensitivity (95% CI, Interval) (N)	QFT-G Sensitivity (95% CI, Interval) (N)	QFT-GIT Sensitivity (95% CI, Interval) (N)	Quality Rating
Kobashi, 2009 ⁷²	Japan (I)	60.0	57.7 (10.2)	1.0	60.1	Data extracted for subjects with culture confirmation. No information available on the timing of testing with respect to treatment.	-	0.81 (0.68 to 0.90) (48)	-	Fair
Kobashi, 2012 ⁸⁸	Japan (I)	77.2	65.2 (10)	0	NR	Study subjects had culture-confirmed pulmonary or extrapulmonary TB. 9% of subjects received previous anti-TB treatment and 14% of subjects received immunosuppressive treatment. No information available on the timing of testing with respect to treatment.	0.95 (0.78 to 0.99) (22)	0.82 (0.61 to 0.93) (22)	0.86 (0.67 to 0.95) (22)	Fair
Lai, 2011 ⁸³	Taiwan (I)	71.0 ^b	57.5 ^b (18.5)	8.0 ^b	NR	Data extracted for subjects with culture confirmation. No information available on timing of testing with respect to treatment.	0.90 (0.60 to 0.98) (10)	-	0.65 (0.55 to 0.74) (98)	Fair
Lai, 2011 ⁸²	Taiwan (I)	51.1 ^b	55.2 ^b 16.4	6.7 ^b	NR	Data extracted for subjects with M.Tb culture confirmation. No information available on timing of testing with respect to treatment.	0.88 (0.80 to 0.93) (98)	-	-	Fair
Lee, 2012 ⁸⁹	South Korea (I)	62.0	61 (19.4)	0	NR	Study subjects had positive nucleic acid amplification PCR or culture confirmation from sputum or pleural fluid. No information available on timing of testing with respect to treatment.	-	-	0.78 (0.67 to 0.87) (65)	Good

Appendix D Table 2. Studies of Sensitivity of IGRA Screening Tests for Tuberculosis (KQ 2)

First Author, Year	Country (TB Burden ^a)	% Male	Mean Age in Years (SD)	% HIV	% BCG	Other Study Population Comments	T-SPOT.TB Sensitivity (95% CI, Interval) (N)	QFT-G Sensitivity (95% CI, Interval) (N)	QFT-GIT Sensitivity (95% CI, Interval) (N)	Quality Rating
Legesse, 2010 ⁷⁶	Ethiopia (H)	54.3 ^b	34.2 ^b (NR)	0	20.0 ^b	Data extracted for subjects with culture confirmation or positive AFB smear. Study excluded patients on TB treatment.	-	-	0.65 (0.47 to 0.79) (31)	Fair
Losi, 2007 ⁶¹	Netherlands, Germany, and Italy (L)	40.0	42.3 (17.4)	NR	NR	Data extracted for subjects with microbiological or PCR confirmation. No information available on timing of test with respect to treatment.	1.00 (0.72 to 1.00) (10)	-	-	Fair
Lui, 2011 ⁸⁴	Hong Kong (I)	74.6	Median 47	1.6	83.0	Data extracted for subjects with culture or histological confirmation, with 3 patients confirmed by clinic-radiological characteristics and response to therapy. Testing performed prior to initiation of treatment.	-	0.60 (0.47 to 0.72) (55)	-	Fair
Metcalfe, 2010 ⁷⁷	United States (L)	69.0	Median 50 IQR 36 to 62	0	18.0	Study subjects had culture confirmed pulmonary or extrapulmonary TB but were AFB smear-negative. Study excluded patients who had received TB treatment for 7 days or longer.	-	0.72 (0.60 to 0.82) (65)	-	Fair
Min, 2013 ⁹⁴	South Korea (I)	56.8 ^b	Median ^b 66 Range 27 to 90	NR	32.4 ^b	Data extracted for subjects with culture confirmation. 7 subjects had history of treatment although no information available on the timing of treatment with respect to testing.	-	-	0.85 (0.68 to 0.94) (27)	Fair (Sn) Poor (Sp)

Appendix D Table 2. Studies of Sensitivity of IGRA Screening Tests for Tuberculosis (KQ 2)

First Author, Year	Country (TB Burden ^a)	% Male	Mean Age in Years (SD)	% HIV	% BCG	Other Study Population Comments	T-SPOT.TB Sensitivity (95% CI, Interval) (N)	QFT-G Sensitivity (95% CI, Interval) (N)	QFT-GIT Sensitivity (95% CI, Interval) (N)	Quality Rating
Pai, 2007 ⁶³	India (H)	75.0 ^b	36.4 ^b Range 18 to 76	0	41.0 ^b	Data extracted for HIV-negative subjects with culture or smear confirmation. Data extracted only from testing before treatment.	-	-	0.76 (0.60 to 0.87) (37)	Good
Painter, 2013 ⁵¹	Vietnam (H)	68.9 ^b	37.3 ^b Range 15 to 65 and older	0.1 ^b	100.0	Data extracted for subjects with culture confirmation. No information available on timing of testing with respect to treatment.	-	-	0.86 (0.79 to 0.91) (132)	Fair
Park, 2009 ⁷³	South Korea (I)	54.0	52.2 (16.5)	0	NR	Data extracted for subjects with culture confirmation. No information available on timing of testing with respect to treatment.	-	-	0.88 (0.82 to 0.92) (153)	Fair
Qian, 2013 ⁹⁵	China (H)	66.2 ^b	45.8 (17.3) ^b	0	84.7 ^b	Data extracted for subjects with positive AFB smear. No subjects were receiving treatment.	-	-	0.82 (0.75 to 0.87) (157)	Fair
Ra, 2011 ⁸⁵	South Korea (I)	42.1	Median 49 Range 22 to 83	0	84.6	Data extracted for subjects with positive AFB smear and culture confirmation. Information not available on timing of testing with respect to treatment. Subjects included 9 patients with prior history of TB and 13 immuno-suppressed patients.	-	0.89 (0.76 to 0.96) (38)	-	Fair (QFT-G) Poor (TST)

Appendix D Table 2. Studies of Sensitivity of IGRA Screening Tests for Tuberculosis (KQ 2)

First Author, Year	Country (TB Burden ^a)	% Male	Mean Age in Years (SD)	% HIV	% BCG	Other Study Population Comments	T-SPOT.TB Sensitivity (95% CI, Interval) (N)	QFT-G Sensitivity (95% CI, Interval) (N)	QFT-GIT Sensitivity (95% CI, Interval) (N)	Quality Rating
Ruhwald, 2011 ⁸⁶	Italy (L), Denmark (L), Sweden (L), Spain (L), Greece (L), Finland (L)	57.0	Median 37 Range 18 to 90	7.0	NR	Study subjects had positive culture, PCR, or microscopy or histology with a response to treatment. Testing completed within the first 2 weeks of treatment.	0.90 (0.78 to 0.95) (48)	-	0.79 (0.72 to 0.85) (168)	Good
Soysal, 2008 ⁶⁹	Turkey (I)	56.0	35 (16)	0	78.0	Data extracted for subjects with culture confirmation. All subjects had been untreated or treated for less than 7 days at the time of testing.	0.83 (0.75 to 0.89) (96)	0.78 (0.69 to 0.85) (100)	-	Fair
Taki-Eddin, 2012 ⁹⁰	Syria (I)	NR	NR	NR	NR	Data extracted for subjects with culture confirmation. No information available on timing of testing with respect to treatment.	-	-	0.87 (0.73 to 0.94) (38)	Fair
Tan, 2010 ⁷⁸	Taiwan (I)	75.0 ^b	67 ^b (12.9)	1.2 ^b	NR	Data extracted for subjects with culture confirmation. All subjects had diabetes. 5 subjects were reported to have received anti-TB treatment prior to testing, but timing of treatment is not described.	0.86 (0.72 to 0.93) (42)	-	-	Fair
Tsiouris, 2006 ⁵⁷	South Africa (H)	62.3 ^b	Male: ^b 38 Female: 36.5 (NR)	0	65.7 ^b	Study subjects had culture confirmation. Data extracted for HIV-negative subjects.	-	-	0.73 (0.48 to 0.89) (15)	Good

Appendix D Table 2. Studies of Sensitivity of IGRA Screening Tests for Tuberculosis (KQ 2)

First Author, Year	Country (TB Burden ^a)	% Male	Mean Age in Years (SD)	% HIV	% BCG	Other Study Population Comments	T-SPOT.TB Sensitivity (95% CI, Interval) (N)	QFT-G Sensitivity (95% CI, Interval) (N)	QFT-GIT Sensitivity (95% CI, Interval) (N)	Quality Rating
Walsh, 2011 ⁸⁷	United States (L), Mexico (I)	T-SPOT.TB: 65.1 GFT-2G: 67.5	T-SPOT.TB: Range 20 to 60 and older GFT-2G: Range 20 to 60 and older	T-SPOT.TB: 7.0 GFT-2G: 3.0	T-SPOT.TB: 87.5 GFT-2G: 74.5	Study excluded patients receiving treatment more than 7 days with culture confirmation or AFB smear positive.	0.93 (0.81 to 0.98) (43)	0.70 (0.63 to 0.77) (169)	-	Fair
Wang, 2013 ⁹⁶	China (H)	65.4	46 Range 20 to 75	0	80.1	Data extracted for subjects with positive AFB smear or sputum culture confirmation. Subjects received testing prior to or within 7 days of beginning treatment.	-	-	0.85 (0.66 to 0.94) (26)	Fair
Włodarczyk, 2014 ⁹⁸	Poland (I)	51.2	48.6 (18.2)	0	100	Data extracted for subjects with culture confirmation. Timing of treatment in relation to testing unstated.	-	-	0.65 (0.50 to 0.78) (43)	Good

^aTB burden according to World Health Organization classification. (L) Low <10 cases/100,000; (I) Intermediate 10–99 cases/100,000; (H) High >100 cases/100,000.

^bRepresents demographics of the overall study population; demographics for subjects eligible for inclusion in analysis were not reported.

AFB = acid fast bacilli; BCG = bacillus Calmette-Guerin; CI = confidence interval; HIV = human immunodeficiency virus; IGRA = interferon gamma release assay; IQR = interquartile range; M.Tb = Mycobacterium tuberculosis; N = number analyzed; NR = not reported; QFT-G = QuantiFERON TB Gold[®] test (2nd generation test); QFT-GIT = QuantiFERON TB Gold-In-Tube[®] test (3rd generation test); SD = standard deviation; TB = tuberculosis; T-SPOT.TB = commercial ELISPOT assay; TST = tuberculin skin test.

Appendix D Table 3. Studies of Specificity of TST Screening Tests for Tuberculosis (KQ 2)

First Author, Year	Country (TB Burden ^a)	% Male	Mean Age in Years (SD)	% HIV	% BCG	Other Study Population Comments	TST 5 mm Specificity (95% CI, Interval) (N)	TST 10 mm Specificity (95% CI, Interval) (N)	TST 15 mm Specificity (95% CI, Interval) (N)	Quality Rating
Bellefleur, 2002 ¹¹⁶	United States (L)	41.1 ^b	NR	NR	NR	Data extracted for study subjects at low risk for TB.	-	-	0.96 (0.87 to 0.99) (52)	Fair
Berkel, 2005 ⁵⁴	Netherlands (L)	41.0	24.2 (6.1)	NR	0	Study only included patients under 40 years of age and excluded patients with BCG vaccination. All study subjects screened due to intended travel.	0.95 (0.94 to 0.96) (2848)	0.97 (0.96 to 0.98) (2848)	0.99 (0.98 to 0.99) (2848)	Fair
Bieneck, 2009 ¹²¹	United States (L)	83.5 ^b	NR	0	3.3 ^b	Data extracted for participants classified as “low risk” for TB.	-	1.00 (0.99 to 1.00) (296)	-	Fair
Dilektasli, 2010 ⁷⁵	Turkey (I)	36.7 ^b	13.7 ^b (NR)	NR	93.4 ^b	Study subjects were healthy controls with no history of TB or exposure.	-	-	0.57 (0.39 to 0.73) (30)	Fair
Fietta, 2003 ⁵³	Italy (L)	57.1	27 (NR)	0	0	Study subjects were healthy, “low- risk” volunteers with no stated possible risk factors for M.tb exposure.	-	0.95 (0.84 to 0.99) (42)	-	Fair
Katsenos, 2010 ¹²²	Greece (L)	100.0	24.3 (4.0)	NR	100.0	Population is Greek army recruits. Study excluded individuals with treatment for active or latent TB, suspected current TB, prior “severe” TST reaction, known TB exposure, or any known immunosuppressive condition.	0.94 (0.92 to 0.95) (1750)	0.95 (0.93 to 0.95) (1750)	0.97 (0.96 to 0.97) (1750)	Good
Mancuso, 2012 ¹²³	United States (L)	65.5 ^b	21.8 ^b (4.6)	NR	3.5 ^b	Data extracted for subjects classified as “low risk” for TB based on history. Population is U.S. military recruits.	-	0.99 (0.98 to 0.99) (1373)	0.99 (0.99 to 1.00) (1373)	Fair
Mazurek, 2001 ¹¹⁵	United States (L)	50.0 ^b	39 ^b (NR)	0	NR	Data extracted for subjects at low risk for latent TB.	-	-	0.98 (0.93 to 0.99) (98)	Good

Appendix D Table 3. Studies of Specificity of TST Screening Tests for Tuberculosis (KQ 2)

First Author, Year	Country (TB Burden ^a)	% Male	Mean Age in Years (SD)	% HIV	% BCG	Other Study Population Comments	TST 5 mm Specificity (95% CI, Interval) (N)	TST 10 mm Specificity (95% CI, Interval) (N)	TST 15 mm Specificity (95% CI, Interval) (N)	Quality Rating
Mazurek, 2007 ¹²⁰	United States (L)	94.3 ^b	20 ^b Median 20 Range 17 to 39	NR	2.2	Data extracted for subjects classified as “low risk” for TB. Population is U.S. Navy recruits.	0.97 (0.95 to 0.98) (551)	0.98 (0.97 to 0.99) (551)	0.99 (0.98 to 1.00) (551)	Fair
Soysal, 2008 ⁶⁹	Turkey (I)	62.0	22 (12)	0	83.0	Population is healthy medical students with no previous clinical patient contact and no history of TB exposure.	0.30 (0.19 to 0.44) (47)	0.45 (0.31 to 0.59) (47)	0.60 (0.45 to 0.72) (47)	Fair
Taggart, 2004 ¹¹⁷	United States (L)	50.0 ^b	31.5 (NR)	0	0	-	-	-	0.92 (0.83 to 0.97) (66)	Fair
Taggart, 2006 ¹¹⁸	United States (L)	42.3 ^b	37.3 Range 20 to 67	NR	0	Data extracted for subjects considered low risk with no known risk factors for TB exposure, non-BCG vaccinated, with no history of active TB infection. Study subjects enrolled at an on-site employee health clinic. Participants originated from 20 countries.	-	-	0.96 (0.90 to 0.99) (81)	Fair
Villarino, 1999 ¹¹³	United States (L)	38.0	Median 26 Range 18 to 50	NR	0	Participants received the TST with the PPD-S1 antigen. Study excluded any person with known immunodeficiency.	-	0.99 (0.98 to 0.99) (1555)	1.00 (0.99 to 1.00) (1555)	Fair
Villarino, 2000 ¹¹⁴	United States (L)	37.8	Median 27	NR	0	Participants received the TST with the PPD-S2 antigen. Study excluded any person known to have a condition that could suppress delayed-type hypersensitivity, including HIV infection.	-	0.98 (0.98 to 0.99) (1189)	1.00 (0.99 to 1.00) (1189)	Fair

^aTB burden according to World Health Organization classification. (L) Low <10 cases/100,000; (I) Intermediate 10–99 cases/100,000; (H) High >100 cases/100,000.

Appendix D Table 3. Studies of Specificity of TST Screening Tests for Tuberculosis (KQ 2)

^b Represents demographics of the overall study population; demographics for subjects eligible for inclusion in analysis were not reported.

BCG = bacillus Calmette-Guerin; CI = confidence interval; HIV = human immunodeficiency virus; IGRA = interferon gamma release assay; N = number analyzed; NR = not reported; M.Tb = mycobacterium tuberculosis; PPD = purified protein derivative; SD = standard deviation; TB = tuberculosis; TST = tuberculin skin test.

Appendix D Table 4. Studies of Specificity of IGRA Screening Tests for Tuberculosis (KQ 2)

First Author, Year	Country (TB Burden ^a)	% Male	Mean Age in Years (SD)	% HIV	% BCG	Other Study Population Comments	T-SPOT.TB Specificity (95% CI, Interval) (N)	QFT-G Specificity (95% CI, Interval) (N)	QFT-GIT Specificity (95% CI, Interval) (N)	Quality Rating
Bienek, 2009 ¹²¹	United States (L)	83.5 ^b	NR	0	3.3 ^b	Data extracted for participants classified as “low risk” for TB.	0.95 (0.91 to 0.97) (291)	-	-	Fair
Bua, 2007 ¹¹⁹	Italy (L)	51.9	Median 45 Range 20 to 70	NR	NR	Data extracted for healthy subjects with negative TST.	-	1.00 (0.81 to 1.00) (16)	-	Fair
Dilektasli, 2010 ⁷⁵	Turkey (I)	36.7 ^b	13.7 ^b (NR)	NR	93.4 ^b	Study subjects were healthy controls with no history of TB or exposure.	0.73 (0.56 to 0.86) (30)	-	-	Fair
Kim, 2013 ⁹³	South Korea (I)	43.8	Median 37 Range 18 to 56	NR	78.1	Data extracted for healthy subjects with no known history of contact with TB patients, normal chest radiographs, and no symptoms of active TB.	-	-	0.60 (0.49 to 0.71) (73)	Fair
Lempp, 2015 ¹²⁴	United States (L)	NR	NR	NR	NR	TST, QFT, and QFT-G results from a portion of subjects previously reported; only abstracted data for QFT-GIT low-risk subjects.	-	-	0.98 (0.97 to 0.99) (525)	Fair
Mancuso, 2012 ¹²³	United States (L)	65.5 ^b	21.8 ^b (4.6)	NR	3.5 ^b	Data extracted for subjects classified as “low risk” for TB based on history. Population is U.S. military recruits.	0.97 (0.96 to 0.98) (1373)	-	0.99 (0.98 to 0.99) (1354)	Fair
Ruhwald, 2011 ⁸⁶	Italy(L), Denmark (L), Spain (I)	59.0 ^b	Median 22 Range 19 to 53	1.0 ^b	NR	Data extracted for subjects with no known exposure to TB and no prior TB diagnosis or treatment. Study subjects were students and nonexposed volunteers.	0.99 (0.92 to 1.00) (70)	-	0.99 (0.95 to 1.00) (101)	Good

Appendix D Table 4. Studies of Specificity of IGRA Screening Tests for Tuberculosis (KQ 2)

First Author, Year	Country (TB Burden ^a)	% Male	Mean Age in Years (SD)	% HIV	% BCG	Other Study Population Comments	T-SPOT.TB Specificity (95% CI, Interval) (N)	QFT-G Specificity (95% CI, Interval) (N)	QFT-GIT Specificity (95% CI, Interval) (N)	Quality Rating
Soysal, 2008 ⁶⁹	Turkey (I)	62.0	22 (12)	0	83.0	Population was healthy medical students with no previous clinical patient contact and no history of TB exposure.	0.85 (0.72 to 0.92) (46)	0.89 (0.77 to 0.95) (47)	-	Fair
Taggart, 2006 ¹¹⁸	United States (L)	42.3 ^b	37.3 Range 20 to 67	NR	0	Data extracted for subjects considered low risk with no known risk factors for TB exposure, non-BCG vaccinated, with no history of active TB infection. Study subjects enrolled at an on-site employee health clinic. Participants originated from 20 countries.	-	1.00 (0.95 to 1.00) (81)	-	Fair

^aTB burden according to World Health Organization classification. (L) Low <10 cases/100,000; (I) Intermediate 10–99 cases/100,000; (H) High >100 cases/100,000.

^b Represents demographics of the overall study population; demographics for subjects eligible for inclusion in analysis were not reported.

BCG = bacillus Calmette-Guerin; CI = confidence interval; HIV = human immunodeficiency virus; IGRA = interferon gamma release assay; NR = not reported; QFT-G = QuantiFERON TB Gold[®] test (2nd generation test); QFT-GIT = QuantiFERON TB Gold-In-Tube[®] test (3rd generation test); SD = standard deviation; TB = tuberculosis; T-SPOT.TB = commercial ELISPOT assay; TST = tuberculin skin test.

Appendix D Table 5. Studies of Sensitivity of TST Screening Tests for Tuberculosis (KQ 2), Sensitivity Analysis

First Author, Year	Country (TB Burden ^a)	% Male	Mean Age in Years (SD)	% HIV	% BCG	Other Study Population Comments	TST 5 mm Sensitivity (95% CI, Interval) (N)	TST 10 mm Sensitivity (95% CI, Interval) (N)	TST 15 mm Sensitivity (95% CI, Interval) (N)	Quality Rating
Kang, 2007 ¹⁰²	South Korea (I)	62.5 ^b	Median 55 ^b Range 16 to 81	0	36.1 ^b	Data extracted for subjects with culture confirmation. 20% of study population had risk factor for immunosuppression. No information available on timing of testing with respect to treatment.	-	0.67 (0.55 to 0.77) (67)	-	Poor
Kim, 2011 ⁸¹	South Korea (I)	54.4	Median 49 Range 16 to 94	0	NR	Data extracted for subjects with culture confirmation. QFT testing completed before treatment initiation.	0.70 ^c (0.60 to 0.78) (96)	0.70 ^c (0.60 to 0.78) (96)	-	Poor (TST only)
Li, 2012 ¹¹⁰	China (H)	58.3	46.9 (21.7)	0	33.3	Data extracted for subjects with culture confirmation. Population includes patients who had been treated for 14 days or less.	0.67 (0.50 to 0.80) (36)	-	-	Poor
Memish, 2000 ¹⁰¹	Saudi Arabia (I)	43.4 ^b	Median 38 ^b Range 1 to 78	NR	NR	Data extracted for subjects with culture confirmation, positive AFB smear, or presence of caseating granulomas in histologic sections or cytologic smears with no clinical evidence of other infectious or noninfectious diseases. No information available on timing of testing with respect to treatment.	-	0.83 (0.67 to 0.92) (35)	-	Poor
Ozekinci, 2007 ¹⁰³	Turkey (I)	NR	41 Range 18 to 63	NR	67.4 ^b	Data extracted for subjects with smear or culture confirmation. Treatment received up to 2 weeks prior to testing.	-	0.82 ^d (0.64 to 0.92) (28)	0.82 ^d (0.64 to 0.92) (28)	Poor
Ra, 2011 ⁸⁵	South Korea (I)	42.1	Median 49 Range 22 to 83	0	84.6	Data extracted for subjects with positive AFB smear and culture confirmation. Information not available on timing of testing with respect to treatment. Subjects included 9 patients with prior history of TB and 13 immunosuppressed patients.	-	0.71 (0.47 to 0.87) (17)	-	Poor (TST only)
Shalabi, 2009 ¹⁰⁸	Egypt (I)	73.3	31 (11.1)	0	76.7	Data extracted for subjects with positive AFB smear. No information available on timing of testing with respect to treatment.	-	0.87 (0.70 to 0.95) (30)	-	Poor

Appendix D Table 5. Studies of Sensitivity of TST Screening Tests for Tuberculosis (KQ 2), Sensitivity Analysis

^a TB burden according to World Health Organization classification. (L) Low <10 cases/100,000; (I) Intermediate 10–99 cases/100,000; (H) High >100 cases/100,000.

^b Represents demographics of the overall study population; demographics for subjects eligible for inclusion in analysis were not reported.

^c Estimate represents use of both the 5-mm and 10-mm threshold, which varied by clinical status of the individual tested.

^d Estimate represents use of both the 10-mm and 15-mm threshold, which varied by BCG vaccination status of the individual tested.

AFB = acid fast bacilli; BCG = bacillus Calmette-Guerin; CI = confidence interval; NR = not reported; QFT-G = QuantiFERON TB Gold[®] test (2nd generation test); QFT-GIT = QuantiFERON TB Gold-In-Tube[®] test (3rd generation test); HIV = human immunodeficiency virus; SD = standard deviation; Sn = sensitivity; TB = tuberculosis; TST = tuberculin skin test.

Appendix D Table 6. Studies of Sensitivity of IGRA Screening Tests for Tuberculosis (KQ 2), Sensitivity Analysis

First Author, Year	Country (TB Burden ^a)	% Male	Mean Age in Years (SD)	% HIV	% BCG	Other Study Population Comments	T-SPOT.TB Sensitivity (95% CI, Interval) (N)	QFT-G Sensitivity (95% CI, Interval) (N)	QFT-GIT Sensitivity (95% CI, Interval) (N)	Quality Rating
Eum, 2008 ¹⁰⁴	South Korea (I)	92.0	43.6 (2.5)	NR	NR	Data extracted for subjects with culture confirmation. All subjects received TB treatment for less than 1 week at the time of testing.	-	-	0.76 (0.57 to 0.89) (25)	Poor
Kalantri, 2009 ¹⁰⁶	India (H)	76.0	Range 24 to 45	NR	NR	Data extracted for subjects with positive AFB smear. No information available on timing of testing with respect to treatment.	-	-	0.96 (0.90 to 0.98) (100)	Poor
Kamiya, 2013 ¹¹²	Japan (L)	Younger age group: 49.2 ^b Older age group: 50.0 ^b	Younger age group: 54.0 ^b Older age group: 78.0 ^b	0	NR	Data extracted for subjects with M.tb confirmation from body site samples. No information available on timing of testing with respect to treatment.	-	-	0.88 (0.70 to 0.96) (25)	Poor
Kang, 2007 ¹⁰²	South Korea (I)	62.5 ^b	Median 55 ^b Range 16 to 81	0	36.1 ^b	Data extracted for subjects with culture confirmation. 20% of study population had risk factors for immunosuppression. No information available on timing of testing with respect to treatment.	0.88 (0.78 to 0.94) (67)	0.87 (0.76 to 0.93) (67)	-	Poor
Kobashi, 2009 ¹⁰⁷	Japan (I)	NR	NR	NR	NR	Data extracted for subjects with microbiological confirmation. No information available on timing of testing with respect to treatment.	-	0.82 (0.75 to 0.88) (140)	-	Poor
Li, 2012 ¹¹⁰	China (H)	58.3	46.9 (21.7)	0	33.3	Data extracted for subjects with culture confirmation. Population includes patients who had been treated for 14 days or less.	0.89 (0.75 to 0.96) (36)	-	-	Poor
Ozekinci, 2007 ¹⁰³	Turkey (I)	NR	41 Range 18 to 63	NR	67.4 ^b	Data extracted for subjects with smear or culture confirmation. Treatment received up to 2 weeks prior to testing.	0.93 (0.77 to 0.98) (28)	-	-	Poor

Appendix D Table 6. Studies of Sensitivity of IGRA Screening Tests for Tuberculosis (KQ 2), Sensitivity Analysis

First Author, Year	Country (TB Burden ^a)	% Male	Mean Age in Years (SD)	% HIV	% BCG	Other Study Population Comments	T-SPOT.TB Sensitivity (95% CI, Interval) (N)	QFT-G Sensitivity (95% CI, Interval) (N)	QFT-GIT Sensitivity (95% CI, Interval) (N)	Quality Rating
Palazzo, 2008 ¹⁰⁵	Italy (L)	NR	36 (2)	0	NR	Data extracted for subjects with culture confirmation and positive AFB smear. No information available on timing of testing with respect to treatment.	-	0.50 (0.29 to 0.71) (18)	0.82 (0.59 to 0.94) (17)	Poor
Shrestha, 2011 ¹⁰⁹	Nepal (H)	NR	NR	NR	NR	Data extracted for subjects with positive AFB smear. No information available on timing of testing with respect to treatment.	0.90 (0.74 to 0.97) (30)	-	-	Poor
Turtle, 2012 ¹¹¹	England (I)	53.0 ^b	36 ^b Range 17 to 78	0	NR	Data extracted for HIV negative subjects with culture confirmation. No information available on timing of testing with respect to treatment.	0.82 (0.52 to 0.95) (11)	-	-	Poor

^aTB burden according to World Health Organization classification. (L) Low <10 cases/100,000; (I) Intermediate 10–99 cases/100,000; (H) High >100 cases/100,000.

^b Represents demographics of the overall study population; demographics for subjects eligible for inclusion in analysis were not reported.

AFB = acid fast bacilli; BCG = bacillus Calmette-Guerin; CI = confidence interval; HIV = human immunodeficiency virus; IGRA = interferon gamma release assay; M.Tb = *Mycobacterium tuberculosis*; NR = not reported; QFT-G = QuantiFERON TB Gold[®] test (2nd generation test); QFT-GIT = QuantiFERON TB Gold-In-Tube[®] test (3rd generation test); N = number; SD = standard deviation; TB = tuberculosis; T-SPOT.TB = commercial ELISPOT assay.

Appendix D Table 7. Studies of Specificity of TST Screening Tests for Tuberculosis (KQ 2), Sensitivity Analysis

First Author, Year	Country (TB Burden ^a)	% Male	Mean Age in Years (SD)	% HIV	% BCG	Other Study Population Comments	TST 5 mm Specificity (95% CI, Interval) (N)	TST 10 mm Specificity (95% CI, Interval) (N)	TST 15 mm Specificity (95% CI, Interval) (N)	Quality Rating
Franken, 2007 ¹²⁶	Netherlands (L)	91.8	19.6 (2.8)	NR	8.8	Population is Dutch armed forces recruits. 2 subjects were known to have been treated previously for TB.	-	0.89 (0.83 to 0.93) (153)	0.92 (0.87 to 0.95) (153)	Poor
Ozekinci, 2007 ¹⁰³	Turkey (I)	NR	30 Range 17 to 61	NR	67.4 ^b	Data extracted for subjects with no history of exposure to TB.		0.46 ^c (0.30 to 0.64) (28)		Poor
Shalabi, 2009 ¹⁰⁸	Egypt (I)	58.1	39.4 (12.6)	0	77.4	Data extracted for healthy control subjects.	-	0.84 (0.67 to 0.93) (31)	-	Poor

^a TB burden according to World Health Organization classification. (L) Low <10 cases/100,000; (I) Intermediate 10–99 cases/100,000; (H) High >100 cases/100,000.

^b Represents demographics of the overall study population; demographics for subjects eligible for inclusion in analysis were not reported.

^c Estimate represents use of both the 10-mm and 15-mm threshold, which varied by BCG vaccination status of the individual tested.

BCG = bacillus Calmette-Guerin; CI = confidence interval; HIV = human immunodeficiency virus; M.Tb = Mycobacterium tuberculosis; NR = not reported; SD = standard deviation; TB = tuberculosis; TST = tuberculin skin test.

Appendix D Table 8. Studies of Specificity of IGRA Screening Tests for Tuberculosis (KQ 2), Sensitivity Analysis

First Author, Year	Country (TB Burden ^a)	% Male	Mean Age in Years (SD)	% HIV	% BCG	Other Study Population Comments	T-SPOT.TB Specificity (95% CI, Interval) (N)	QFT-G Specificity (95% CI, Interval) (N)	QFT-GIT Specificity (95% CI, Interval) (N)	Quality Rating
Franken, 2007 ¹²⁶	Netherlands (L)	91.8	19.6 (2.8)	NR	8.8	Population is Dutch armed forces recruits; 2 subjects were known to have been previously treated for TB.	-	-	0.97 (0.93 to 0.99) (171)	Poor
Min, 2013 ⁹⁴	South Korea (I)	57.6	Median 28 Range 23 to 42	NR	75.6	Data extracted for health volunteer study subjects with neither a history of TB treatment nor contact with active TB patients.	-	-	0.94 (0.80 to 0.98) (33)	Poor (Sp) Fair (Sn)
Ozekinci, 2007 ¹⁰³	Turkey (I)	NR	30 Range 17 to 61	NR	67.4 ^b	Data extracted for subjects with no history of exposure to TB.	0.89 (0.73 to 0.96) (28)	-	-	Poor
Palazzo, 2008 ¹⁰⁵	Italy (L)	NR	37 (2)	0	21.0	Data extracted for healthy control subjects.	-	0.94 (0.72 to 0.99) (16)	1.00 (0.78 to 1.00) (14)	Poor

^aTB burden according to World Health Organization classification. (L) Low <10 cases/100,000; (I) Intermediate 10–99 cases/100,000; (H) High >100 cases/100,000.

^bRepresents demographics of the overall study population; demographics for subjects eligible for inclusion in analysis were not reported.

BCG = bacillus Calmette-Guerin; CI = confidence interval; HIV = human immunodeficiency virus; IGRA = interferon gamma release assay; NR = not reported; QFT-G = QuantiFERON TB Gold[®] test (2nd generation test); QFT-GIT = QuantiFERON TB Gold-In-Tube[®] test (3rd generation test); M.Tb = mycobacterium tuberculosis; SD = standard deviation; Sn = sensitivity; Sp = specificity; TB = tuberculosis; T-SPOT.TB = commercial ELISPOT assay.

Appendix D Table 9. Studies of Reliability of Screening Tests for Tuberculosis (KQ 2)

First Author, Year	Country (TB Burden ^a)	% Male	Mean Age in Years (SD)	% HIV	% BCG	Study Population Comments	Test (N)	Reliability Measure	Result	Quality Rating
Cummings 2009 ¹³²	United States(L)	NR	28	NR	7	U.S. HCWs at low risk of TB in a single institution.	QFT-GIT (3-Gen) N=182; N analyzed at 4 weeks=85	Test-retest	2 of 5 positive results on first test were confirmed on subsequent testing At 4 weeks: 85 (47%) of 182 HCWs who had an initial test had the second test; 84 of 85 had consistent results (98.8%)	Poor
Dorman, 2014 ¹²⁷	United States(L)	25	Median 36 (IQR: 28-48)	0.4	9	U.S. HCWs at 4 U.S. health care institutions	T-SPOT.TB and QFT-GIT N=130	Reproducibility	Number of discordant results in participants who had 2 samples drawn simultaneously: QFT-GIT: 10 /172 (5.8%) T-SPOT.TB: 10/153 (6.5%)	Good
								Test-retest	Test-retest at 2 weeks: T-SPOT.TB: 9/111 (8.1%) tests changed from negative to positive and 10/19 (52.6%) changed from positive to negative QFT-GIT:10/134 (7.5%) results changed from negative to positive and 5/15 (33.3%) changed from positive to negative	
Dilektasli 2010 ⁷⁵	Turkey(I)	36.7	39	NR	90.3	Study included multiple groups, including those with pulmonary TB, close contacts of people with TB, and healthy controls.	T-SPOT.TB N=91	Interrater reliability	Interrater reliability ^b = 96% (k=0.92; p<0.05) Manual read versus automated Elispot reader = 85.8% (k=0.73; p<0.05)	Fair
Franken, 2009 ¹²⁸	Netherlands	NR	NR	NR	NR	Immigrants that were close contacts of smear-positive TB patients.	T-SPOT.TB N=313	Interrater reliability ^b	Kappas for agreement among 6 raters were all above 0.6.	Fair
Mancuso, 2012 ¹²³	United States(L)	66	21.8	NR	3.5	Population is U.S. military recruits at low risk of exposure to TB.	TST N=1826	Interrater reliability ^b	Kappa=0.79	Fair

Appendix D Table 9. Studies of Reliability of Screening Tests for Tuberculosis (KQ 2)

First Author, Year	Country (TB Burden ^a)	% Male	Mean Age in Years (SD)	% HIV	% BCG	Study Population Comments	Test (N)	Reliability Measure	Result	Quality Rating
O'Shea, 2014 ¹³¹	Nepal(H)	166	NR; range 18-21	0.9	63	Nepalese military recruits who had left Nepal and recently entered the U.K.	T-SPOT-TB and QFT-GIT N=166	Test-retest	Test-retest at 1 week: T-SPOT-TB: kappa for agreement between initial test and retest: 0.66 (95% CI, 0.50 to 0.83) QFT-GIT: kappa for agreement between initial test and retest: 0.48 (95% CI, 0.26 to 0.7)	Fair
Villarino 2000 ¹¹⁴	United States(L)	37-81 ^c	50	NR	NR	2 study populations: persons with pulmonary TB and those at low risk of exposure to TB.	TST (PPD S2) N=1189	Interrater reliability ^b	Kappa=0.52 to 0.78 across all groups	Fair
Villarino 1999 ¹¹³	United States(L)	38	26	NR	NR	Persons at low risk for TB.	TST (PPD S1) N=127	Interrater reliability ^b	Kappa=0.69	Fair
Whitworth, 2012 ¹²⁹	United States(L)	49	NR; all ≥18	NR	28	Subjects with self-reported positive TST recruited from U.S Air Force and CDC staff located in San Antonio, TX, and Atlanta, GA	QFT-GIT(3Gen) N=91	Interlaboratory reliability ^d	Across 3 labs, 7/ 91 (7.7%) subjects had discordant results (none had indeterminate results); Kappas of pairwise lab sample comparisons ranged from 0.87, 0.89, and 0.93	Good
Whitworth, 2014 ¹³⁰	United States(L)	46	NR; all ≥18	NR	21	Subjects with self-reported positive TST recruited from U.S Air Force and CDC staff located in San Antonio, TX, and Atlanta, GA	QFT-GIT(3Gen) N=146	Interrater reliability	2 samples from each participant both processed via manual read and automated ELISA; across all 4 tests, 88.6% were concordant (16% concordant positive and 72.6% concordant negative) and 11% were discordant. Discordance by method: Automated vs. automated: 4.8% (kappa 0.85) Manual vs. manual: 6.9% (kappa 0.80) Automated vs. manual: 3.4% to 9.0% across comparisons (kappa 0.73- 0.90)	Good

^a TB burden according to World Health Organization classification. (L) Low <10 cases/100,000; (I) Intermediate 10–99 cases/100,000; (H) High >100 cases/100,000.

^b Agreement between first and second observer.

^c Among the population with pulmonary TB 81 percent were male. Among the population at low risk of exposure to TB, 37 percent were male.

^d To measure interlaboratory reliability, three tubes of blood were collected from each subject so that the assay could be completed at three different labs noted to have “extensive experience and demonstrated proficiency.”

Appendix D Table 9. Studies of Reliability of Screening Tests for Tuberculosis (KQ 2)

BCG = bacillus Calmette-Guerin; CDC= Centers for Disease Control and Prevention; HIV = human immunodeficiency virus; HCW = health care worker; IQR= intraquartile range; NR = not reported; N = number analyzed; NR = not reported; QFT-GIT = QuantiFERON TB Gold-In-Tube[®] test (3rd generation test); PPD-S1 or S2 = purified protein derivative standard 1 or standard 2; SD = standard deviation; TB = tuberculosis; TST = tuberculin skin test; U.K. = United Kingdom; U.S. = United States

Appendix D Table 10. Characteristics of Included Randomized, Controlled Trials (KQs 3, 5), Main Analysis

First Author, Year Trial Name N	Drug, Dose X Duration (N)	Followup	Population	LTBI Confirmed?	Country; TB Burden ^a	TB Risk Factors	Mean (Range) Age	% F	% Non- white	% BCG	Quality
Menzies, 2004 ¹⁴³ 116	RIF 10mg/kg of body weight, up to 600 mg/day x 4 months; up to 20 weeks, if needed, depending on missed doses (58). INH 5mg/kg, up to 300 mg/day x 9 months; up to 43 weeks, if needed, depending on missed doses (58).	16-20 weeks 36-43 weeks Duration of both arms depending upon whether treatment was extended due to missed doses.	≥18 years Positive TST following Canadian guidelines; physician recommend 9 INH for LTBI. <5% HIV positive	Yes (TST≥5-, 10- and 15-mm, based upon risk status under Canadian guidelines). Abnormal CXR: 29 (50) 31 (53)	Canada: low	Contact with active TB case: 10 (17) 10 (17) COB high TB ^b : 45 (78) 48 (83) Randomization stratified by TB risk (high if HIV infected close contacts with active TB ^c , or fibronodular changes CXR; low to moderate for all others).	32.9 (10.8 SD) 34.8 (13.0 SD)	38 50	NR	Yes: 21 Unknown: 19 Yes: 28 Unknown: 21	Fair
Menzies, 2008 ¹³³ 847	RIF 10 mg/kg of body weight, up to 600 mg/day x 4 months (420). INH 5 mg/kg, up to 300 mg/day x 9 months (427).	4 months 9 months	18 years or older with a documented positive TST and if physician recommend INH for LTBI following national or international guidelines; 9 university hospitals (7 were in Canada).	Yes	Canada; low ^d Saudi Arabia; intermediate, Brazil; high	HIV infection: 6 (1) 7 (2) Abnormal chest radiograph: 117 (28) 105 (25) Contact with active TB case: 131 (31) 135 (32) Recent immigrant: 29 (7) 33 (8) Of the Canadian participants (who comprised 80% of the sample), born in high TB incidence country: 227 (54) 235 (55)	Age 18-34: 229 (55) 242 (57) Age≥35: 191 (45) 185 (43)	48 47	NR	Yes: 54 47 Unk: 33 25	Good

Appendix D Table 10. Characteristics of Included Randomized, Controlled Trials (KQs 3, 5), Main Analysis

First Author, Year Trial Name N	Drug, Dose X Duration (N)	Followup	Population	LTBI Confirmed?	Country; TB Burden ^a	TB Risk Factors	Mean (Range) Age	% F	% Non-white	% BCG	Quality
Sterling, 2011 ^{134e} PREVENT TB 6,886	RPT 900 mg + INH 900 mg/week x 12 weeks (3,556) INH 300 mg/day x 36 weeks (3,330)	33 months	≥18 years, TST or IGRA positive excluding HIV-positive patients; Close contacts of patients with culture-confirmed TB, recent converters, and small percentage with fibrosis.	Yes ^e	U.S., Canada, Brazil, and Spain; low to high	Close contact within the past 2 years with patient with culture-confirmed TB.	Median: 37 ^e	45.8 ^e	42.9 ^e	NR	Fair
Thompson, 1982 ¹³⁵ IUAT 27,830	INH 300 mg x 12 weeks (6,956). INH 300 mg x 24 weeks (6,965). INH 300 mg x 52 weeks (6,919). Placebo (6,990).	5 years	Age 20-64 ^f with fibrotic pulmonary lesions ^g not previously treated with anti-TB meds.	Yes (6 mm or greater Mantoux test) ^h	7 European countries ⁱ low to intermediate	NR	Median 50 years (NR); 38% were between 55 and 65 years	47	NR	NR	Good (for KQ 3) Fair (for KQ 5)
White, 2012 ¹⁴⁴ 364	RIF 600 mg/day x 4 months; up to 6 months, if needed, depending on missed doses for a total of 120 doses (180). INH 900 mg 2x week x 9 months; up to 12 months, if needed, depending on missed doses for a total of 76 doses (184).	16-18 weeks 36-40 weeks Duration of both arms depended upon whether treatment was extended due to missed doses, unless necessary to	Inmates ≥18 years in the San Francisco City and County Jail diagnosed with LTBI at jail entry.	Yes, diagnosis method NR	U.S.: low	Foreign-born: 278 (76); p=0.5 Jailed before: 255 (70); p=0.80 Drug/alcohol problem: 186 (51); p=0.21	<35: 258 (71) ≥35: 106 (29)	7	92	NR	Fair

Appendix D Table 10. Characteristics of Included Randomized, Controlled Trials (KQs 3, 5), Main Analysis

First Author, Year Trial Name N	Drug, Dose X Duration (N)	Followup	Population	LTBI Confirmed?	Country; TB Burden ^a	TB Risk Factors	Mean (Range) Age	% F	% Non- white	% BCG	Quality
		restart (RIF, restart if missed doses >2 weeks); INH restart if missed doses >1 month									

^aTB burden according to World Health Organization classification. Low <10 cases/100,000; intermediate 10–99 cases/100,000; high >100 cases/100,000.

^bCountries classified as high TB according to TB incidence as suggested by the World Health Organization.

^cNumber of subjects with who have been in close contact with an individual with active tuberculosis unspecified.

^dAlthough TB burden in Canada is low, 54–55 percent of the Canadian participants (a total of 462 participants) were born in countries with high TB incidence.

^eData extracted from supplemental data provided by personal communication source for eligible study subgroup (HIV-negative subjects with IGRA or TST confirmation).

^fInclusion criteria initially limited to age 20–64, but a few persons are included outside these limits.

^gDefined as well-delineated radiographic lesions of probable tuberculous origin, usually in the upper half of the lung, which had been stable during the year prior to entry. For participants, the lesions had been known to exist for a median of 8 years (range 11 months to 58 years).

^hMedian induration of participants was 15 mm (range 6–90 mm).

ⁱCzechoslovakia (low), Finland (low), Germany (low), Hungary (intermediate), Poland (intermediate), Romania (intermediate), Yugoslavia (low-intermediate).

BCG = bacillus Calmette-Guérin vaccine; CXR = chest x-ray; F = female; HIV = human immunodeficiency virus; IGRA = Interferon gamma release assays; INH = isoniazid; IUAT = International Union Against Tuberculosis and Lung Disease; kg = kilogram; LTBI = latent tuberculosis infection; mg = milligram; N = sample size; NR = not reported; RIF = rifampin; SD = standard deviation; TB = tuberculosis; TST = tuberculin skin test; Unk = unknown.

Appendix D Table 11. Results of Included Randomized, Controlled Trials for Benefits (KQ 3), Main Analysis

First Author, Year Trial Name N	Drug, Dose X Duration (N)	Active TB Disease, N (%)	Transmission, N (%)	Quality of Life	Overall Mortality, N (%)	Disease-Specific Mortality, N (%)
Menzies, 2008 ¹³³ 847	RIF 10 mg/kg of body weight, up to 600 mg/day x 4 months (420). INH 5 mg/kg, up to 300 mg/day x 9 months (427).	NR	NR	NR	0 (0) 1 (0.2)	0 (0) 0 (0)
Sterling, 2011 ¹³⁴	RPT 900 mg + INH 900 mg/week x 12 weeks (3,556).	5 (0.15) 10 (0.32)	NR	NR	30 (0.8) 34 (1.0)	NR
PREVENT TB 6,886	INH 300 mg/day x 36 weeks (3,330).	Rate per 100 person years 0.05 0.12 Difference in cumulative TB rate -0.17 Upper bound of the 95% CI, (%) 0.07				
Thompson, 1982 ¹³⁵	INH 300 mg x 12 weeks (6,956).	76 (1.1) 34 (0.5) 24 (0.3)	NR	NR	All groups combined: 1124 (4.0)	Due to tuberculosis: 0 (0.00) 0 (0.00)
IUAT 27,830	INH 300 mg x 24 weeks (6,965).	97 (1.4)			NR by group	0 (0.00) 3 (0.042)
	INH 300 mg x 52 weeks (6,919).	Percent reduction compared with placebo ^{a, b} 21 65				
	Placebo (6,990).	75 NA (reference)				
		RR compared with 52 weeks of INH ^c 3.1 1.4 1.0 (reference) 4.0				
		Benefit-to-risk ratio by regimen (cumulative TB cases prevented/ cumulative hepatitis cases incurred), 5				

Appendix D Table 11. Results of Included Randomized, Controlled Trials for Benefits (KQ 3), Main Analysis

First Author, Year Trial Name N	Drug, Dose X Duration (N)	Active TB Disease, N (%)	Transmission, N (%)	Quality of Life	Overall Mortality, N (%)	Disease-Specific Mortality, N (%)
		years: 1.2 2.6 ^{d, e} 2.1 NA (reference)				

^a Percent reduction by size of lesion: for lesions <2 cm², 20, 66, 64, and NA (reference); for lesions >2 cm², 24, 67, 89, and NA (reference).

^b When limited to “completer-compliers” the percent reductions were 31, 69, 93, and NA (reference), respectively.

^c The differences between the 52-week and 24-week INH regimens and between the 12-week INH and placebo were not statistically significant (0.20>P >0.10). All other interregimen differences were statistically significant.

^d RR by size of lesion: for lesions <2 cm², 2.2, 1.0, 1.0 (reference), and 2.8; for lesions >2 cm², 6.8, 2.9, 1.0 (reference), and 8.9.

^e When limited to “completer-compliers” the RRs were 9.4, 4.3, 1.0 (reference), and 13.6, respectively.

INH = isoniazid; IUAT = International Union Against Tuberculosis and Lung Disease; mg = milligram; N = sample size; NA = not applicable; NR = not reported; RR = relative risk; TB = tuberculosis.

Appendix D Table 12. Results of Included Randomized, Controlled Trials for Harms (KQ 5), Main Analysis

First Author, Year Trial Name N	Drug, Dose X Duration (N)	DC Due to AEs, N (%)	Hepatotoxicity, N (%)	Mortality From Hepatotoxicity, N (%)	Gastrointestinal, N (%)	Other Specific AEs, N (%) ^a
Menzies, 2004 ¹⁴³ 116	RIF 10 mg/kg of body weight, up to 600 mg/day x 4 months; up to 20 weeks, if needed, depending on missed doses (58) INH 5 mg/kg, up to 300 mg/day x 9 months; up to 43 weeks, if needed, depending on missed doses (58)	2 (3.4) 8 (13.8) RR: 0.25 (95% CI, 0.1 to 1.1)	0 (0) 3 (5.2) Drug-induced hepatitis after 74, 105, and 137 doses of INH	0 (0) 0 (0)	Severe nausea and vomiting: 4 (3.4) ^b	Other overall AEs 2 (3.4) 5 (8.6) Calculated RR: 0.40 (95% CI, 0.08 to 1.98) Persistent debilitating fatigue: 2 (1.7) Rash: 1 (0.8) ^c
Menzies, 2008 ¹³³ 847	RIF 10 mg/kg of body weight, up to 600 mg/day x 4 months (420) INH 5 mg/kg, up to 300 mg/day x 9 months (427)	Among protocol-adherent: 16 (3.8) 24 (5.6) Subtotal for any grade 3 or 4 AE ^{d-h} 7 (1.7) 17 (4.0) RD -2.3% (95% CI, -5.0 to -0.1) Subtotal for any grade 1 or 2 AE: ^{i-m} 9 (2.1) 7 (1.6) RD 1% (95% CI, 1.0 to 3.0)	Grade 3 or 4 hepatotoxicity ^d 3 (0.7) 16 (3.7) RD -3.1% (95% CI, -5.0 to -1.0)	0 (0) 0 (0)	Minor AEs reported “similar” between groups GI intolerance (grade 1 or 2 AEs): ^l 1 (0.2) 2 (0.5) Calculated RR: 0.51 (95% CI, 0.05 to 5.59)	Hematologic (grade 3 or 4 AEs): ^d 2 (0.5) 1 (0.2) Calculated RR: 2.0 (95% CI, 0.19 to 22.34) Drug interaction (grade 3 or 4 AEs): ^h 1 (0.2) 0 (0) Calculated RR: 3.05 (95% CI, 0.13 to 74.66) Rash (grade 3 or 4 AEs) ^e 1 (0.2) 0 (0) Calculated RR: 3.05 (95% CI, 0.13 to 74.66) Rash (grade 1 or 2 AEs) ^l 8 (1.9) 5 (1.2) Calculated RR: 1.63 (95% CI, 0.54 to 4.93)

Appendix D Table 12. Results of Included Randomized, Controlled Trials for Harms (KQ 5), Main Analysis

First Author, Year Trial Name N	Drug, Dose X Duration (N)	DC Due to AEs, N (%)	Hepatotoxicity, N (%)	Mortality From Hepatotoxicity, N (%)	Gastrointestinal, N (%)	Other Specific AEs, N (%) ^a
Sterling, 2011 ^{134b} PREVENT TB 6,886	RPT 900 mg + INH 900 mg/week x 12 weeks (3,556) INH 300 mg/day x 36 weeks (3,330)	DC due to adverse drug reaction: 186 (5.2) 136 (4.1) Calculated RR: 1.28 (95% CI, 1.03 to 1.59)	Grade 3 toxicity: ^p 176 (4.9) 184 (5.5) Calculated RR: 0.90 (95% CI, 0.73 to 1.10) Grade 4 toxicity: ^p 34 (1.0) 35 (1.1) Calculated RR: for Grade 3 or 4 toxicity: 0.90 (95% CI, 0.75 to 1.08)	Grade 5 (death): NR 30 (0.8) 34 (1.0) Calculated RR: 0.83 (95% CI, 0.51 to 1.35)		Possible hypersensitivity: 146 (4.1) 17 (0.5) Calculated RR: 8.04 (95% CI, 4.88 to 13.26)
Thompson, 1982 ¹³⁵ IUAT 27,830	INH 300 mg x 12 weeks (6,956) NH 300 mg x 24 weeks (6,965) INH 300 mg x 52 weeks (6,919) Placebo (6,990)	Overall DC: INH (8.1) Placebo (5.8) ¹⁴⁷ Due to AEs (GI distress, liver disease, or gallbladder disease): INH (1.8) Placebo (1.2) ¹⁴⁷ DC due to liver disease: INH (0.4) Placebo (0.1) ¹⁴⁷	Hepatitis: INH 99 ^q (0.5) Placebo 7 (0.1) Cumulative excess hepatitis rates per 1000 cases for INH: 12 weeks: 2.5 24 weeks: 3.6 52 weeks: 5.2 Calculated number of cases: 12 weeks: 24 24 weeks: 32 52 weeks: 43 Hepatitis cases prevented per 1000 persons by reducing duration of INH from 52 weeks to: 24 weeks, 1.6 12 weeks, 2.7	2 (0.03) 0 (0.00) 1 (0.01) 0 (0.00) 0.14 per 1,000 persons receiving INH 0 cases in placebo group. Calculated RR: 2.35 (95% CI, 0.12 to 45.46)	GI distress resulting in stopping: INH (1.2) Placebo (0.9) ¹⁴⁷ Calculated RR: 1.33 (95% CI, 1.01 to 1.75)	Gallbladder disease resulting in stopping: INH (0.2) Placebo (0.2)

Appendix D Table 12. Results of Included Randomized, Controlled Trials for Harms (KQ 5), Main Analysis

First Author, Year Trial Name N	Drug, Dose X Duration (N)	DC Due to AEs, N (%)	Hepatotoxicity, N (%)	Mortality From Hepatotoxicity, N (%)	Gastrointestinal, N (%)	Other Specific AEs, N (%) ^a
White, 2012 ¹⁴⁴ 364	RIF 600 mg/day x 4 months; up to 6 months, if needed, depending on missed doses for a total of 120 doses (180) INH 900 mg 2x/week x 9 months; up to 12 months, if needed, depending on missed doses for a total of 76 doses (184)	2 (1.1) 0 (0)	Grade 3 for LFT was AST or ALT >5.0–10.0 times ULN ≥3 elevated LFT: 8 (4.4) 21 (11.4)	0 (0) 0 (0)	GI 16 (9) 9 (10) Calculated RR: 1.82 (95% CI, 0.82 to 4.01)	Other AEs ^c Rash/pruritis 16 (9) 12 (6) Calculated RR: 1.36 (95% CI, 0.66 to 2.80) Central nervous system 6 (3) 20 (11) Calculated RR: 0.31 (95% CI, 0.13 to 0.75) Allergic reaction 1 (1) 0 (0) Calculated RR: 3.07 (95% CI, 0.13 to 74.78) Other ^e 13 (7) 14 (8) Calculated RR: 0.95 (95% CI, 0.46 to 1.96)

^a No studies reported peripheral neuropathy or development of drug-resistant TB outcomes.

^b Other adverse events not presented by drug regimen, but for entire population.

^c Categories are not mutually exclusive; participants could experience symptoms in more than one body system category. Therefore, the number and percentage represent the number of participants and the percentage of the study group or total that had an adverse event in the category.

^d Liver aminotransferase levels that increased to 5 to 10 or 3 to 10 times the upper limit of normal in the presence of compatible symptoms met criteria for grade 3 hepatotoxicity, whereas those that exceeded 10 times the upper limit of normal met criteria for grade 4 toxicity.

^e Criteria for a grade 3 rash is a rash that affects 100 percent of body surface area or mucus membranes, conjunctivae are affected, vital signs are abnormal (fever or low blood pressure), or there is wheezing.

^f Neutrophil counts <1.00 to 0.50 x 10⁹ cells/L or platelet counts <50 to 25 x 10⁹ cells/L met the criteria for grade 3 hematologic effects, whereas Neutrophil counts that exceeded 0.50 x 10⁹ cells/L or platelet counts greater than 25 x 10⁹ cells/L met the criteria for grade 4.

^g Protracted nausea and vomiting or severe abdominal pain that disrupts daily life (for example, cannot sleep), severe diarrhea (more than 5 bowel movements per day) met the criteria for a grade 3 gastrointestinal adverse event.

^h Under drug interaction grade 3, drug interaction was noted and therapy was modified repeatedly but eventually successful; patient did not have any untoward clinical effect, and LTBI therapy was continued. Under grade 4, care providers unable to adjust therapy successfully to achieve therapeutic effects; LTBI therapy was discontinued.

ⁱ Liver aminotransferase levels that increased to 1 to 3 times the upper limit of normal in the presence of symptoms suggestive of hepatotoxicity (nausea, anorexia, vomiting, fatigue, abdominal pain) met criteria for grade 1, whereas levels 1 to 5 times the upper limit of normal with no symptoms met criteria for grade 2 toxicity.

Appendix D Table 12. Results of Included Randomized, Controlled Trials for Harms (KQ 5), Main Analysis

^j Criteria for a grade 1 involves itching only or limited to limbs, trunk, or face only; no abnormality of vital signs and no mucosal or conjunctival involvement. Grade 2 rash affects limbs and trunk or more than 50 percent of total body surface area or rash is confluent in areas.

^k Neutrophil levels <1.50 to 1.00×10^9 cells/L or platelet counts <100 to 50×10^9 cells/L met the criteria for grades 1-2.

^l Some stomach upset with nausea or loss of appetite, but no vomiting and no change in bowel habits met that criteria for a grade 1 gastrointestinal adverse event.

^m Under drug interaction grade 1, a potential drug interaction was noted, but no change in therapy was required and neither short- nor long-term effect detected. Under grade 2, a potential drug interaction was noted, but after an initial change in therapy, no further problems and therapy did not have to be changed.

ⁿ Data extracted from supplemental data provided by personal communication source for eligible study subgroup (HIV-negative subjects with IGRA or TST confirmation).

^o Other category includes symptoms such as appetite loss, muscle/body pain, fatigue, weight loss, malaise, cold symptoms, change of urine color, fever, and eye redness.

^p Common toxicity criteria version 2.0. Bethesda, MD: Cancer Therapy Evaluation Program, 1999 (http://www.eortc.be/services/doc/ctc/ctcv20_4-30-992.pdf).

^q The total number of hepatotoxicity cases among isoniazid patients was calculated based on the cumulative excess hepatitis rates per 1000 cases for INH presented in the paper.

AE = adverse events; ALT = alanine aminotransferase; AST = aspartate aminotransferase; CI = confidence interval; DC = discontinuation; GI = gastrointestinal; INH = isoniazid; kg = kilogram; LFT = liver function test; mg = milligram; N = sample size; NR = not reported; RD = risk difference; RIF = rifampin; RPT = rifapentine; RR = relative risk; TB = tuberculosis; ULN = upper limit of normal.

Appendix D Table 13. Characteristics of Randomized, Controlled Trials Used Only in Sensitivity Analyses (KQs 3, 5)

First Author, Year Trial Name N	Drug, Dose X Duration (N)	Followup	Population	LTBI Confirmed?	Country; TB Burden ^a	TB Risk Factors	Mean (Range) Age	% F	% Non- white	% BCG	Quality
Bailey, 1974 ¹⁴⁵ 178	INH 300 mg + 50 mg pyridoxine (vitamin B6)/day x 12 months (85) Control (93)	Rolling enrollment, follow-up ranged between 1- 10 months	Adult tuberculin- reactive employees in a U.S. hospital (1,900-bed general; 80 beds for TB patients) considered for anti-TB Chemo- prophylaxis with INH Normal levels (SGOT <40): Normal: 63 (74) 62 (67) Abnormal levels (SGOT ≥40): 22 (26) 31 (33) ^b	No, but positive TST (5 TU PPD); cut-offs unspecified	U.S.: low	Health care workers: 178 (100%)	38.54 (13.78 SD) 40.56 (11.39 SD)	74.2	70.2	NR	Fair
Bush, 1965 ⁴³ All subjects: 2,238 ≥15 years 1309 ≥20 years 1140	INH 250 mg/day x 12 months (571) Placebo (569)	1 year after end of medication regimen	Subjects ≥20 years who were HH contacts of active TB cases Total HHs: 328 322 HHs ≥1 cases active TB: 220	No, but chest film and TST (5 TU PPD-S); 90% of the adults with ≥5 mm TST	Japan: low	HH contacts (all ages) who lived with an adult index case >9 months: (78.5) (78.9)	Subjects 20-49 years: 818 Subjects 50+ years: 322	59.4 (subjects ≥20 years)	NR; appro- ximate- ly 100%	NR	Fair

Appendix D Table 13. Characteristics of Randomized, Controlled Trials Used Only in Sensitivity Analyses (KQs 3, 5)

First Author, Year Trial Name N	Drug, Dose X Duration (N)	Followup	Population	LTBI Confirmed?	Country; TB Burden ^a	TB Risk Factors	Mean (Range) Age	% F	% Non- white	% BCG	Quality
			189								
			Study population ≥20 years: 569 571								
			Study population ≥15 years: 646 663								
Byrd, 1977 ¹⁴⁶ 120	Round 1: INH 300 mg/day x 3 months (60) Placebo (60) Round 2: INH 300 mg/day x 3 months (60) ^c .	3 months	≥18 years with baseline SGOT <20 for Round 1; SGOT levels unspecified for Round 2 ATS criteria for undergoing chemopro- phylaxis	Positive TST (Tween stabilized intermediate strength); cutoffs unspecified; CXR	U.S.: low	NR	Subjects <30 years: 19 22 Subjects 30-39 years: 23 25 Subjects >40 years 18 13	26.7	30	NR	Fair
Falk, 1978 ^{41,148} 7,036	INH 300 mg/day x 2 years (2,166). INH 300 mg/day x 1 year, followed by placebo x 1 year (2,553). Placebo daily x 2 years (2,317)	7 years	Veterans with pulmonary TB classified as inactive ^{d,e}	NR; required to have inactive pulmonary TB	U.S.: low	NR	78% were 30-50; 16% were 51-70	2	24 23 21	NR	Fair

Appendix D Table 13. Characteristics of Randomized, Controlled Trials Used Only in Sensitivity Analyses (KQs 3, 5)

First Author, Year Trial Name N	Drug, Dose X Duration (N)	Followup	Population	LTBI Confirmed?	Country; TB Burden ^a	TB Risk Factors	Mean (Range) Age	% F	% Non-white	% BCG	Quality
Ferebee, 1963 ⁴⁴ 27,924 patients (566 psychiatric wards randomized); 25,210 patients included in morbidity analyses ^{f, g}	INH 4-7 mg/kg/day (average of 5mg/kg) ^h x 12 months (14,407 in randomized sample; 12,884 in morbidity analyses). Placebo x 12 months (13,517; 12,326).	10 years	Those residing in mental institutions	No (not required to have positive TST to be included; 57% had positive TST, ≥5 mm)	U.S. ⁱ : low	Residing in mental institutions 100% Abnormal CXR 1216 (9.5%) 1071 (8.7%) Tuberculin positive 7242 (56%) 7253 (59%)	Males: 48 Females: 54 Listed overall range: 2-80+ years Proportion <15 years of age: 63% 58%	51 54	13 11	NR	Fair
Veening, 1968 ⁴² 261	INH 600 mg (8-10mg/kg) x 4 months, then 400 mg (5-7mg/kg) until 1 year (133) ^j . Placebo (128).	7 years	Military service members with Mantoux conversion after exposure to an active case	Yes	Netherlands ; low	All were close contact of an active case	Mean NR; military recruits 18 to 20 years old at baseline	0 (0) 0 (0)	NR	NR	Poor

^a TB burden according to World Health Organization classification. Low <10 cases/100,000; intermediate 10–99 cases/100,000; high >100 cases/100,000.

^b Chi-square = 0.8479; *P*=0.3 (not significant, according to authors).

^c Placebo subjects received treatment after initial 3-month trial.

^d Determine by NTA diagnostic standards current at that time.

^e TB had been inactive for 5 years or more in 95 percent of participants.

^f Morbidity analyses did not include patients who moved to a new ward and crossed over; only included people who took either INH or placebo.

^g All data entered for Ferebee 1963 for subsequent rows of this table are based on the N included in morbidity analyses.

^h Those 15 and older received 300 mg/day.

ⁱ Wisconsin, Georgia, Michigan, and Massachusetts.

^j This is a higher dose than is currently recommended by CDC.

ATS = American Thoracic Society; BCG = bacillus Calmette-Guérin vaccine; CXR = chest x-ray; F = female; HH = household; INH = isoniazid; kg = kilogram; LTBI = latent tuberculosis infection; mg = milligram; N = sample size; NR = not reported; PPD = purified protein derivative; PPD-S = polysorbate 80 stabilized solution of tuberculin purified protein derivative; SD = standard deviation; SGOT = serum glutamic-oxalacetic transaminase; TB = tuberculosis; TST = tuberculin skin test; TU = tuberculin units; U.S. = United States.

Appendix D Table 14. Results of Randomized, Controlled Trials Used Only in Sensitivity Analyses for Benefits (KQ 3)

First Author, Year Trial Name N	Drug, Dose X Duration (N)	Active TB Disease, N (%)	Transmission, N (%)	Quality of Life	Overall Mortality, N (%)	Disease-Specific Mortality, N (%)
Bush, 1965 ⁴³ All subjects: 2,238 ≥15 years 1,309 ≥20 years 1,140	INH 250 mg/day x 12 months (569). Placebo (571).	All subjects (adults and children): 8 (0.73) ^a 11 (0.96) ^b Subjects ≥15 years: 4 (0.60) 7 (1.08)	NR	NR	NR NR	NR NR
Falk, 1978 ^{41,148} 7,036	INH 300 mg daily x 2 years (2166). INH 300 mg daily x 1 year, followed by placebo x 1 year (2,553). Placebo daily x 2 years (2,317).	17 (0.8) 20 (0.8) 26 (1.1) Among those with no prior treatment for TB (2,389 subjects): 8 5 15 Reactivators who had received previous treatment: 9 (0.6) 15 (0.9) 11 (0.7) Reactivators who had received previous adequate ^c treatment 5 13 5 ^d	NR	NR	Total deaths: 357 Rate: 4/1,000 6.5/1,000 4.4/1,000 <i>p</i> =0.0001 for INH 1 year vs. placebo; NR for other comparisons	2 (0.03) deaths from TB (both received INH; 1 occurred at the 6th month of INH therapy and 1 in a patient who completed only 2 months of INH and died 11 months later)

Appendix D Table 14. Results of Randomized, Controlled Trials Used Only in Sensitivity Analyses for Benefits (KQ 3)

First Author, Year Trial Name N	Drug, Dose X Duration (N)	Active TB Disease, N (%)	Transmission, N (%)	Quality of Life	Overall Mortality, N (%)	Disease-Specific Mortality, N (%)
Ferebee, 1963 ⁴⁴ 27,924 patients (566 psychiatric wards randomized); 25,210 patients included in morbidity analyses	INH 4-7 mg/kg/day (average of 5mg/kg) x 12 months (14,407 in randomized sample; 12,884 analyzed). Placebo x 12 months (13,517; 12,326).	Cases diagnosed during first 15 months of the trial: Total 4 (0.03) 21 (0.17) Among those with abnormal CXR 3 (0.25) 14 (1.31) Among those who were tuberculin positive 0 (0.0) 7 (0.11) Tuberculin negative 0 (0.0) 0 (0.0) Unknown tuberculin status 1 (0.11) 0 (0.0) Cases developing after medication year Total 15 (0.12) 30 (0.24) Among those with abnormal CXR 5 (0.41) 9 (0.84) Among those who were tuberculin positive 5 (0.08) 17 (0.26)	NR	NR	During treatment year, among the full randomized sample ^e : 752 (5.2) 611 (4.5) Among patients who took only one medication (excluding crossovers): 695 (5.4%) 547 (4.4%)	NR
Ferebee, 1963 ⁴⁴ (continued)		Among the placebo group: Cases appearing by May 1962 in those ≥20 years Males <150 lbs 21 (1.14) Males ≥150 lbs 3 (0.20)				

Appendix D Table 14. Results of Randomized, Controlled Trials Used Only in Sensitivity Analyses for Benefits (KQ 3)

First Author, Year Trial Name N	Drug, Dose X Duration (N)	Active TB Disease, N (%)	Transmission, N (%)	Quality of Life	Overall Mortality, N (%)	Disease-Specific Mortality, N (%)
		Females <130 lbs 8 (0.46)				
		Females ≥130 lbs 2 (0.10)				
		Among the placebo group, cases based on TB infection status: Initial tuberculin reactions <5 mm 4 (0.10)				
		Initial tuberculin ≥5 mm 24 (0.37)				
		Abnormal roentgenogram 23 (2.15)				
Veening, 1968 ⁴² 261	INH 600 mg (8-10 mg/kg) x 4 months, then 400 mg (5-7 mg/kg) until 1 year (133). Placebo (128).	1 year: 1 (0.8) 9 (7.0) 4 years: 1 (0.8) 12 (9.4) 7 years: 1 (0.8) 12 (9.4)	NR	NR	NR	NR

^a No cases first 3 months after starting treatment; one case between months 6 and 11; seven cases 11 months or more after starting treatment. Days index case in home by new cases: 1-60 days: 2; 61-180 days: 4; 181-270 days: 1; 270-300 days: 1.

^b No cases first 3 months after starting treatment; 2 cases between month 6 and 11; 9 cases 11 months or more after starting treatment. Days index case in home by new cases: 1-60 days: 2; 61-180 days: 2; 181-270 days: 5; 270-300 days: 2.

^c Adequate treatment was defined as at least 18 months of therapy with two drugs.

^d Rate of reactivation was 7.3/1,000 for those with adequate prior chemotherapy, and 12.7/1,000 for those with inadequate or no prior chemotherapy.

^e Deaths in wards participating in the trial during the year prior to the trial: INH 801 (5.6), placebo 698 (5.2). Change in percent of deaths from year prior to the trial to the medication year: INH -0.4 percent; placebo -0.7 percent.

CXR = chest x-ray; INH = isoniazid; kg = kilogram; lbs = pounds; mg = milligram; N = sample size; NR = not reported; TB = tuberculosis.

Appendix D Table 15. Results of Randomized, Controlled Trials Used Only in Sensitivity Analyses for Harms (KQ 5)

First Author, Year Trial Name N	Drug, Dose X Duration (N)	DC due to AEs, N (%)	Hepatotoxicity, N (%)	Mortality From Hepatotoxicity, N (%)	Gastrointestinal, N (%)	Other Specific AEs, N (%) ^a
Bailey, 1974 ¹⁴⁵ 178	INH 300 mg + 50 mg pyridoxine (vitamin B ₆)/day x 12 months (85). Control (93).	10 (11.8) NR	SGOT elevations ≥100 mU/ml ^b : 10 (11.8) ^c 0 (0.0) ^d Among INH: with ≥100 mU/ml: Average age: 50.2 (SD 12.09) with <100 mU/ml: Average age: 36.99 (SD 13.3)	NR NR	NR	NR
Bush, 1965 ⁴³ All subjects: 2,238 ≥15 years 1,309 ≥20 years 1,140	INH 250 mg/day x 12 months (569). Placebo (571).	All ages: 8 (0.7) 12 (1.1)	NR NR	NR NR	GI (all ages): 8 (0.7) 3 (0.3) Calculated RR: 2.68 (95% CI, 0.71 to 10.04)	Other AEs (all ages) Rash: 0 (0.0) 3 (0.3) Calculated RR: 0.14 (95% CI, 0.01 to 2.77) Other, unspecified: 0 (0.0) 6 (0.5) Calculated RR: 0.08 (95% CI, 0.004 to 1.37)
Byrd, 1977 ¹⁴⁶ 120	Round 1: INH 300 mg/day x 3 months (60). Placebo (60). Round 2: INH 300 mg/day x 3 months (60).	Round 1 7 (11.7) 1 (1.7) Round 2 3 (5.0)	SGOT elevations ≥100 IU: Round 1: 3 (5.0) 0 (0.0) Round 2: 1 (1.7) SGOT elevations ≥30 IU, overall: Round 1: (18.3) (6.9) <i>p</i> <05 Round 2: (22.4)	Round 1: 0 (0) 0 (0) Round 2: NR	Round 1: Nausea: 2 (3.3) 1 (1.7) Calculated RR: 2.00 (95% CI, 0.19 to 21.47) Clay-colored Stools: 6 (10.0) 3 (5.0) Calculated RR: 2.00 (95% CI, 0.52 to 7.63) Anorexia:	Other adverse effects; Round 1: Muscle aching: 18 (30.0) 17 (28.3) Calculated RR: 1.06 (95% CI, 0.61 to 1.85) Joint aching: 14 (23.3) 11 (18.3) Calculated RR: 1.27 (95% CI, 0.63 to 2.57) Flulike symptoms:

Appendix D Table 15. Results of Randomized, Controlled Trials Used Only in Sensitivity Analyses for Harms (KQ 5)

First Author, Year Trial Name N	Drug, Dose X Duration (N)	DC due to AEs, N (%)	Hepatotoxicity, N (%)	Mortality From Hepatotoxicity, N (%)	Gastrointestinal, N (%)	Other Specific AEs, N (%) ^a
			$p < .025$		5 (8.3)	8 (13.3)
			SGOT		5 (8.3)	10 (16.7)
			Elevations		Calculated RR: 1.00	Calculated RR: 0.80
			≥ 30 IU, by month:		(95% CI, 0.31 to 3.28)	(95% CI, 0.34 to 1.89)
			Round 1 ^e :		Round 2:	Fever:
			Month 1:		NR for any	4 (6.7)
			(5.0)		gastrointestinal	4 (6.7)
			(3.3)		events	Calculated RR: 1.00
			$p = NS$			(95% CI, 0.26 to 3.82)
			Month 2:			Chills:
			(14.0)			9 (15.0)
			(3.4)			5 (8.3)
			$p < .05$			Calculated RR: 1.8
			Month 3:			(95% CI, 0.64 to 5.06)
			(14.0)			Skin rash:
			(1.7)			7 (11.7)
			$p < .025$			6 (10.0)
			Round 2:			Calculated RR: 1.17
			NR by month			(95% CI, 0.42 to 3.27)
						Dark urine:
						6 (10.0)
						0 (0.0)
						Calculated RR: 13.0
						(95% CI, 0.75 to
						225.75)
						Yellow cast
						to sciera:
						1 (1.7)
						1 (1.7)
						Calculated RR: 1.00
						(95% CI, 0.06 to
						15.62)
						Round 2: NR for any
						other specific AE

Appendix D Table 15. Results of Randomized, Controlled Trials Used Only in Sensitivity Analyses for Harms (KQ 5)

First Author, Year Trial Name N	Drug, Dose X Duration (N)	DC due to AEs, N (%)	Hepatotoxicity, N (%)	Mortality From Hepatotoxicity, N (%)	Gastrointestinal, N (%)	Other Specific AEs, N (%) ^a
Falk, 1978 ^{41,148} 7,036	INH 300 mg/day x 2 years (2,166). INH 300 mg/day x 1 year, followed by placebo x 1 year (2,553). Placebo daily x 2 years (2,317).	NR	1 (0.01) taking INH (NR which group); mild hepatitis that resolved after stopping INH 0 in placebo group	NR	NR (reported that nausea occurred equally among the 3 regimens)	Rash INH regimens: 44 (0.9) Placebo: 8 (0.3) Calculated RR: 2.7 (95% CI, 1.27 to 5.73)
Ferebee, 1963 ⁴⁴ 27,924 patients (566 psychiatric wards randomized); 25,210 patients included in these harms analyses	INH 4-7 mg/kg/day (average of 5mg/kg) x 12 months (14,407 in randomized sample; 12,884 in harms analyses). Placebo x 12 months (13,517; 12,326).	Made "sick" from pills 141 (1.1) 58 (0.47)	NR	NR	NR	NR

^a No studies reported peripheral neuropathy or development of drug-resistant TB outcomes.

^b Statistical analysis yielded a chi-square value of 9.15 and a $p < 0.01$.

^c Comparison using Fisher's exact test for 2 x 2 contingency tables. Number of subjects not provided.

^d Reported rate of elevated transaminases for the placebo group is based on the 90 individuals who had baseline SGOT < 100, not on the full placebo group (N=93).

^e Liver aminotransferase levels that increased to 5 to 10 or 3 to 10 times the upper limit of normal in the presence of compatible symptoms met criteria for grade 3 hepatotoxicity, whereas those that exceeded 10 times the upper limit of normal met criteria for grade 4 toxicity.

AE = adverse event; DC = discontinuation; INH = isoniazid; IU = international units; mg = milligram; mU/ml = milli units/milliliter; N = sample size; NR = not reported; NS = not sufficient; SD = standard deviation; SGOT = serum glutamic-oxalacetic transaminase; TB = tuberculosis.

Appendix E Table 1. Quality Ratings for Studies of Accuracy and Reliability of Screening Tests for Tuberculosis (KQ 2)

First Author, Year	Were selection criteria clearly described?	Was the spectrum of patients representative of the patients who will receive the test in PC?	Were withdrawals from the study explained (post-enrollment)?	Was the screening test relevant and adequately described?	Was the reference test performed regardless of screening test result?	Were the reference standard and screening test interpreted independently? (i.e., each test interpreted blinded to the result of the other)	Were methods for calculating accuracy (e.g. sensitivity/specificity) clearly reported and valid?	Did the study provide raw data on indeterminate results or enough information to understand how indeterminate results were handled?	Quality Rating
Adetifa, 2007 ⁵⁸	Partially	NA	Partially	Partially	Yes	NR	NA	Yes	Fair
Ak, 2009 ⁷⁰	Yes	NA	NA	Yes	Yes	NR	Yes	Yes	Good
Bellef, 2002 ¹¹⁶	Partially	Yes	Yes	Yes	NA	NA	NA	NA	Fair
Berkel, 2005 ⁵⁴	Yes	No	NA	No	No	NR	Partially	NA	Fair
Bienek, 2009 ¹²¹	Yes	Partially	Yes	Yes	NA	NR	Partially	Yes	Fair
Bocchino, 2010 ⁷⁴	Partially	NA	NA	No	Yes	NR	NA	Yes	Fair
Boyd, 2011 ⁷⁹	Yes	NA	NA	Yes	Yes	Yes	Yes	Yes	Good
Bua, 2007 ¹¹⁹	No	NR	NA	Partially	NA	NR	NA	Yes	Fair
Chee, 2008 ⁶⁴	Yes	NA	Yes	Yes	Yes	NR	NA	Yes	Good
Cho, 2011 ⁸⁰	Yes	NA	NA	Partially	Yes	Yes	Yes	Yes	Good
Cummings, 2009 ¹³²	No	Partially	No	Yes	NA	NR	No	Yes	Poor
Dewan, 2007 ⁵⁹	Yes	NA	Partially	Yes	Yes	NR	Yes	Yes	Fair
Dilektasli, 2010 ⁷⁵	Yes	Partially	Yes	Partially	Yes	Partially	NA	Yes	Fair
Dorman, 2014 ¹²⁷	Yes	No	Yes	Yes	NA	NR	Yes	Yes	Good
Erdem, 2014 ⁹⁹	No	NA	NA	Yes	No	NR	No	NR	Fair
Eum, 2008 ¹⁰⁴	Partially	NA	No	Yes	Yes	NR	Partially	No	Poor
Feng, 2013 ⁹¹	Partially	NA	Yes	Yes	Partially	NR	NA	Yes	Fair
Fietta, 2003 ⁵³	Yes	Yes	NA	Yes	NR	NR	NA	NA	Fair
Franken, 2007 ¹²⁶	No	No	No	Partially	NA	NR	Partially	No	Poor
Franken,	Yes	Partially	NA	Yes	NA	NR	NA	Yes	Fair

Appendix E Table 1. Quality Ratings for Studies of Accuracy and Reliability of Screening Tests for Tuberculosis (KQ 2)

First Author, Year	Were selection criteria clearly described?	Was the spectrum of patients representative of the patients who will receive the test in PC?	Were withdrawals from the study explained (post-enrollment)?	Was the screening test relevant and adequately described?	Was the reference test performed regardless of screening test result?	Were the reference standard and screening test interpreted independently? (i.e., each test interpreted blinded to the result of the other)	Were methods for calculating accuracy (e.g. sensitivity/specificity) clearly reported and valid?	Did the study provide raw data on indeterminate results or enough information to understand how indeterminate results were handled?	Quality Rating
2009 ¹²⁸									
Goletti, 2006 ⁵⁶	Yes	NA	NR	Partially	Yes	Yes	Yes	Partially	Fair
Harada, 2008 ⁶⁵	Yes	NA	NA	Yes	Yes	NR	NA	Yes	Good
Higuchi, 2009 ⁷¹	Partially	NA	Yes	Yes	Yes	NR	NA	Yes	Fair
Janssens, 2007 ⁶⁰	Yes	NA	NA	Yes	Yes	NR	Yes	No	Fair
Jeon, 2013 ⁹²	Yes	NA	NA	Yes	Yes	NR	NA	No	Fair
Kalantri, 2009 ¹⁰⁶	No	NA	NA	Yes	Yes	NR	NA	No	Poor
Kamiya, 2013 ¹¹²	No	NA	Partially	Yes	Partially	NR	Yes	NA	Poor
Kang, 2007 ¹⁰²	Partially	NA	NA	Partially	Partially	NR	No	No	Poor
Kang, 2005 ⁵⁵	Partially	NA	NR	Yes	Partially	No	Partially	Yes	Fair
Katsenos, 2010 ¹²²	Yes	Partially	NA	Yes	NA	Yes	Yes	Yes	Good
Kim, 2014 ¹⁰⁰	Yes	NA	Yes	Yes	Yes	NR	Yes	Yes	Good
Kim, 2013 ⁹³	Partially	Yes	NA	Yes	Partially	NR	Yes	Yes	Fair
Kim, 2011 ⁸¹	Yes	NA	NA	Yes	Yes	NR	NA	Partially	Good (QFT-GIT) Poor (TST)
Kobashi, 2008 ⁶⁶	Partially	NA	Yes	Yes	Yes	NR	NA	Yes	Fair
Kobashi, 2009 ¹⁰⁷	No	NA	NA	Partially	Yes	NR	NA	No	Poor
Kobashi, 2009 ⁷²	Partially	NA	NA	Yes	Yes	NR	No	Partially	Fair
Kobashi, 2008 ⁶⁷	Yes	NA	NA	Yes	Yes	NR	NA	Yes	Good
Kobashi,	Partially	NA	NA	Yes	Yes	NR	NA	Yes	Fair

Appendix E Table 1. Quality Ratings for Studies of Accuracy and Reliability of Screening Tests for Tuberculosis (KQ 2)

First Author, Year	Were selection criteria clearly described?	Was the spectrum of patients representative of the patients who will receive the test in PC?	Were withdrawals from the study explained (post-enrollment)?	Was the screening test relevant and adequately described?	Was the reference test performed regardless of screening test result?	Were the reference standard and screening test interpreted independently? (i.e., each test interpreted blinded to the result of the other)	Were methods for calculating accuracy (e.g. sensitivity/specificity) clearly reported and valid?	Did the study provide raw data on indeterminate results or enough information to understand how indeterminate results were handled?	Quality Rating
2008 ⁶⁸									
Kobashi, 2012 ⁸⁸	Yes	NA	Yes	Partially	Yes	NR	Yes	Yes	Fair
Lai, 2011 ⁸³	Partially	NA	NA	Partially	Yes	NR	NA	Yes	Fair
Lai, 2011 ⁸²	Partially	No	NA	Yes	Yes	NR	Yes	Yes	Fair
Lee, 2012 ⁸⁹	Partially	NA	NA	Yes	Yes	Yes	Yes	Yes	Good
Lee, 2011 ⁹⁷	Partially	No	NA	Partially	Yes	NR	NA	NR	Fair
Legesse, 2010 ⁷⁶	Yes	No	NA	Yes	Yes	NR	Yes	Yes	Fair
Lempp, 2015 ¹²⁴	No	NA	NA	Yes	No	NR	No	NR	Fair
Li, 2012 ¹¹⁰	Partially	NA	NA	Partially	No	Partially	Yes	Yes	Poor
Losi, 2007 ⁶¹	Partially	NA	NA	Partially	Yes	NR	Yes	Partially	Fair
Lui, 2011 ⁸⁴	Yes	No	NA	Yes	Partially	Partially	Yes	Yes	Fair
Mancuso, 2012 ¹²³	Partially	No	Yes	Yes	NA	Yes	Partially	No	Fair
Mazurek, 2007 ⁶²	Yes	NA	Yes	Yes	Yes	NR	Yes	Yes	Good
Mazurek, 2007 ¹²⁰	Yes	Yes	Yes	Yes	NA	NR	No	No	Fair
Mazurek, 2001 ¹¹⁵	Yes	Yes	NA	Yes	NA	NA	NA	NA	Good
Memish, 2000 ¹⁰¹	No	NA	NA	No	NR	NR	NA	NA	Poor
Metcalfe, 2010 ⁷⁷	Yes	NA	Yes	Yes	Partially	NR	Partially	Yes	Fair
Min, 2013 ⁹⁴	No	NR	NA	Yes	Yes	NR	Yes	Partially	Poor (Sp) Fair (Sn)
O'Shea, 2014 ¹³¹	Yes	No	NA	Yes	Yes	NR	Yes	Partially	Fair

Appendix E Table 1. Quality Ratings for Studies of Accuracy and Reliability of Screening Tests for Tuberculosis (KQ 2)

First Author, Year	Were selection criteria clearly described?	Was the spectrum of patients representative of the patients who will receive the test in PC?	Were withdrawals from the study explained (post-enrollment)?	Was the screening test relevant and adequately described?	Was the reference test performed regardless of screening test result?	Were the reference standard and screening test interpreted independently? (i.e., each test interpreted blinded to the result of the other)	Were methods for calculating accuracy (e.g. sensitivity/specificity) clearly reported and valid?	Did the study provide raw data on indeterminate results or enough information to understand how indeterminate results were handled?	Quality Rating
Ozekinci, 2007 ¹⁰³	Partially	Yes	NA	No	Yes	NR	No	Yes	Poor
Pai, 2007 ⁶³	Yes	NA	Yes	Yes	Yes	NR	Yes	Yes	Good
Painter, 2013 ⁵¹	Yes	NA	Partially	Yes	Yes	Yes	Partially	No	Fair
Palazzo, 2008 ¹⁰⁵	Partially	Partially	No	No	Yes	NR	Partially	No	Poor
Park, 2009 ⁷³	Partially	Partially	Yes	Partially	Partially	NR	Partially	Yes	Fair
Qian, 2013 ⁹⁵	Yes	NA	NA	Yes	Yes	NR	NA	No	Fair
Ra, 2011 ⁸⁵	Partially	No	NA	Partially	Yes	NR	No	Yes	Fair (QFT-G) Poor (TST)
Ruhwald, 2011 ⁸⁶	Yes	Partially	NA	Yes	Yes	NR	NA	Yes	Good
Seibert, 1991 ⁵²	Partially	NA	Partially	Yes	Yes	NR	NA	NA	Fair
Shalabi, 2009 ¹⁰⁸	Partially	NR	NA	No	Yes	NR	NA	NA	Poor
Shrestha, 2011 ¹⁰⁹	No	NA	NA	Partially	Yes	NR	NA	Yes	Poor
Soysal, 2008 ⁶⁹	Yes	Partially	No	Yes	Yes	NR	Partially	Partially	Fair
Taggart, 2006 ¹¹⁸	Yes	Partially	NA	Yes	NA	NR	Partially	No	Fair
Taggart, 2004 ¹¹⁷	Partially	Yes	Yes	Yes	NA	NA	NR	NA	Fair
Taki-Eddin, 2012 ⁹⁰	Partially	NA	NA	Yes	Yes	NR	NA	NR	Fair
Tan, 2010 ⁷⁸	Partially	NA	NA	Yes	Yes	NR	NA	Yes	Fair
Tsiouris, 2006 ⁵⁷	Yes	NA	NR	Yes	Yes	Yes	Yes	Yes	Good

Appendix E Table 1. Quality Ratings for Studies of Accuracy and Reliability of Screening Tests for Tuberculosis (KQ 2)

First Author, Year	Were selection criteria clearly described?	Was the spectrum of patients representative of the patients who will receive the test in PC?	Were withdrawals from the study explained (post-enrollment)?	Was the screening test relevant and adequately described?	Was the reference test performed regardless of screening test result?	Were the reference standard and screening test interpreted independently? (i.e., each test interpreted blinded to the result of the other)	Were methods for calculating accuracy (e.g. sensitivity/specificity) clearly reported and valid?	Did the study provide raw data on indeterminate results or enough information to understand how indeterminate results were handled?	Quality Rating
Turtle, 2012 ¹¹¹	No	NA	Partially	Partially	NR	NR	No	No	Poor
Villarino, 2000 ¹¹⁴	Partially	Partially	Yes	Partially	NA	Yes	NA	Yes	Fair
Villarino, 1999 ¹¹³	Partially	Partially	Yes	Yes	NA	Partially	NA	Partially	Fair
Walsh, 2011 ⁸⁷	Yes	NA	NA	Partially	NR	NR	No	Yes	Fair
Wang, 2013 ⁹⁶	Yes	NA	NA	Yes	Yes	NR	Yes	No	Fair
Whitworth, 2014 ¹³⁰	Partially	Yes	NA	Yes	NA	NR	Yes	Yes	Fair
Whitworth, 2012 ¹²⁹	Partially	NA	NA	Yes	Yes	NR	Yes	Yes	Good
Włodarczyk, 2014 ⁹⁸	Partially	Partially	Yes	Yes	NA	NR	Yes	Yes	Good

Good: Relevant and adequately described study populations for the outcome of interest (i.e., Sensitivity, Specificity), screening test well described in terms of test procedures followed and threshold used for a “positive” or “negative” test, credible reference standard used for outcome of interest (i.e., Sensitivity or Specificity), generally interprets reference standard independently of screening test, outcomes clearly reported and valid, handles indeterminate results in a reasonable manner.

Fair: Mostly includes a relevant and adequately described study population for the outcome of interest (i.e., Sensitivity, Specificity), screening test described although may include some ambiguity about test procedures followed or threshold for a “positive” or “negative” test, credible reference standard mostly used for outcome of interest (i.e., Sensitivity or Specificity), interpretation of reference standard may or may not be independent of screening test, outcomes mostly clearly reported although may have some ambiguity regarding how indeterminate results were handled.

Poor: Has fatal flaw such as study population not appropriate for outcome of interest (i.e., Sensitivity, Specificity), screening test improperly administered or not at all described, use of noncredible reference standard, reference and screening test not independently assessed, outcomes not clearly or accurately reported with no information about how indeterminate tests were handled.

NA = not applicable; NR = not reported; QFT-GIT = QuantiFERON TB Gold-In-Tube (3rd Generation) assay; Sn = sensitivity; Sp = specificity; TST = tuberculin skin test.

Appendix E Table 2. Quality Ratings for Randomized, Controlled Trials (KQs 3, 5): Main Analysis, Part 1

First Author, Year Trial name N	Was random- ization adequate?	Was allocation concealment adequate?	Were groups similar at baseline?	Was adherence to the intervention adequate?	What was the overall attrition?	What was the differential attrition?	Did the study have differential attrition or overall high attrition raising concern for bias?	Did the study have crossovers or contamination raising concern for bias?
Menzies, 2004 ¹⁴³ 116	Yes	Partially	Yes	Yes, RIF: 53 (91) took 80% of doses, 50 (86) took more than 90% of doses within 20 weeks INH: 44 (76) took 80% doses; 36 (62) took 90% doses 43 weeks 80% doses: RR: 1.2 (95% CI, 1.02 to 1.4) 90% of doses: RR: 1.4 (95% CI, 1.1 to 1.7)	Did not complete: 19 (16.4) Dropout/default: 9 (7.8) RR: 0.5 (95% CI, 0.1 to 1.9)	Total did not complete: RIF: 5 (9) INH: 14 (24) Dropout/default: RIF: 3 (4) INH: 6 (10) RR: 0.5 (95% CI, 0.1 to 1.9)	Partially	No
Menzies, 2008 ¹³³ 847	Yes	Yes	Yes	Yes	Not included in primary analyses for serious AEs: 8 (0.9%) Stopped therapy early and were followed; nonprotocol- adherent: 205 (24%) Stopped therapy early and were followed; protocol- adherent: 45 (5.3%) Did not complete therapy: 264 (31%)	Not included in primary analyses for serious AEs: RIF 2 (0.5%) INH 6 (1.4%) Stopped therapy early and were followed; nonprotocol- adherent: RIF 72 (17%) INH 133 (31%) Stopped therapy early and were followed; protocol- adherent:	No	No

Appendix E Table 2. Quality Ratings for Randomized, Controlled Trials (KQs 3, 5): Main Analysis, Part 1

First Author, Year Trial name N	Was random- ization adequate?	Was allocation concealment adequate?	Were groups similar at baseline?	Was adherence to the intervention adequate?	What was the overall attrition?	What was the differential attrition?	Did the study have differential attrition or overall high attrition raising concern for bias?	Did the study have crossovers or contamination raising concern for bias?
						RIF 17 (4.0%) INH 28 (6.6%) Did not complete therapy: RIF 92 (22%) INH 172 (40%)		
Sterling, 2011 ^{134a} PREVENT TB 6,886	Partially	NR	Yes	Yes	Treatment completion: ^a 2895 (80.8%) 2264 (68.2%)	Differential treatment completion: ^a 12.6%	Partially	No
Thompson, 1982 ¹³⁵ IUAT 27,830	Yes	Yes	Unclear	Yes	5-year followup not complete for 781 (2.8%)	<5%	No	No
White, 2012 ¹⁴⁴ 364	Yes	Partially	Yes	No, nearly 1/2 participants started on either INH or RIF were lost to followup by transfer to another facility or deportation Adherence higher for those who remained in jail: RIF: (79) INH: (83)	Did not complete: 257 (70.6)	Did not complete: RIF: 120 (66.7) INH: 137 (74.5) Lost/ withdrawn: RIF: 33 (18.3) INH: 44 (23.9) Deported/ transferred: RIF: 85 (47.2) INH: 93 (50.5) Withdrawn by physician: RIF: 2 (1.1) INH: 0 (0)	Yes	No

^a Data extracted from supplemental data provided by personal communication source for eligible study subgroup (HIV-negative subjects with IGRA or TST confirmation).
AE = adverse event; CI = confidence interval; INH = isoniazid; IUAT = International Union Against Tuberculosis and Lung Disease; N = sample size; RIF = rifampin; RR = relative risk.

Appendix E Table 3. Quality Ratings for Randomized, Controlled Trials (KQs 3, 5): Main Analysis, Part 2

First Author, Year Trial name N	Were outcome measurements equal, valid, and reliable?	Were patients masked?	Were providers masked?	Were outcome assessors masked?	Was the duration of followup adequate to assess the outcome?	Was an appropriate method used to handle missing data?	Did the study use an ITT analysis?	Did the study use acceptable statistical methods?	Quality Rating	Comments (Explain Poor Ratings)
Menzies, 2004 ¹⁴³ 116	Yes	No	No	No	No	Yes	Yes	Yes	Fair	Open label; authors state unblinded study justified because the primary study outcome, treatment completion, was likely strongly influenced by duration of therapy. Primary outcome % prescribed doses taken as measured by electronic device in the pill container cap; patient compliance may be overestimated. Duration of treatment may have influenced judgement of severity of more subjective AEs (e.g. fatigue, nausea).
Menzies, 2008 ¹³³ 847	Yes	No	No	Yes, blinded review panel	Yes	Yes	Yes	Yes	Good	Open label, but used fairly rigorous methods with masked review panel to ascertain AEs.
Sterling, 2011 ¹³⁴ PREVENT TB 6,886	Yes	NR	NR	NR	Yes	Yes	Yes	Yes	Fair	Masking unclear and higher overall attrition.
Thompson, 1982 ¹³⁵ IUAT 27,830	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Good (For KQ3) Fair (For KQ5)	

Appendix E Table 3. Quality Ratings for Randomized, Controlled Trials (KQs 3, 5): Main Analysis, Part 2

First Author, Year Trial name N	Were outcome measurements equal, valid, and reliable?	Were patients masked?	Were providers masked?	Were outcome assessors masked?	Was the duration of followup adequate to assess the outcome?	Was an appropriate method used to handle missing data?	Did the study use an ITT analysis?	Did the study use acceptable statistical methods?	Quality Rating	Comments (Explain Poor Ratings)
White, 2012 ¹⁴⁴ 364	Yes	No	No	No	No	Yes	Yes	Yes	Fair	Open label; nearly 1/2 participants started on either INH or RIF were lost to followup by transfer to another facility or deportation. However, those who remained in jail had higher adherence.

AE = adverse event; INH = isoniazid; ITT = intent to treat; IUAT = International Union Against Tuberculosis and Lung Disease; RIF = rifampin.

Appendix E Table 4. Additional Quality Ratings for Randomized, Controlled Trials for Harms (KQ 5): Main Analysis

First Author, Year Trial Name N	Were harms prespecified and defined?	Were ascertainment techniques for harms adequately described?	Were ascertainment techniques for harms equal, valid, and reliable?	Was duration of followup adequate for harms assessment?	Harms Quality Rating	Comments (Explain Poor Quality Ratings)
Menzies, 2004 ¹⁴³ 116	Yes	Yes	Partially	No	Fair	Followup likely insufficient; some AEs subject to judgement of severity (e.g., fatigue, nausea)
Menzies, 2008 ¹³³ 847	Yes	Yes	Yes	Yes	Good	
Sterling, 2011 ¹³⁴ PREVENT TB 6,886	Yes	Yes	Yes	Yes	Fair	
Thompson, 1982 ¹³⁵ IUAT 27,830	Partially; INH-induced hepatotoxicity was prespecified; NR how it was defined; unclear for other harms	Partially; specific criteria for ascertaining/confirming hepatotoxicity NR	They were equal. Unclear how valid and reliable (dispensary staff were told to be particularly alert for symptoms of INH-induced hepatitis; participants were advised to call the dispensary if they had any unexpected reactions)	Yes	Fair	
White, 2012 ¹⁴⁴ 364	Yes	Yes	Yes	No	Fair	Nearly 1/2 participants started were lost to followup by transfer to another facility or deportation, thus unable to adequately track harms.

INH = Isoniazid; IUAT = International Union Against Tuberculosis; N = sample size; NR = not reported.

Appendix E Table 5. Quality Ratings for Randomized, Controlled Trials Used Only in Sensitivity Analysis (KQs 3, 5), Part 1

First Author, Year Trial name N	Was random- ization adequate?	Was allocation concealment adequate?	Were groups similar at baseline?	Was adherence to the intervention adequate?	What was the overall attrition?	What was the differential attrition?	Did the study have differential attrition or overall high attrition raising concern for bias?	Did the study have crossovers or contamination raising concern for bias?
Bailey, 1974 ¹⁴⁵ 178	Yes; random assignment, although no details on methods	No; control group not given placebo	Partially, although baseline characteristics sparse. Control group had higher proportion of subjects with elevated SGOT levels at baseline	Adherence levels NR	NR	NR	Attrition rates not report, thus unable to assess bias	Partially
Bush, 1965 ⁴³ All subjects: 2,238 ≥15 years 1309 ≥20 years 1140	Partially, randomized by HH instead of by individual	Partially, since randomized by HH, plausible families realized whether they were under treatment regime	Partially, subjects randomized by HH. Baseline characteristics details sparse	Yes, Completed 9 months of drug regiment; all subjects (all ages): 780 (68.3) 748 (68.3) Completed 12 months of drug regiment; all subjects: 557 (48.8) 609 (55.6)	Total discontinued treatment; all subjects 441	Total discontinued treatment; all subjects 215 (18.8) 226 (20.6) Reasons for discontinuing treatment; all subjects: Moved or left household 46 (4.0) 48 (4.4) Not interested: 48 (4.2) 62 (5.6) Suspected TB: 2 (0.2) 2 (0.2) Non-TB illness: 13 (1.1) 14 (1.3) Busy: 18 (1.6) 17 (1.5) Forgot: 32 (2.6) 31 (2.8) Other reason:	Partially	Yes; randomization at HH level

Appendix E Table 5. Quality Ratings for Randomized, Controlled Trials Used Only in Sensitivity Analysis (KQs 3, 5), Part 1

First Author, Year Trial name N	Was random- ization adequate?	Was allocation concealment adequate?	Were groups similar at baseline?	Was adherence to the intervention adequate?	What was the overall attrition?	What was the differential attrition?	Did the study have differential attrition or overall high attrition raising concern for bias?	Did the study have crossovers or contamination raising concern for bias?
						42 (3.7) 37 (3.4) No reason given: 6 (0.5) 3 (0.3)		
Byrd, 1977 ¹⁴⁶ 120	Yes	Round 1: Yes, double-blinded; only chief hospital pharmacist knew content of pills. Same appearance/imprint for both intervention and control Round 2: No, no concealment	Round 1: Yes Round 2: No, baseline SGOT levels not given for baseline; patients had progressed 3 months in to disease	Round 1 and 2: No, index of how treatment compliance based on positive INH in monthly urine specimens. Patients could feasibly register positive if medication taken shortly before followup visit and not throughout month, thus not a true indicator of 30-day compliance	NR	NR	Attrition rates NR, thus unable to assess bias	Round 1: No Round 2: Partially, given prior 3 months as placebo arm
Falk, 1978 ^{41,148} 7,036	Yes	Yes	Unclear. Article reports the groups were “balanced,” but no further details given by group other than race information.	Yes (78% completed more than 12 months of pill taking; 75% completed 19 months or more of pill taking; 73% completed the full 24 months)	19% (81% were observed for 5 years or more) 19% = 1337 participants	NR. Stated distribution of factors related to stopping pill-taking were “similar” among the groups, but no information about differences/similarities in completion of followup or for missing data	Yes	Unclear

Appendix E Table 5. Quality Ratings for Randomized, Controlled Trials Used Only in Sensitivity Analysis (KQs 3, 5), Part 1

First Author, Year Trial name N	Was random- ization adequate?	Was allocation concealment adequate?	Were groups similar at baseline?	Was adherence to the intervention adequate?	What was the overall attrition?	What was the differential attrition?	Did the study have differential attrition or overall high attrition raising concern for bias?	Did the study have crossovers or contamination raising concern for bias?
Ferebee, 1963 ⁴⁴ 27,924 patients (566 psychiatric wards randomized); 25,210 patients included in morbidity analyses	Yes	Yes	Yes	Reported completion of over 39 weeks (from records of ward attendants): INH 70.9% Placebo 76.4% Percentage of ward attendant records accepted as “probably correct”: INH 66% Placebo 69%	Subjects crossing over were dropped from most analyses (except for some of the mortality analyses): n=2714 (9.7%) For the 12- month examination, health status was unknown for <0.05% of participants in the morbidity analyses	Subjects crossing over who were dropped from most analyses: 1.8% For the 12-month examination, unknown health status: <0.05%	No	Yes; 1191 (8.8%) patients from wards randomized to placebo spent part of the year on INH (e.g., transferred to a ward where INH was being given) and 1523 (10.6%) randomized to INH also received some placebo
Veening, 1968 ⁴² 261	Unclear. No details given other than that they were divided “at random”	NR	NR, no data provided to allow comparability of groups at baseline	NR	Missing data for 51 (19.5%) at 7 years; unclear how much missing data for earlier time points, but implied 0% at 1 year; and 43 (16.5%) left military service in the first half of year 2, but unclear how many of those were lost to followup	NR	Yes, moderate concern for risk of attrition bias for overall attrition for the later time points (4 years and 7 years); unclear for differential attrition	No

HH = household; INH = isoniazid; N = sample size; NR = not reported; SGOT = serum glutamic-oxalacetic transaminase; TB = tuberculosis.

Appendix E Table 6. Quality Ratings for Randomized, Controlled Trials Used Only in Sensitivity Analysis (KQs 3, 5), Part 2

First Author, Year Trial name N	Were outcome measurements equal, valid, and reliable?	Were patients masked?	Were providers masked?	Were outcome assessors masked?	Was the duration of followup adequate to assess the outcome?	Was an appropriate method used to handle missing data?	Did the study use an ITT analysis?	Did the study use acceptable statistical methods?	Quality Rating	Comments (Explain Poor Quality Ratings)
Bailey, 1974 ¹⁴⁵ 178	Yes	No	No	No	No	Partially	No	No	Fair	Adherence to treatment and data points for followup unclear
Bush, 1965 ⁴³ All subjects: 2,238 ≥15 years 1309 ≥20 years 1140	No, data by age group varied	Yes	Yes	Yes	Yes	Yes	No	No	Fair	Randomly assigned double-blind study; subjects randomized by HH groups; low retention; data presented inconsistently
Byrd, 1977 ¹⁴⁶ 120	Round 1: Yes Round 2: No, unclear baseline SGOT levels	Round 1: Yes Round 2: No	Round 1: Yes Round 2: No	Round 1: Yes Round 2: No	Round 1: No Round 2: No	Round 1: Partially Round 2: Partially	Round 1: Partially Round 2: Partially	Round 1: Partially Round 2: Partially	Fair	Round 1 randomly assigned double-blind study; Round 2 prospective cohort study Study data difficult to discern in some outcomes (whether Round 1 and 2 treatment results were or were not combined; percentages given without n); adherence to treatment questionable in both Rounds
Falk, 1978 ^{41,148} 7,036	Yes, they were equal; used a masked referee to review all reports of TB reactivation; somewhat unclear what the exact criteria were (reports only)	Yes	Yes	Yes	Yes	No (appears nothing done to consider missing data; NR how much missing data there really was because no information about attrition by study group)	Yes	Unclear	Fair	Good methods of randomization, allocation concealment, and masking; inadequately described statistical analyses; inadequate description of baseline characteristics to allow for assessment of comparability of groups at baseline (but methods of

Appendix E Table 6. Quality Ratings for Randomized, Controlled Trials Used Only in Sensitivity Analysis (KQs 3, 5), Part 2

First Author, Year Trial name N	Were outcome measurements equal, valid, and reliable?	Were patients masked?	Were providers masked?	Were outcome assessors masked?	Was the duration of followup adequate to assess the outcome?	Was an appropriate method used to handle missing data?	Did the study use an ITT analysis?	Did the study use acceptable statistical methods?	Quality Rating	Comments (Explain Poor Quality Ratings)
	that it was “by x-ray, positive bacteriology, or both”)					or about how much followup time subjects contributed)				randomization and allocation concealment were good and the trial is very large). Overall attrition almost 20%; information NR about differential attrition; moderate concern for risk of bias due to attrition
Ferebee, 1963 ⁴⁴ 27,924 patients (566 psychiatric wards randomized); 25,210 patients included in morbidity analyses	Yes	Yes	Yes	Yes	Yes	NR (but seems that nothing was done to handle missing data)	No, dropped crossovers from analyses (more like a per protocol analysis)	Did not report data allowing a true ITT analysis, given how crossovers were handled	Fair	
Veening, 1968 ⁴² 261	Unclear, methods of determining cases of active TB not clearly specified (they did x-rays every 2 months during the earlier part of the study; after 7 years they did x-rays, tracheal lavage, and urine, but article does not report criteria for case definition)	Yes	Yes	NR (study was reported as double-blind, but no information about outcome assessor masking)	Yes	NR, appears nothing done to handle missing data	Yes	Yes	Poor	Very limited reporting to allow risk of bias assessment in this 2-page publication; concern for risk of selection bias, attrition bias, confounding, and measurement bias given the very limited information provided; unclear if groups similar at baseline; unclear methods for randomization, allocation concealment, masking, and ascertainment of outcomes

HH = household; ITT = intent to treat; N = sample size; NR = not reported; SGOT = serum glutamic-oxalacetic transaminase; TB = tuberculosis

Appendix E Table 7. Additional Quality Ratings for Randomized, Controlled Trials Used Only in Sensitivity Analysis for Harms

First Author, Year Trial name N	Were harms prespecified and defined?	Were ascertainment techniques for harms adequately described?	Were ascertainment techniques for harms equal, valid, and reliable?	Was duration of followup adequate for harms assessment?	Harms Quality Rating	Comments (Explain Poor Quality Ratings)
Bailey, 1974 ¹⁴⁵ 178	Yes	Yes	Yes	No	Fair	Adherence to treatment and data points for followup unclear
Bush, 1965 ¹⁴³ All subjects: 2,238 ≥15 years 1309 ≥20 years 1140	Yes	No	No	Yes	Poor	Low retention; thus, full extent of harms unavailable in data
Byrd, 1977 ¹⁴⁶ 120	Round 1: Yes Round 2: Partially	Round 1: Partially, although unclear if INH patient data presenting SGOT values by symptoms were limited to Round 1 INH patients Round 2: No; unclear if INH patient data presenting SGOT values by symptoms included Round 2 INH patients; data limited to those who had high SGOT levels and/or DC treatment	Round 1: Yes Round 2: Partially	Round 1: Yes Round 2: Partially	Fair	Patients only followed for 3 months of 3-month treatment; limited data presented from Round 2

Appendix E Table 7. Additional Quality Ratings for Randomized, Controlled Trials Used Only in Sensitivity Analysis for Harms

First Author, Year Trial name N	Were harms prespecified and defined?	Were ascertainment techniques for harms adequately described?	Were ascertainment techniques for harms equal, valid, and reliable?	Was duration of followup adequate for harms assessment?	Harms Quality Rating	Comments (Explain Poor Quality Ratings)
Falk, 1978 ^{41,148} 7,036	No	No	No	Yes	Poor	No consistent method for obtaining information on harms No followup labs or other formal process to adequately assess for elevated LFTs, hepatotoxicity, or other AEs They surveyed the investigators to determine any known cause of toxicity (unclear, but seems to have been a post-hoc survey; and no further information about what the survey contained or how the investigators collected information to respond to the survey)
Ferebee, 1963 ⁴⁴ 27,924 patients (566 psychiatric wards randomized); 25,210 patients included in morbidity analyses	No	No	NR	Yes	Poor	Only harm reported is number stopping pills because they were made "sick" from the pills

AE = adverse event; DC = discontinuation; INH = isoniazid; LFT = liver function test; SGOT = serum glutamic-oxalacetic transaminase.

Appendix E Table 8. Quality Ratings for Observational Studies Used Only in Sensitivity Analysis for Harms (KQ 5), Part 1

First Author, Year Study Design	Were eligibility criteria clearly de-scribed?	Were subjects representative of the overall source population?	Did the study apply inclusion/exclusion criteria uniformly to all comparison groups of the study?	Did the study avoid inappropriate exclusions?	Is the selection of the comparison group appropriate, after taking into account feasibility and ethical considerations?	Did the study guard against risk of survivor bias?	Were groups similar at baseline?	Were outcome assessors masked to the exposure status of participants?	What was the overall attrition ?	What was the differential attrition?	Did the study have high attrition raising concern for bias?
Polesky 1996 ¹⁵⁶ Retrospective cohort 87	Yes	NR (no data provided on the source population)	Yes	Yes	Yes	Yes	Yes, as reported, but baseline data not available for all subjects (e.g., limited data available on HIV status, IV drug use)	No	14%	27 patients reported lost to followup immediately after skin test conversions in the no-therapy group (38%); 0 (0) in other groups	Not overall, but high differential attrition when compared with the no-therapy group

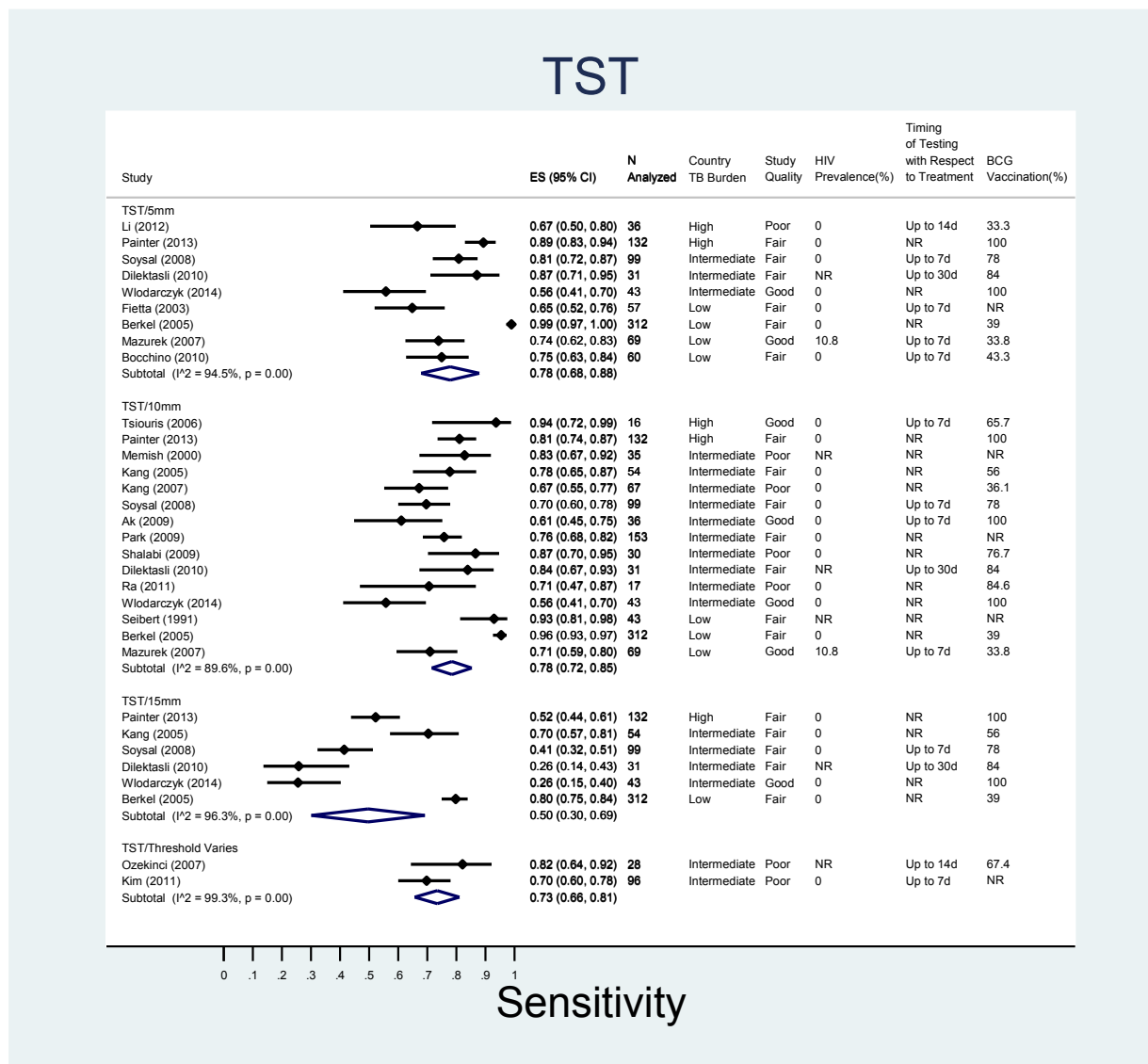
HIV = human immunodeficiency virus; IV = intravenous; NR = not reported.

Appendix E Table 9. Quality Ratings for Observational Studies Used Only in Sensitivity Analysis for Harms (KQ 5), Part 2

First Author, Year Study Name	Were harms pre-specified and defined?	Were ascertainment techniques for harms adequately described?	Were ascertainment techniques (outcome measures) for harms equal, valid, and reliable?	Was the duration of followup adequate to assess the outcome?	Does the analysis control for baseline differences between groups?	Does the analysis control for potential confounders?	Was an appropriate method used to handle missing data?	Did the study use appropriate statistical methods?	Quality Rating	Comments (explain Poor Quality Ratings)
Polesky, 1996 ¹⁵⁶ Retrospective cohort 87	No	No	NR, Unclear	Yes	No; and they had limited information available to determine similarity of groups at baseline	No	NR; for harms information, it is unclear how much missing data there were	Yes	Poor	Retrospective study designed aiming to assess benefits; methods for ascertaining harms not adequately described; high risk of selection bias and confounding Frequency of harms in no-treatment group was not reported for comparison; some differences in followup for those in the TB clinic

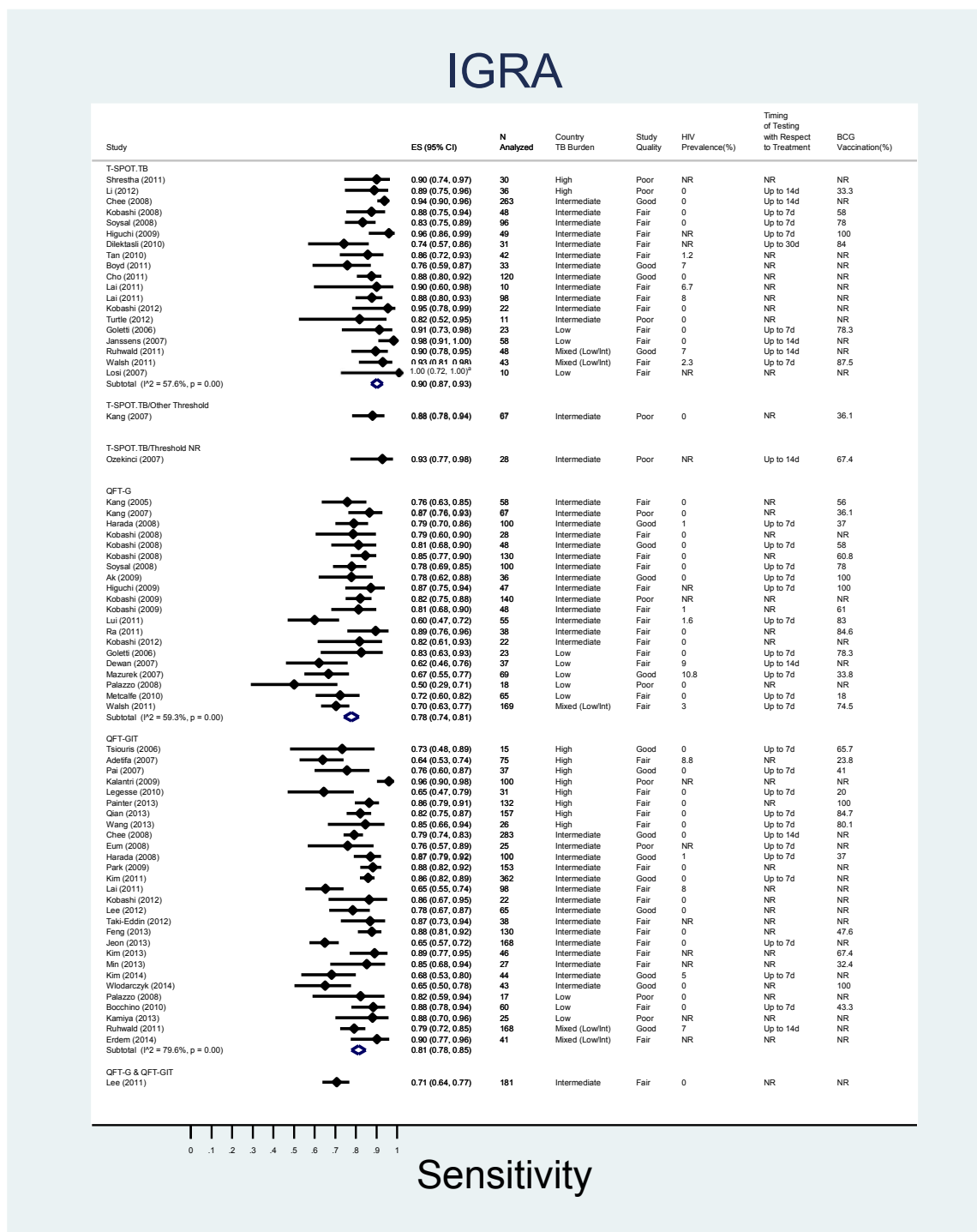
NR = not reported; TB = tuberculosis.

Appendix F Figure 1. Individual Study and Pooled Estimates (Where Applicable) of Sensitivity for Various Thresholds of TST for Tuberculosis Infection, Including Poor-Quality Studies



BCG = bacillus Calmette-Guerin; CI = confidence interval; d = day; ES = effect size; HIV = Human immunodeficiency virus; mm = millimeter; N = number; NR = not reported; TB = tuberculosis; TST = tuberculin skin test.

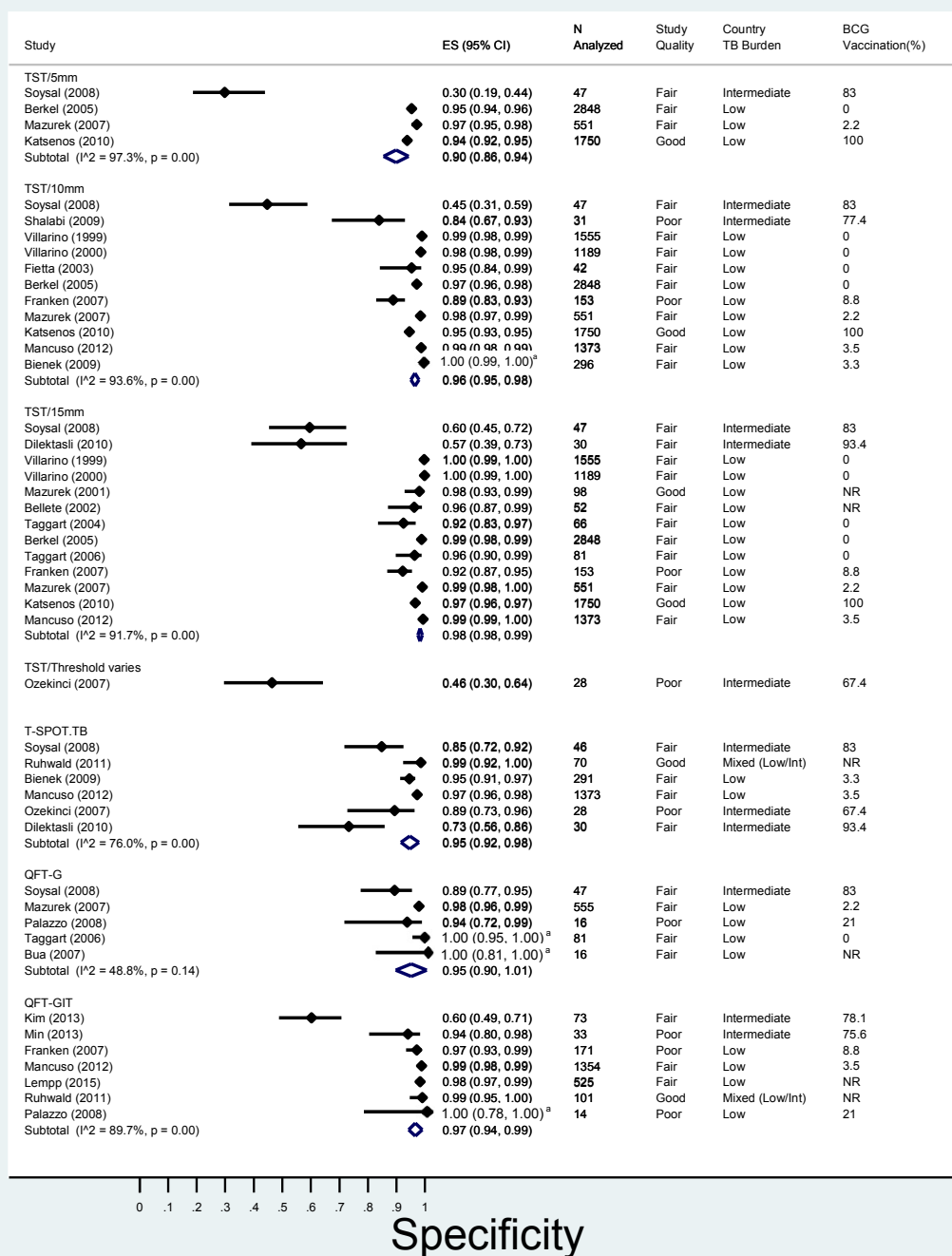
Appendix F Figure 2. Individual Study and Pooled Estimates (Where Applicable) of Sensitivity for Various Thresholds of IGRA Tests for Tuberculosis Infection, Including Poor-Quality Studies



^a Excluded from pooled estimate due to point estimate of 1.0.

BCG = bacillus Calmette-Guerin; CI = confidence interval; d = day; ES = effect size; HIV = Human immunodeficiency virus; mm = millimeter; N = number; NR = not reported; TB = tuberculosis; TST = tuberculin skin test.

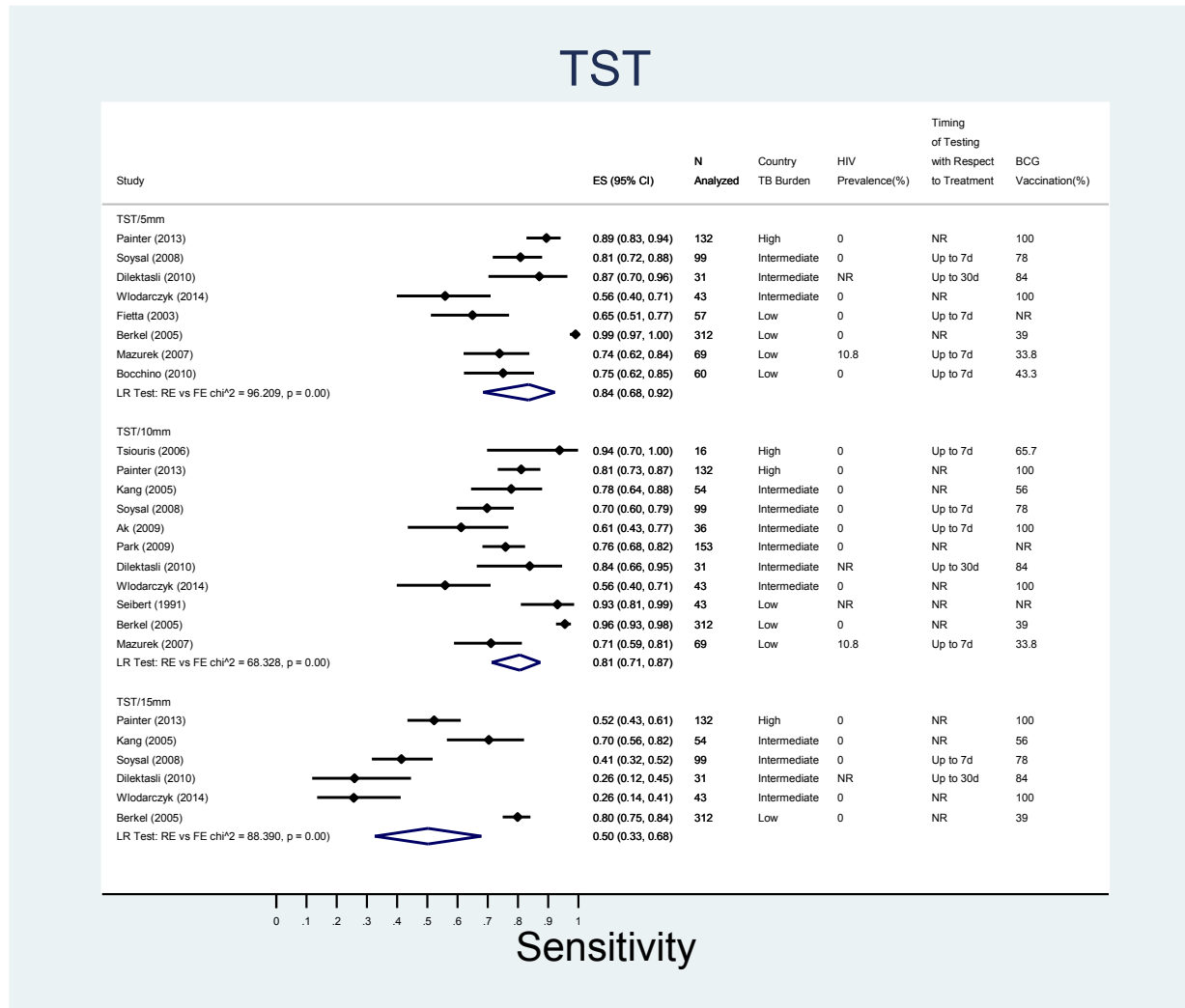
Appendix F Figure 3. Individual Study and Pooled Estimates (Where Applicable) of Specificity for Various Thresholds of TST and IGRA Tests for Tuberculosis Infection, Including Poor-Quality Studies



^a Excluded from pooled estimate due to point estimate of 1.0.

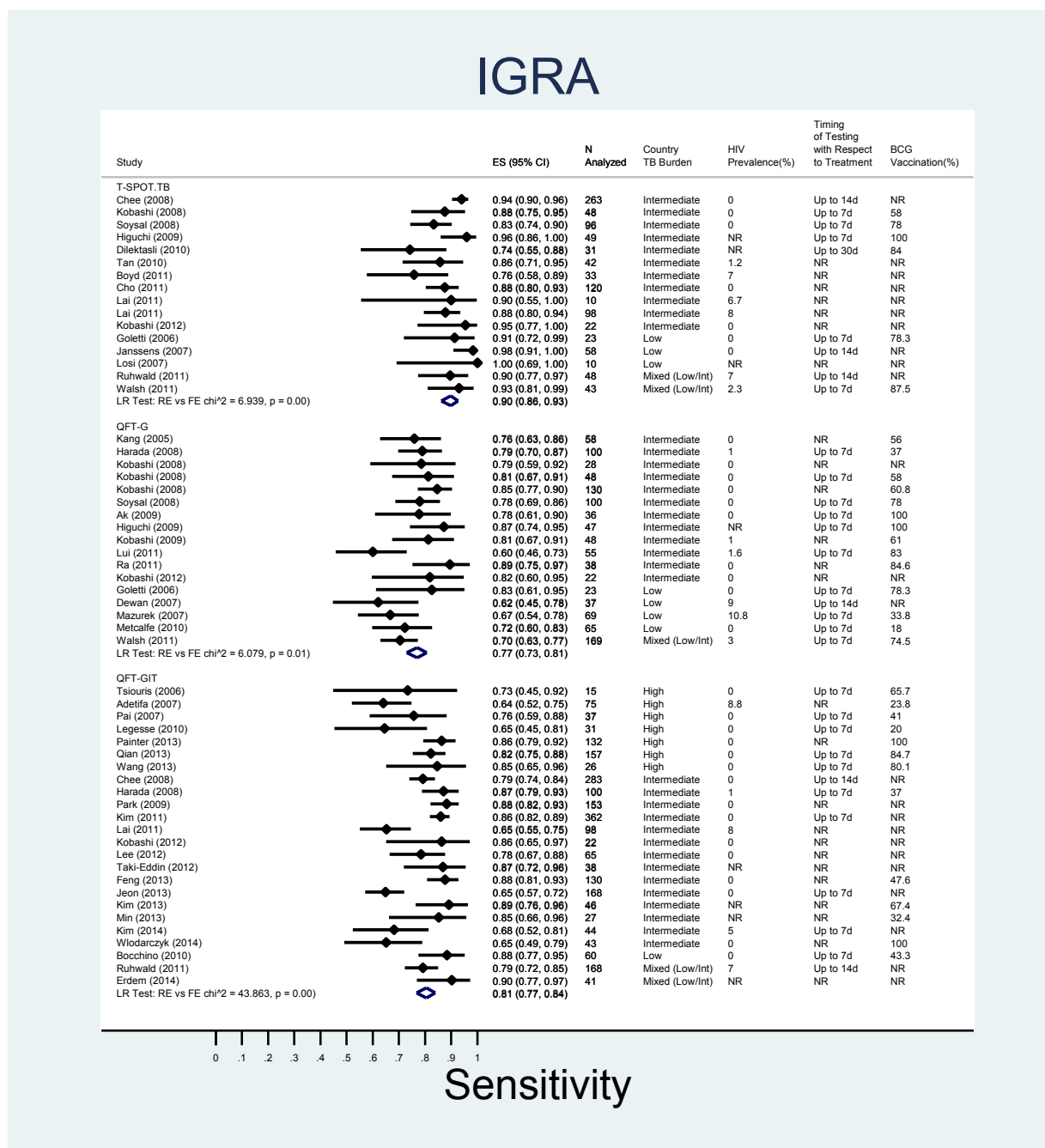
BCG = bacillus Calmette-Guerin; CI = confidence interval; d = day; ES = effect size; HIV = human immunodeficiency virus; mm = millimeter; N = number; NR = not reported; TB = tuberculosis; TST = tuberculin skin test.

Appendix F Figure 4. Individual Study and Pooled Estimates (Where Applicable) of Sensitivity for Various Thresholds of TST for Tuberculosis Infection Using Maximum Likelihood Random-Effects Model



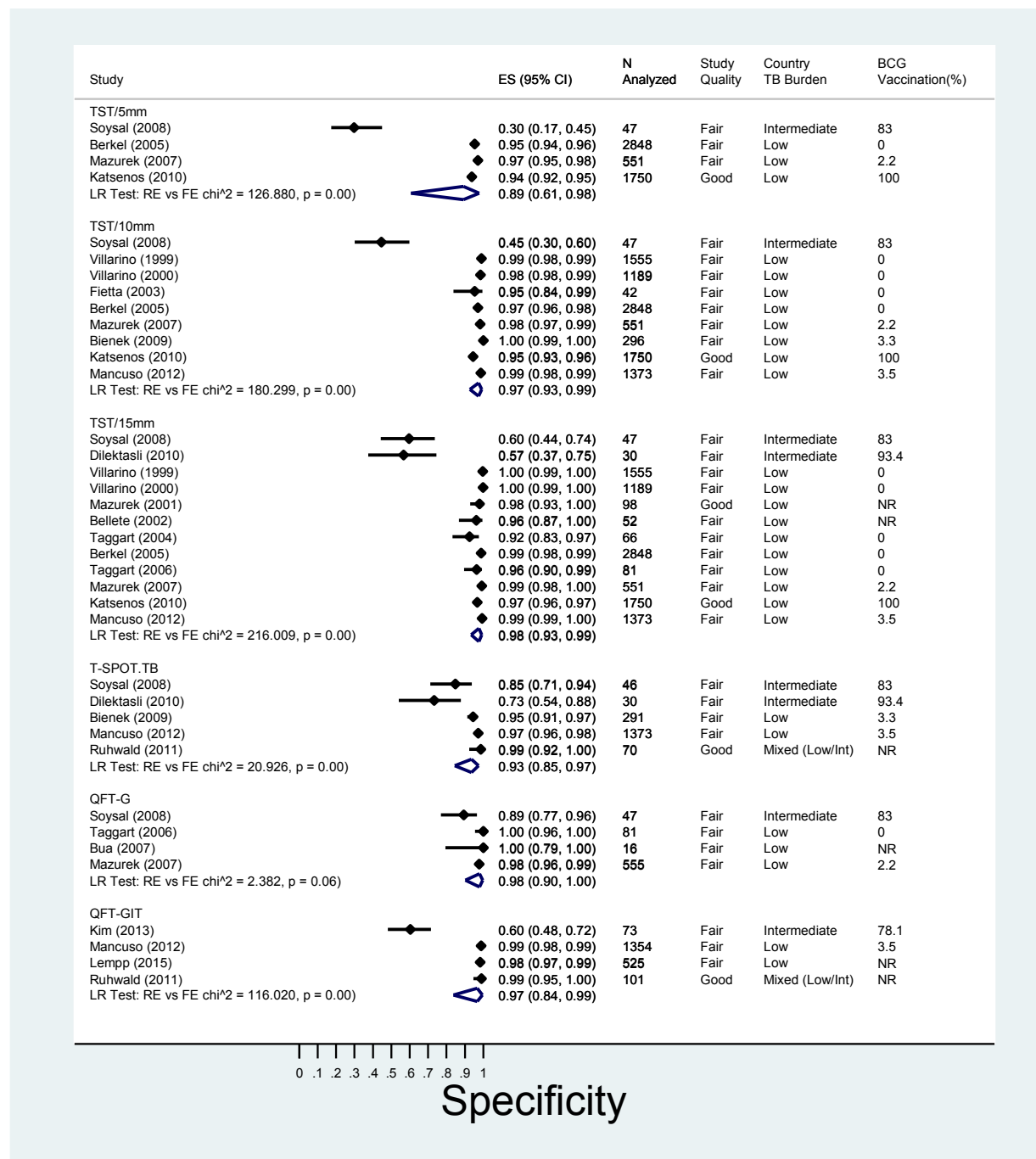
BCG = bacillus Calmette-Guerin; CI = confidence interval; d = day; ES = effect size; HIV = human immunodeficiency virus; mm = millimeter; N = number; NR = not reported; TB = tuberculosis; TST = tuberculin skin test.

Appendix F Figure 5. Individual Study and Pooled Estimates (Where Applicable) of Sensitivity for Various Thresholds of IGRA Tests for Tuberculosis Infection Using Maximum Likelihood Random-Effects Model



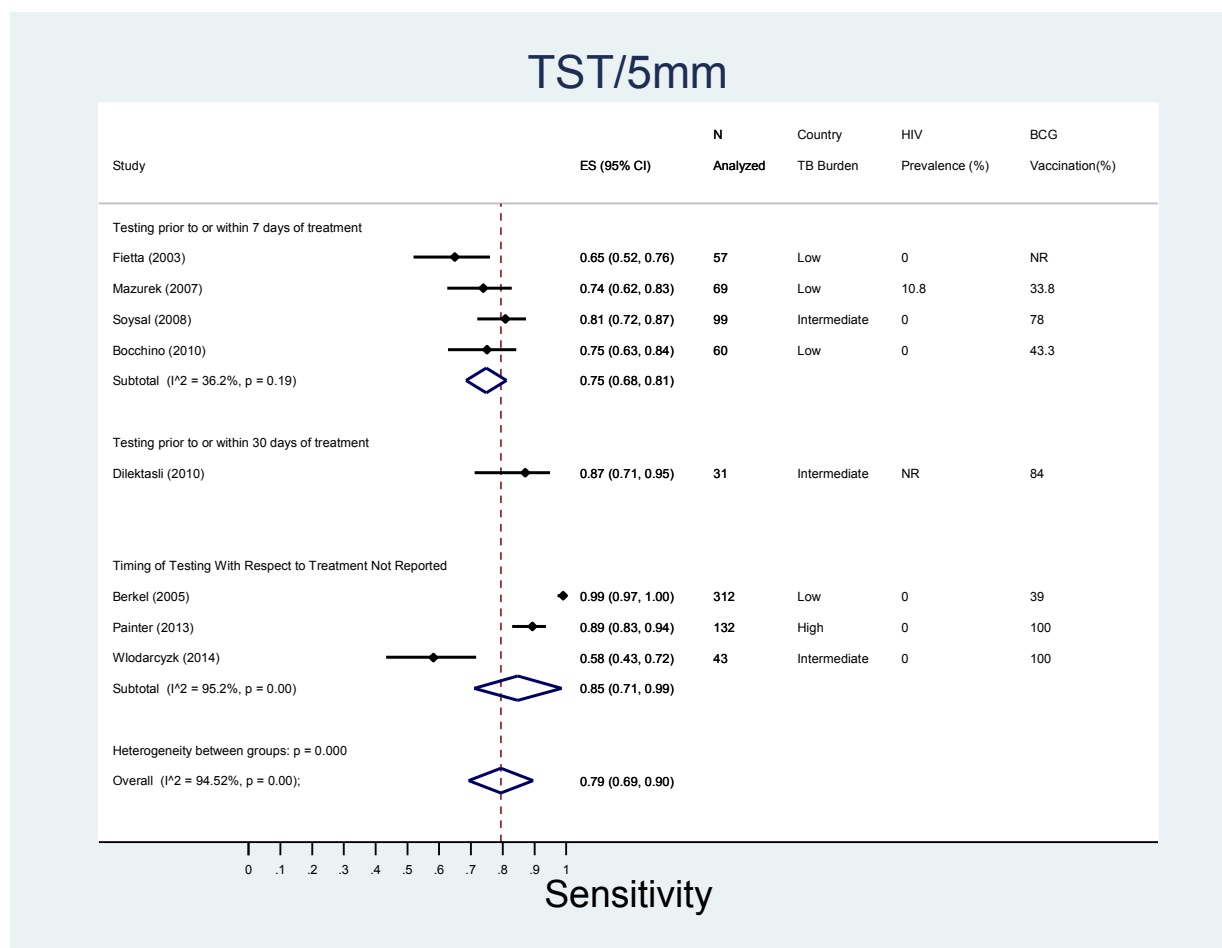
BCG = bacillus Calmette-Guerin; CI = confidence interval; d = day; ES = effect size; HIV = human immunodeficiency virus; IGRA = interferon gamma release assay; N = number; NR = not reported; TB = tuberculosis; TST = tuberculin skin test.

Appendix F Figure 6. Individual Study and Pooled Estimates (Where Applicable) of Specificity for Various Thresholds of TST and IGRA Tests for Tuberculosis Infection Using Maximum Likelihood Random-Effects Model



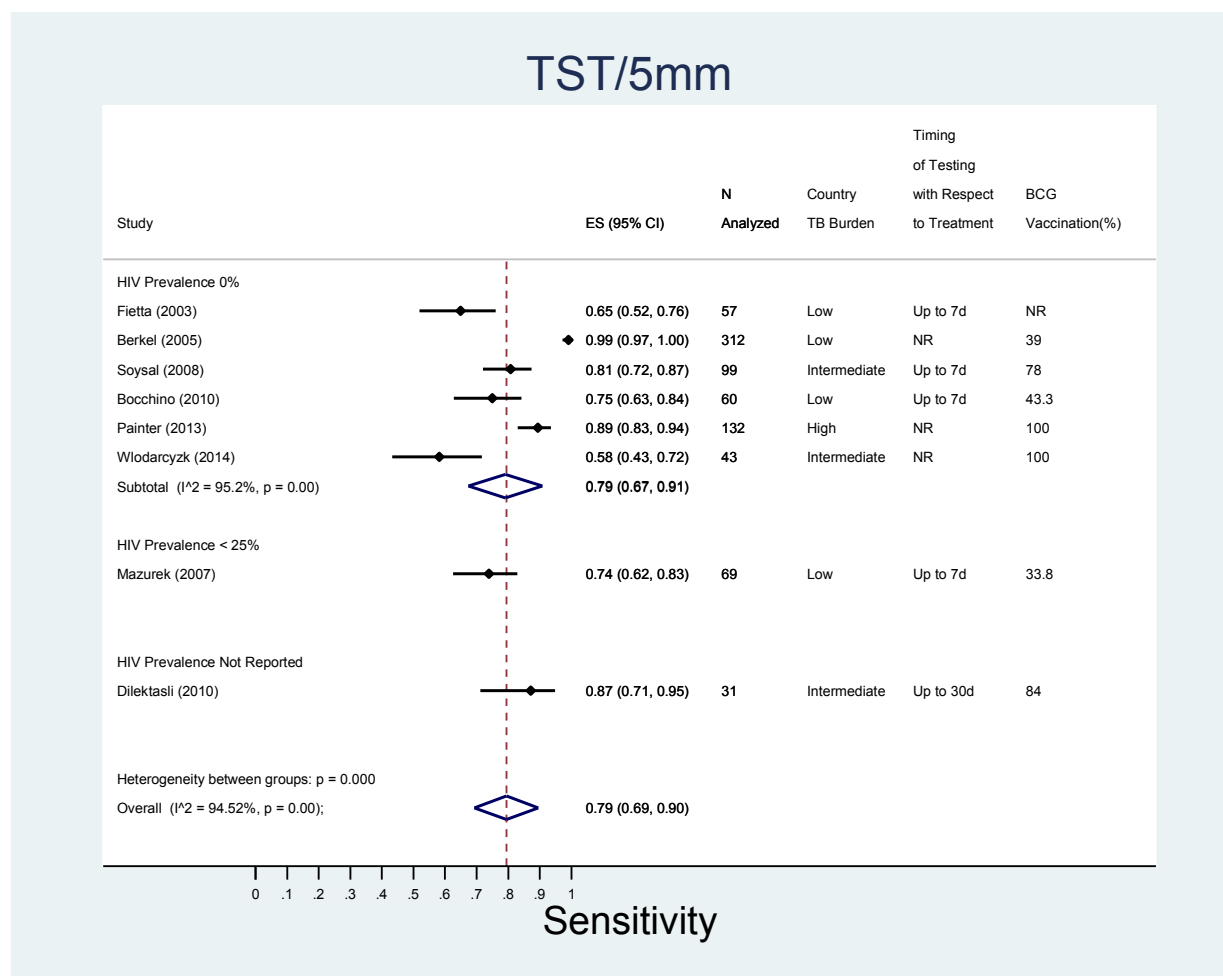
BCG = bacillus Calmette-Guerin; CI = confidence interval; d = day; ES = effect size; HIV = human immunodeficiency virus; mm = millimeter; N = number; NR = not reported; TB = tuberculosis; TST = tuberculin skin test.

Appendix F Figure 7. Individual Study and Pooled Estimates (Where Applicable) of Sensitivity for TST at 5-mm Threshold Stratified by Timing of Testing With Respect to Antituberculosis Treatment



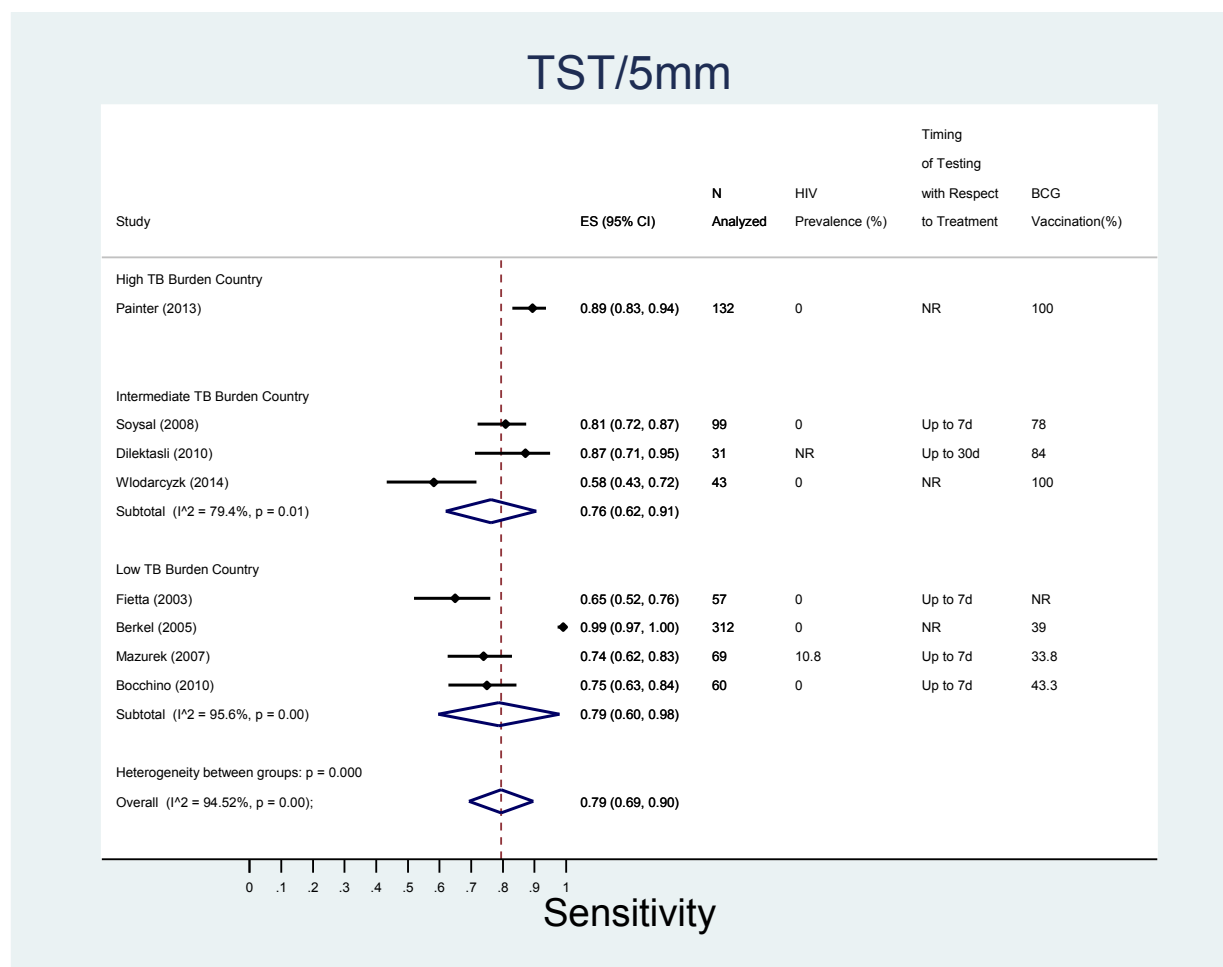
BCG = bacillus Calmette-Guerin; CI = confidence interval; ES = effect size; HIV = human immunodeficiency virus; mm = millimeter; NR = not reported; TB = tuberculosis.

Appendix F Figure 8. Individual Study and Pooled Estimates (Where Applicable) of Sensitivity for TST at 5-mm Threshold Stratified by HIV Prevalence of the Study Population



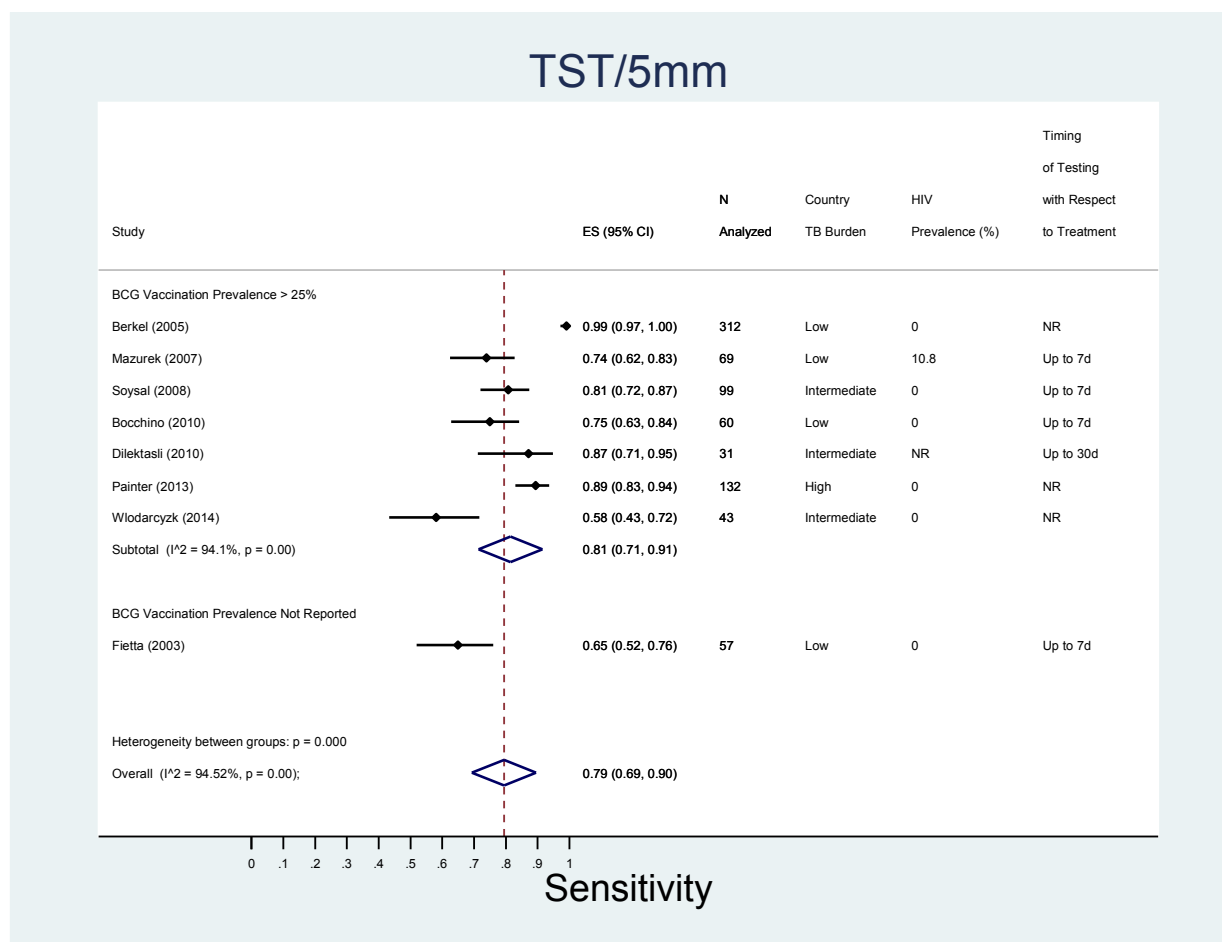
BCG = bacillus Calmette-Guerin; CI = confidence interval; d = day; ES = effect size; HIV = human immunodeficiency virus; mm = millimeter; N = number; NR = not reported; TB = tuberculosis; TST = tuberculin skin test.

Appendix F Figure 9. Individual Study and Pooled Estimates (Where Applicable) of Sensitivity for TST at 5-mm Threshold Stratified by Country TB Burden of the Study Setting



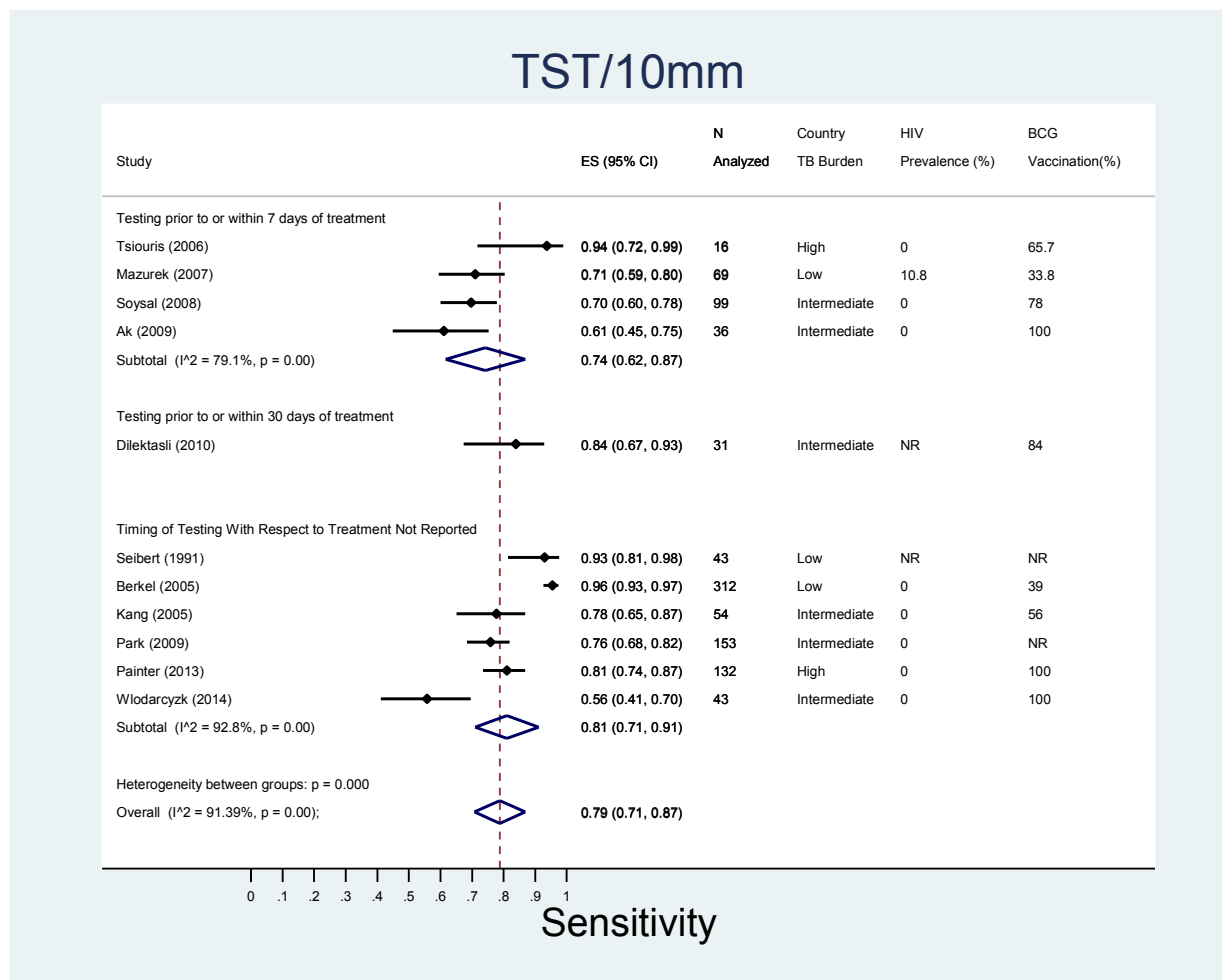
BCG = bacillus Calmette-Guerin; CI = confidence interval; d = day; ES = effect size; HIV = human immunodeficiency virus; mm = millimeter; N = number; NR = not reported; TB = tuberculosis; TST = tuberculin skin test.

Appendix F Figure 10. Individual Study and Pooled Estimates (Where Applicable) of Sensitivity for TST at 5-mm Threshold Stratified by Country BCG Vaccination Prevalence of the Study Population



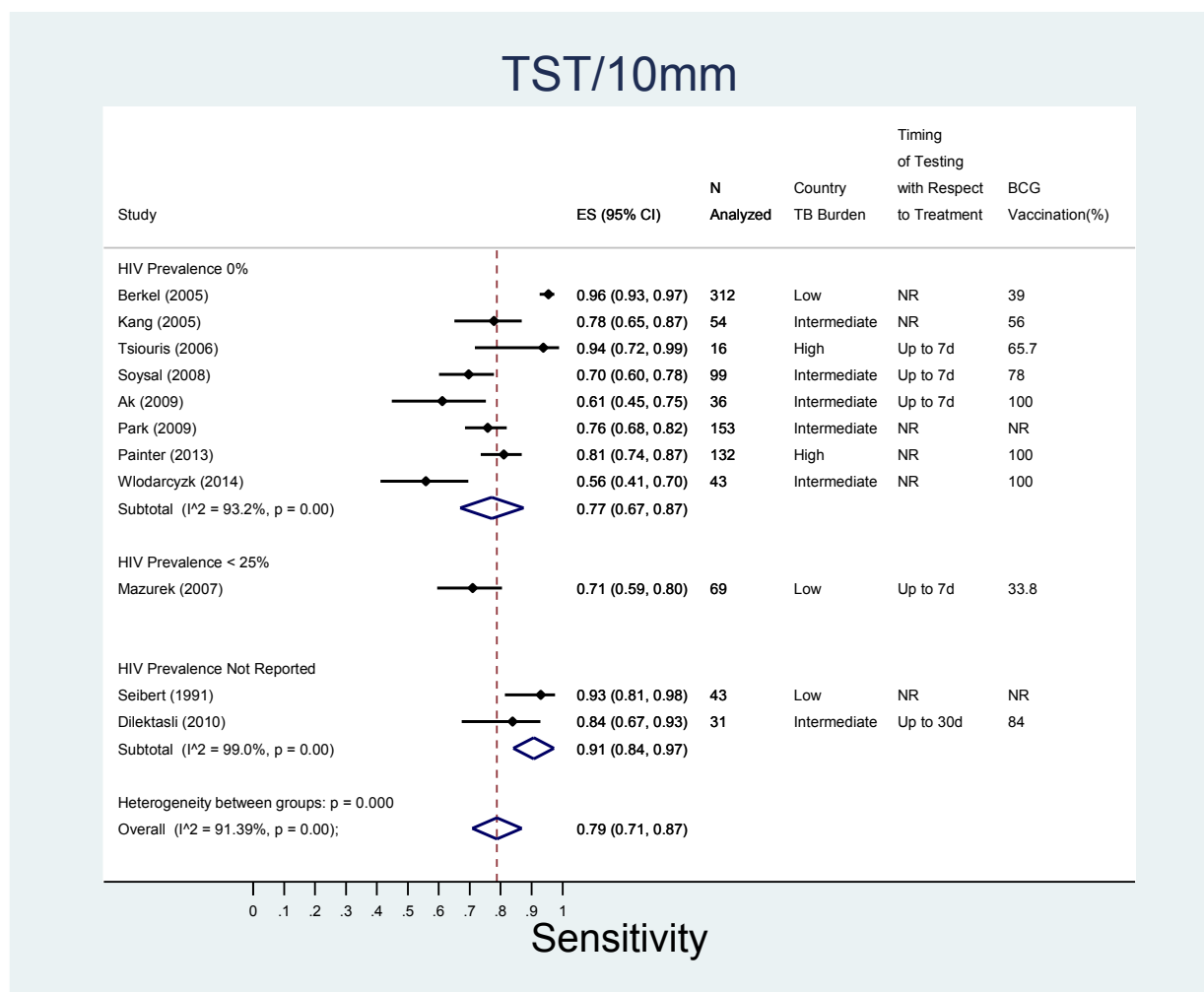
BCG = bacillus Calmette-Guerin; CI = confidence interval; d = day; ES = effect size; HIV = human immunodeficiency virus; mm = millimeter; N = number; NR = not reported; TB = tuberculosis; TST = tuberculin skin test.

Appendix F Figure 11. Individual Study and Pooled Estimates (Where Applicable) of Sensitivity for TST at 10-mm Threshold Stratified by Timing of Testing With Respect to Antituberculosis Treatment



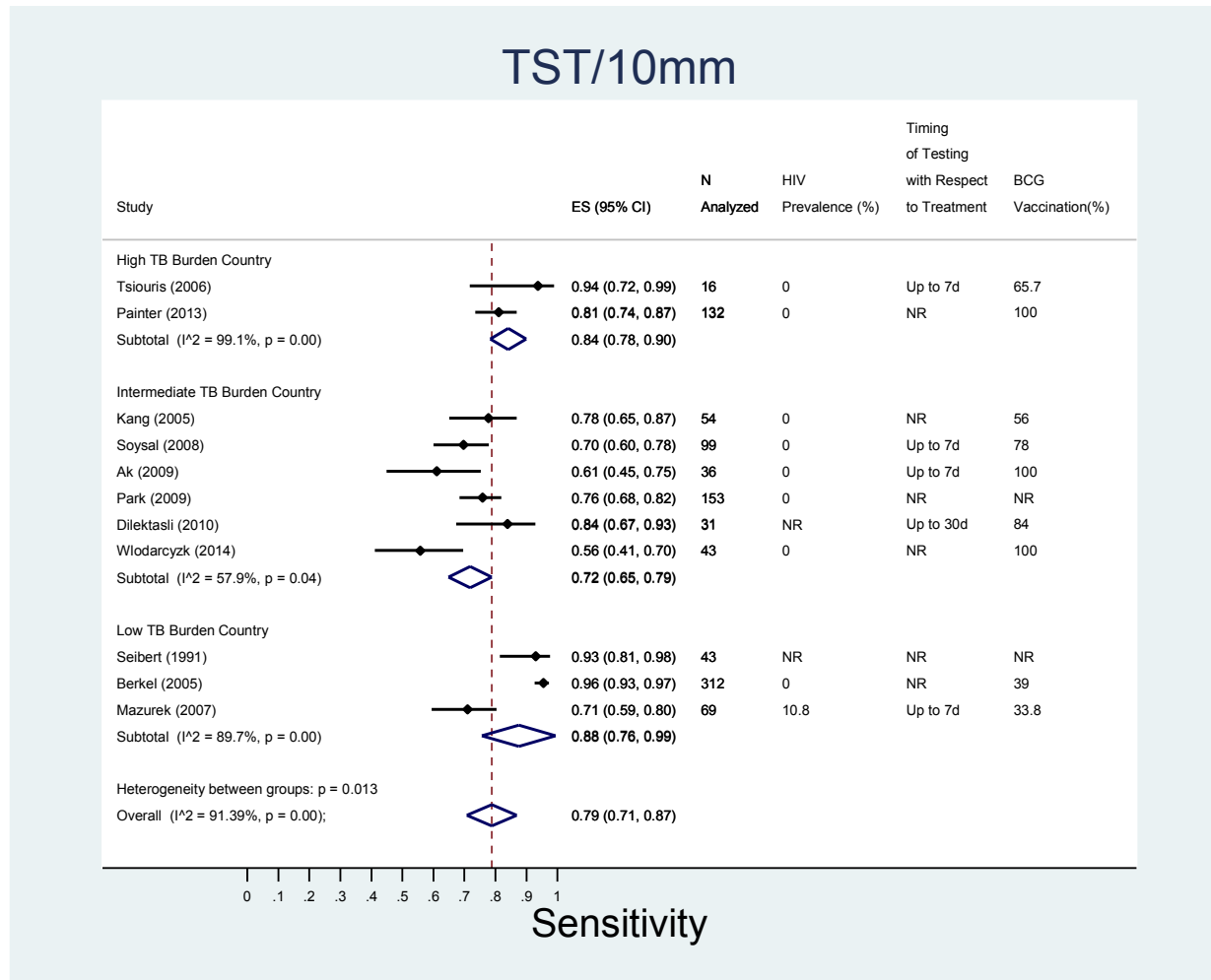
BCG = bacillus Calmette-Guerin; CI = confidence interval; ES = effect size; HIV = human immunodeficiency virus; mm = millimeter; N = number; NR = not reported; TB = tuberculosis; TST = tuberculin skin test.

Appendix F Figure 12. Individual Study and Pooled Estimates (Where Applicable) of Sensitivity for TST at 10-mm Threshold Stratified by HIV Prevalence of the Study Population



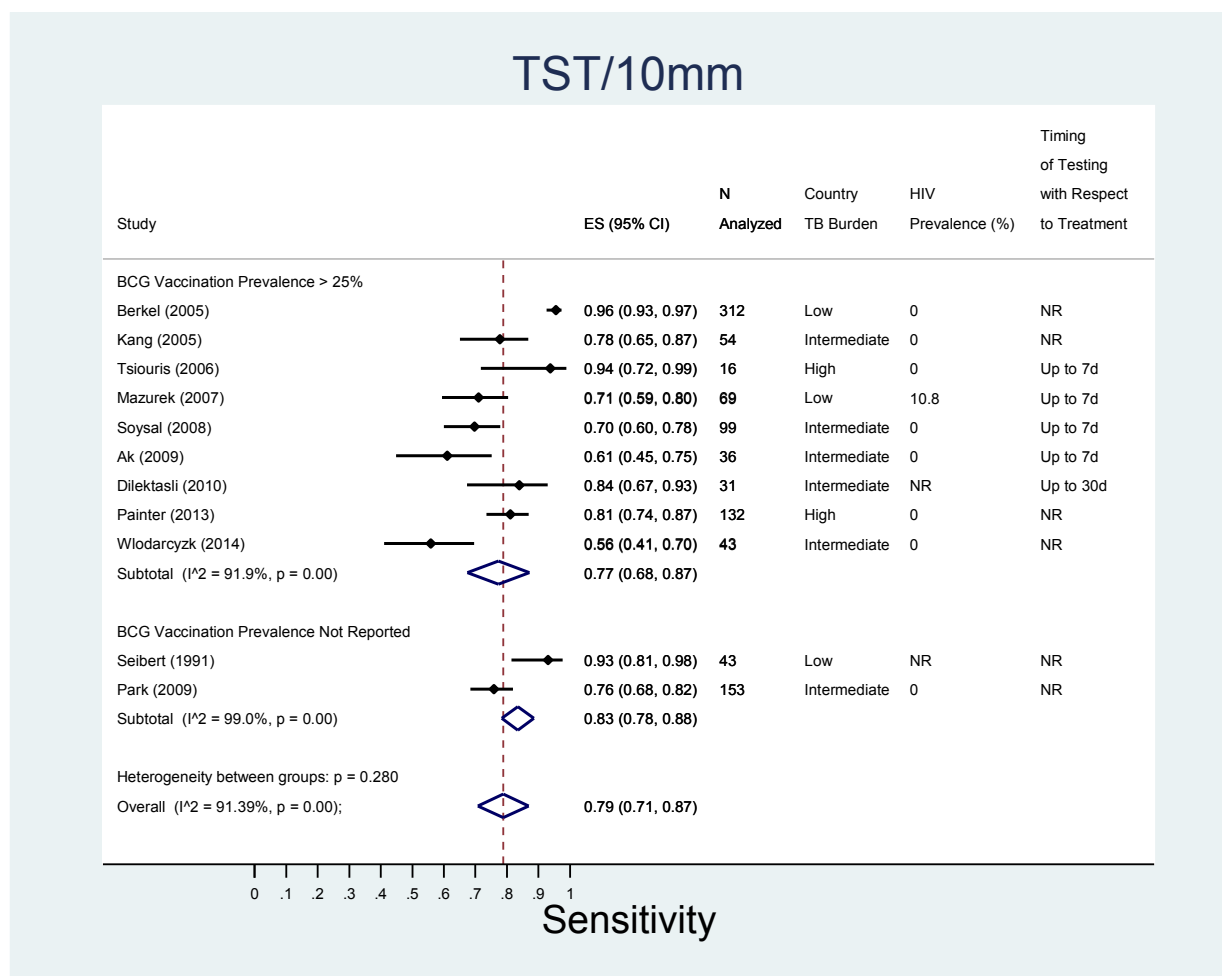
BCG = bacillus Calmette-Guerin; CI = confidence interval; d = day; ES = effect size; HIV = human immunodeficiency virus; mm = millimeter; N = number; NR = not reported; TB = tuberculosis; TST = tuberculin skin test.

Appendix F Figure 13. Individual Study and Pooled Estimates (Where Applicable) of Sensitivity for TST at 10-mm Threshold Stratified by Country TB Burden of the Study Setting



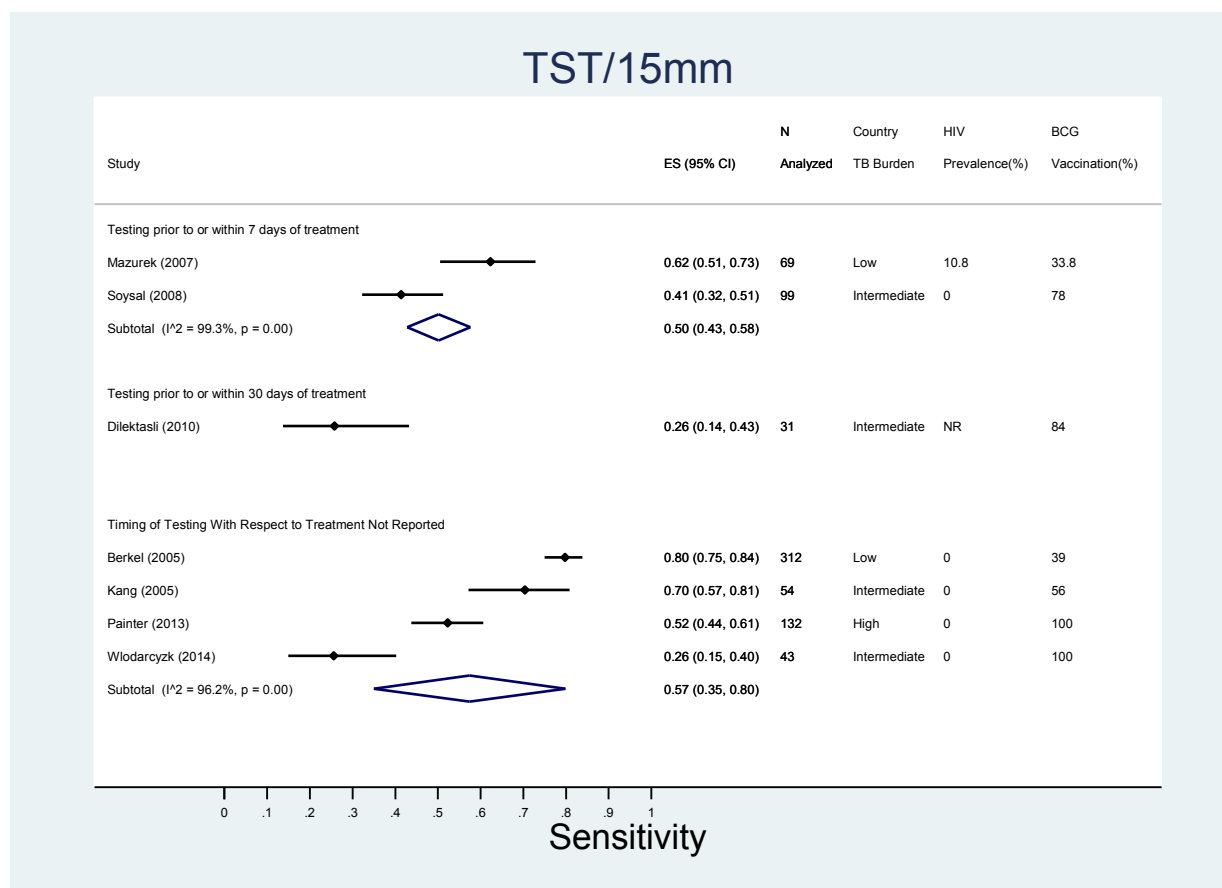
BCG = bacillus Calmette-Guerin; CI = confidence interval; d = day; ES = effect size; HIV = human immunodeficiency virus; mm = millimeter; N = number; NR = not reported; TB = tuberculosis; TST = tuberculin skin test.

Appendix F Figure 14. Individual Study and Pooled Estimates (Where Applicable) of Sensitivity for TST at 10-mm Threshold Stratified by Country BCG Vaccination Prevalence of the Study Setting



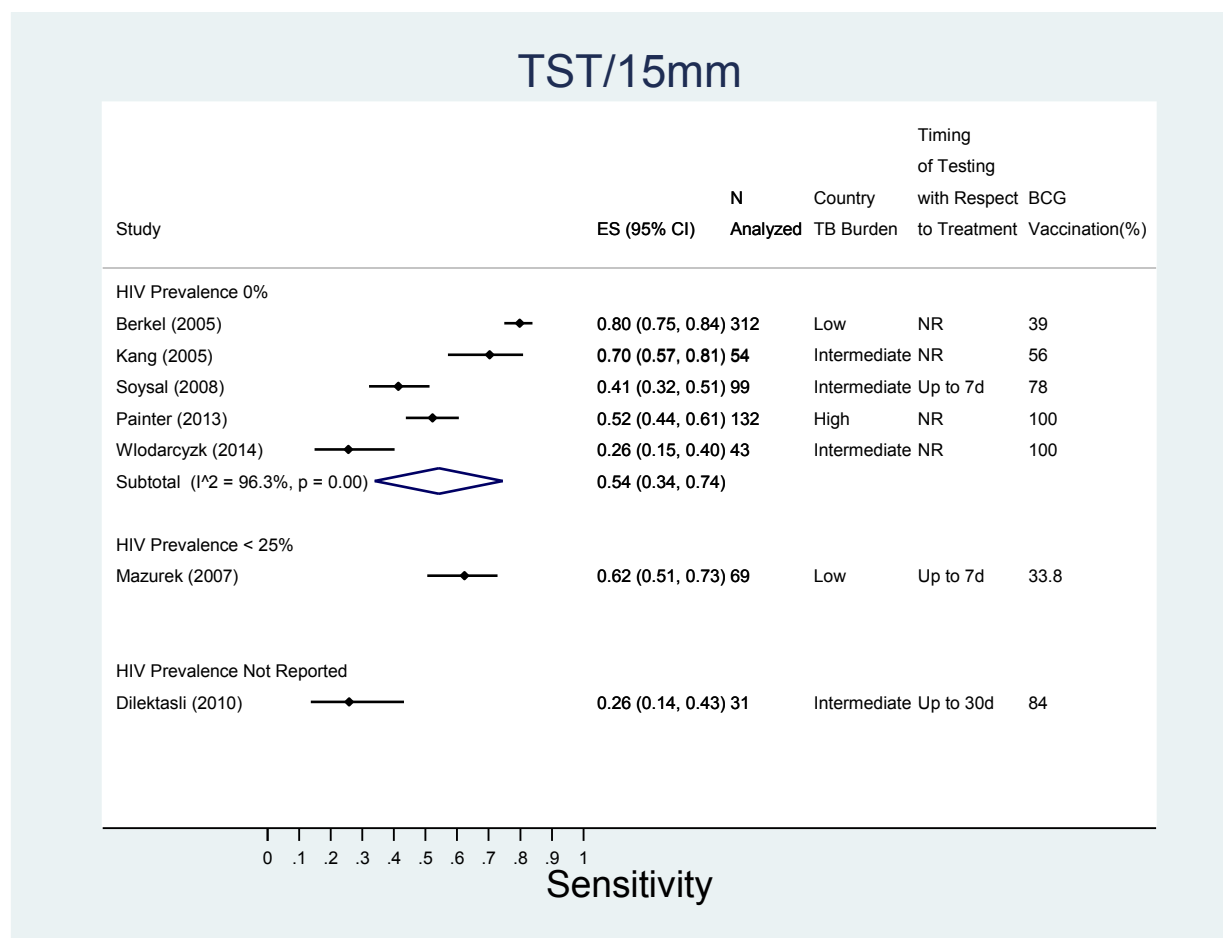
BCG = bacillus Calmette-Guerin; CI = confidence interval; d = day; ES = effect size; HIV = human immunodeficiency virus; mm = millimeter; N = number; NR = not reported; TB = tuberculosis; TST = tuberculin skin test.

Appendix F Figure 15. Individual Study and Pooled Estimates (Where Applicable) of Sensitivity for TST at 15-mm Threshold Stratified by Timing of Testing With Respect to Antituberculosis Treatment



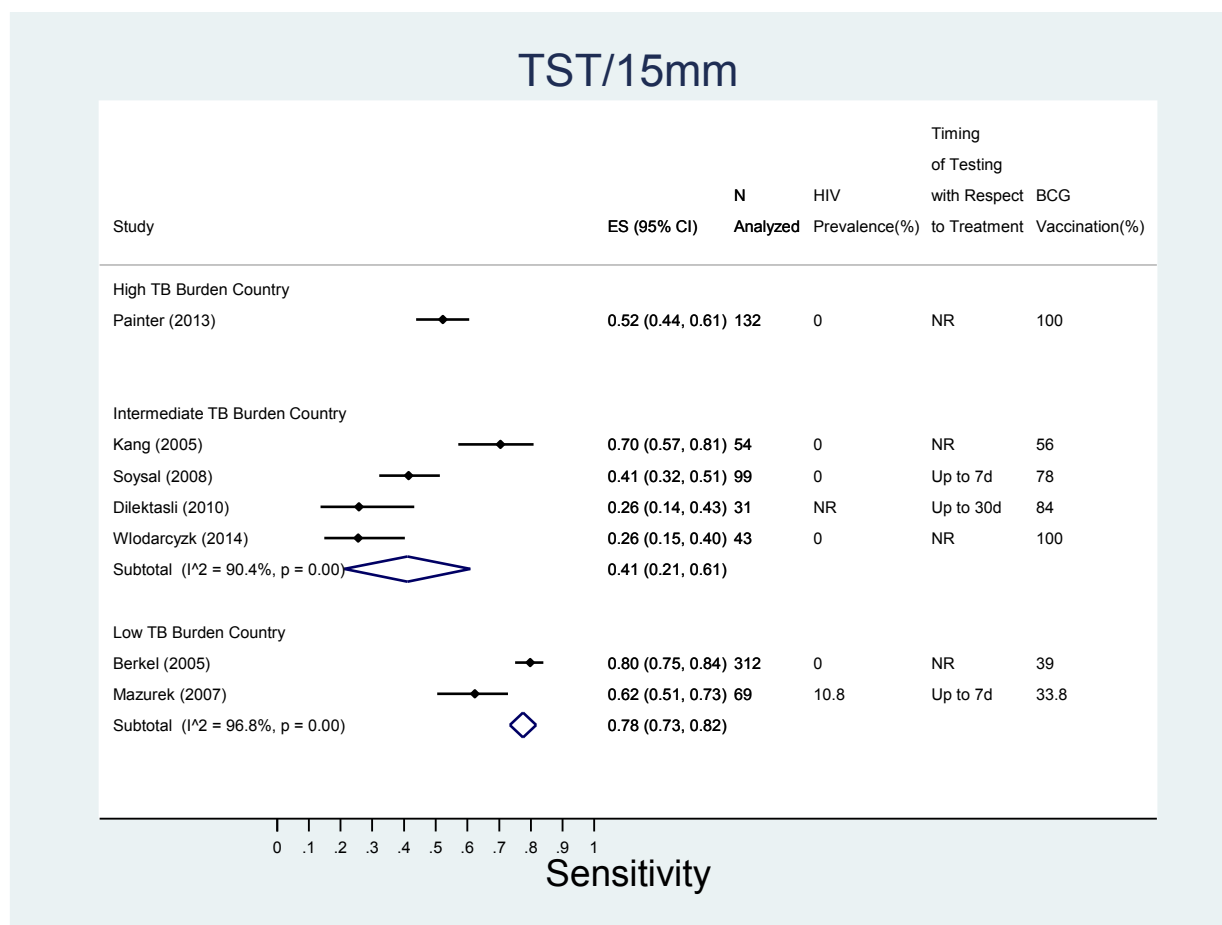
BCG = bacillus Calmette-Guerin; CI = confidence interval; ES = effect size; HIV = human immunodeficiency virus; mm = millimeter; N = number; NR = not reported; TB = tuberculosis; TST = tuberculin skin test.

Appendix F Figure 16. Individual Study and Pooled Estimates (Where Applicable) of Sensitivity for TST at 15-mm Threshold Stratified by HIV Prevalence of the Study Population



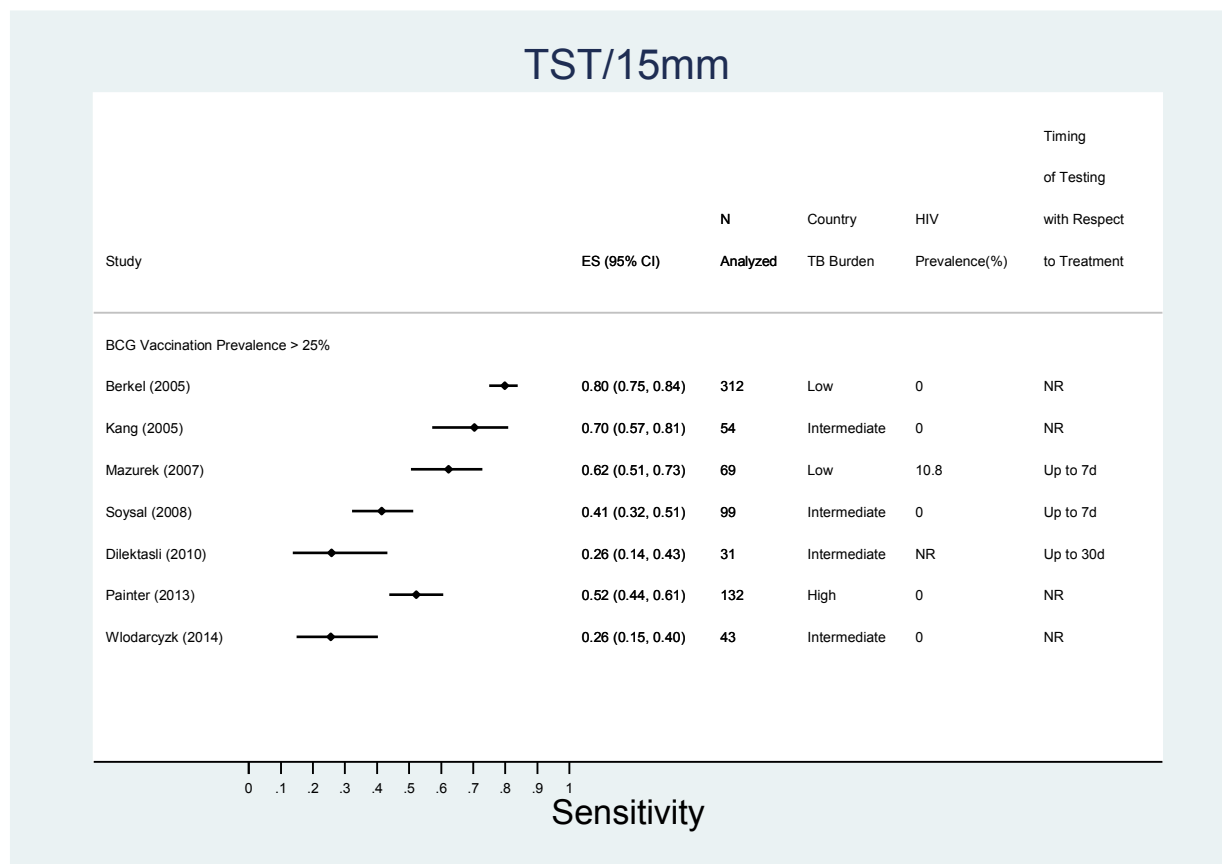
BCG = bacillus Calmette-Guerin; CI = confidence interval; d = day; ES = effect size; HIV = human immunodeficiency virus; mm = millimeter; N = number; NR = not reported; TB = tuberculosis; TST = tuberculin skin test.

Appendix F Figure 17. Individual Study and Pooled Estimates (Where Applicable) of Sensitivity for TST at 15-mm Threshold Stratified by Country TB Burden of the Study Setting



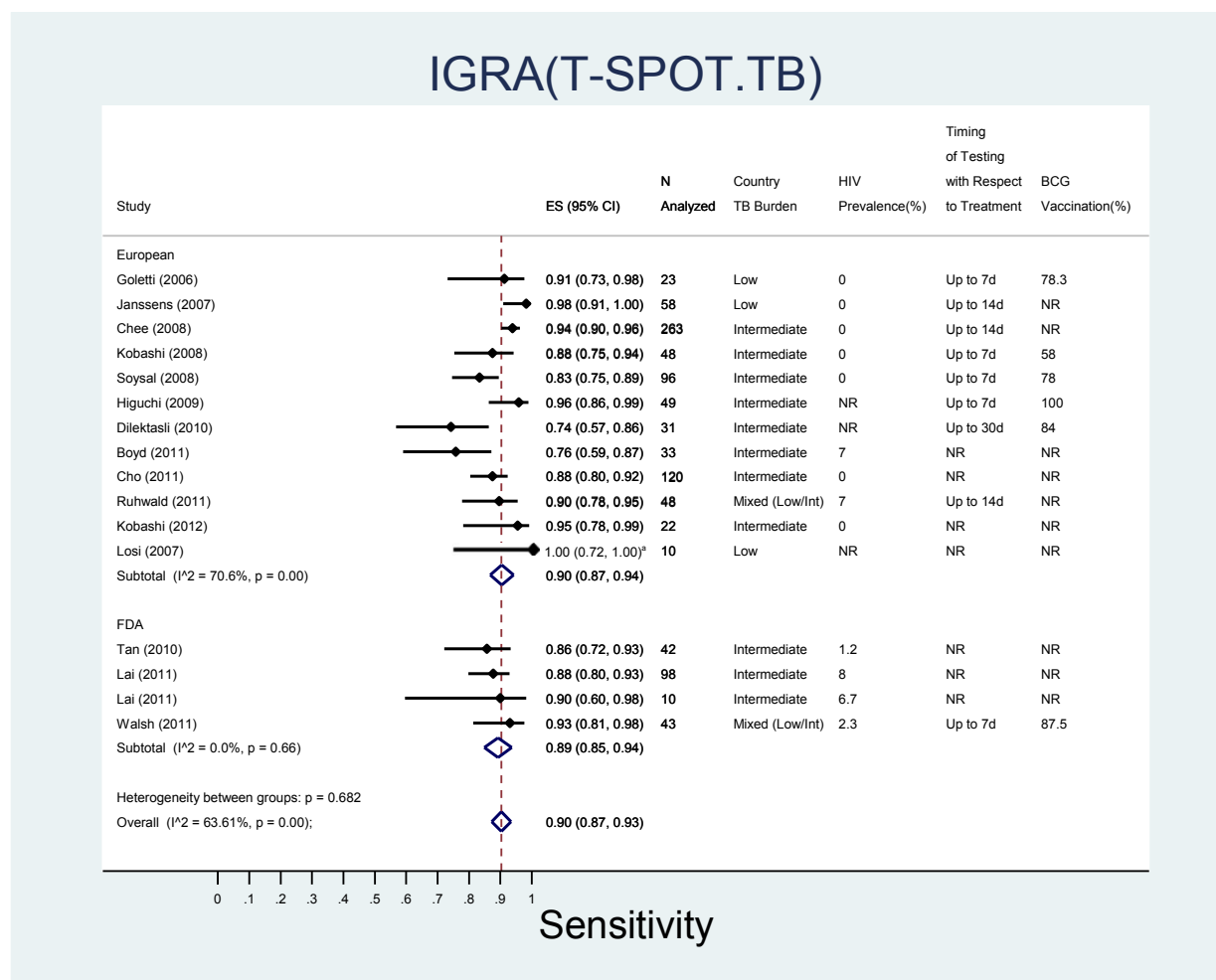
BCG = bacillus Calmette-Guerin; CI = confidence interval; d = day; ES = effect size; HIV = human immunodeficiency virus; mm = millimeter; N = number; NR = not reported; TB = tuberculosis; TST = tuberculin skin test.

Appendix F Figure 18. Individual Study and Pooled Estimates (Where Applicable) of Sensitivity for TST at 15-mm Threshold Stratified by Country BCG Vaccination Prevalence of the Study Population



BCG = bacillus Calmette-Guerin; CI = confidence interval; d = day; ES = effect size; HIV = human immunodeficiency virus; mm = millimeter; N = number; NR = not reported; TB = tuberculosis; TST = tuberculin skin test.

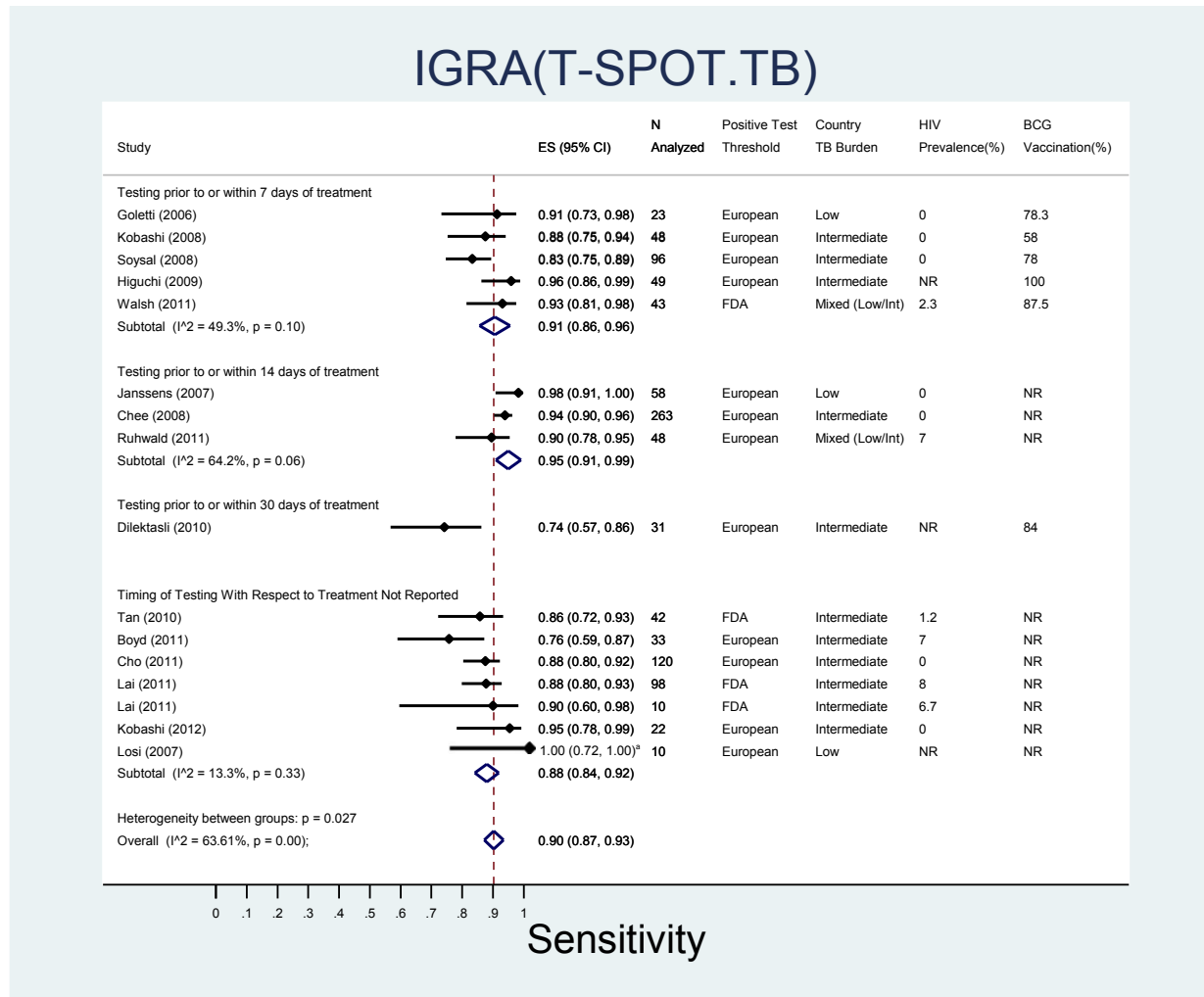
Appendix F Figure 19. Individual Study and Pooled Estimates (Where Applicable) of Sensitivity of T-SPOT.TB Test Stratified by Threshold Used for Positive Test



^a Excluded from pooled estimate due to point estimate of 1.0.

BCG = bacillus Calmette-Guerin; CI = confidence interval; d = day; ES = effect size; HIV = human immunodeficiency virus; IGRA = interferon gamma release assay; mm = millimeter; N = number; NR = not reported; TB = tuberculosis; TST = tuberculin skin test.

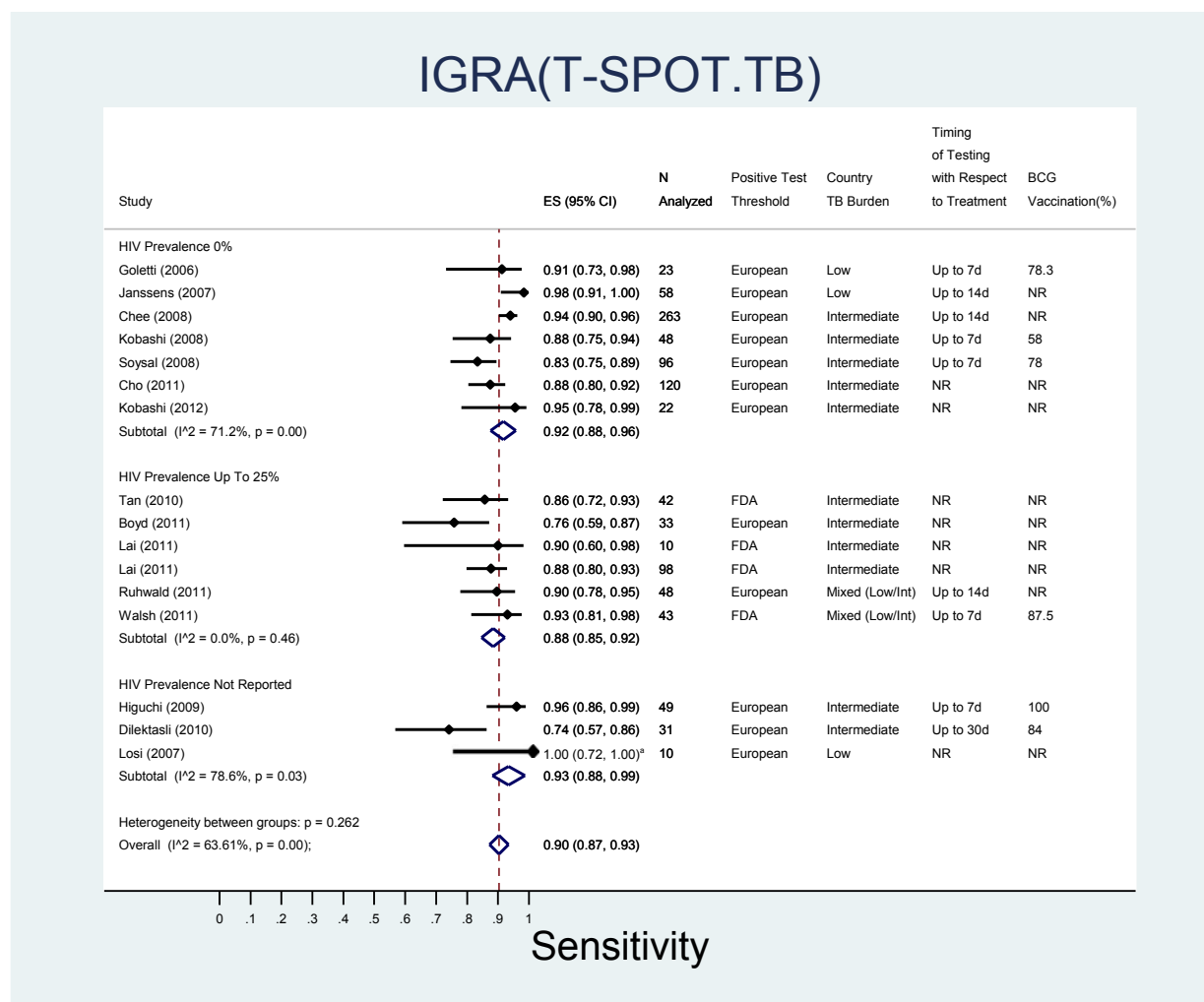
Appendix F Figure 20. Individual Study and Pooled Estimates (Where Applicable) of Sensitivity of T-SPOT.TB Test Stratified by Timing of Testing With Respect to Antituberculosis Treatment



^a Excluded from pooled estimate due to point estimate of 1.0

BCG = bacillus Calmette-Guerin; CI = confidence interval; ES = effect size; FDA = Food and Drug Administration; HIV = human immunodeficiency virus; IGRA = interferon gamma release assay; N = number; NR = not reported; TB = tuberculosis.

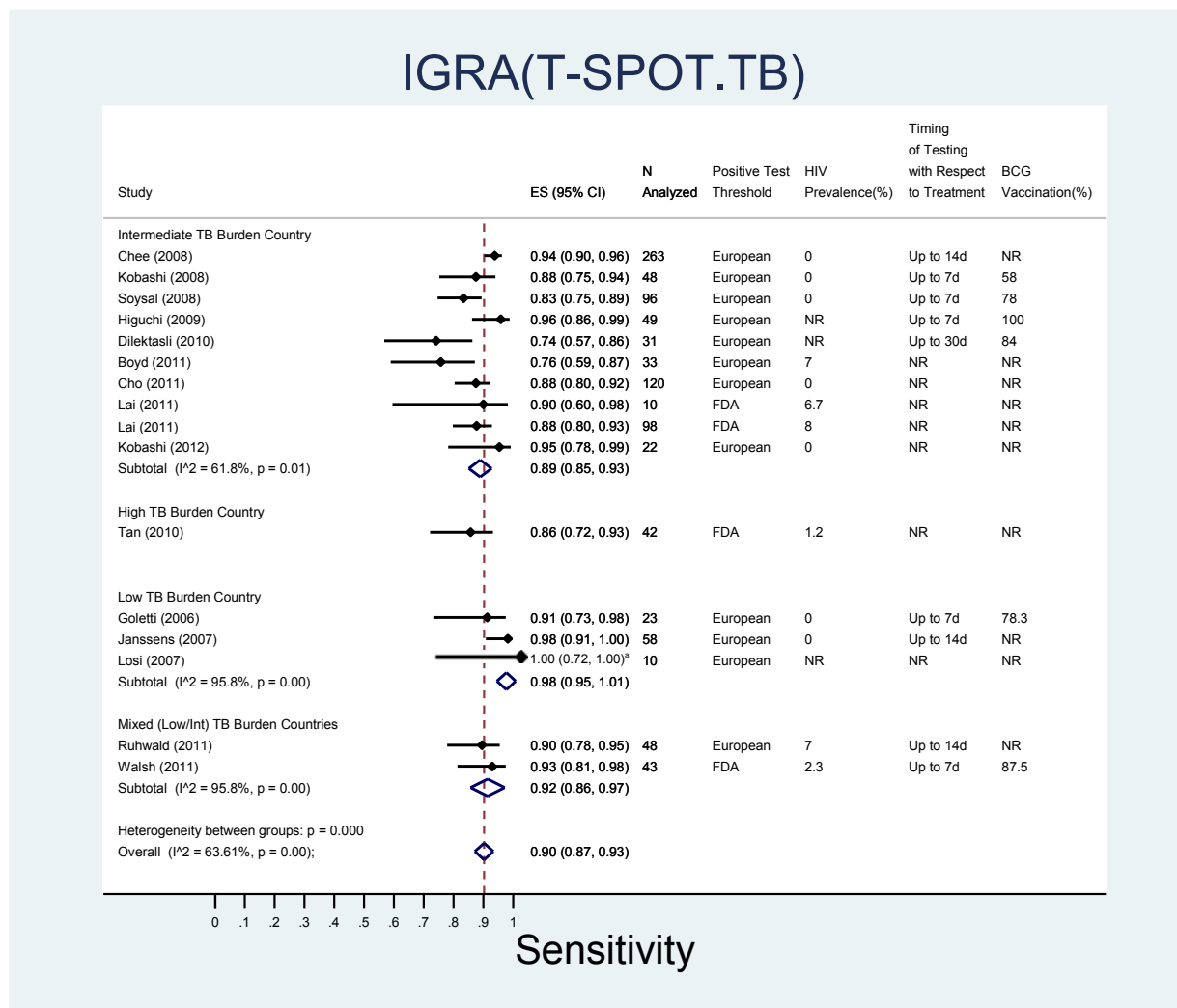
Appendix F Figure 21. Individual Study and Pooled Estimates (Where Applicable) of Sensitivity of T-SPOT.TB Test Stratified by HIV Prevalence of the Study Population



^a Excluded from pooled estimate due to point estimate of 1.0.

BCG = bacillus Calmette-Guerin; CI = confidence interval; d = day; ES = effect size; FDA = Food and Drug Administration; HIV = human immunodeficiency virus; IGRA = interferon gamma release assay N = number; NR = not reported; TB = tuberculosis.

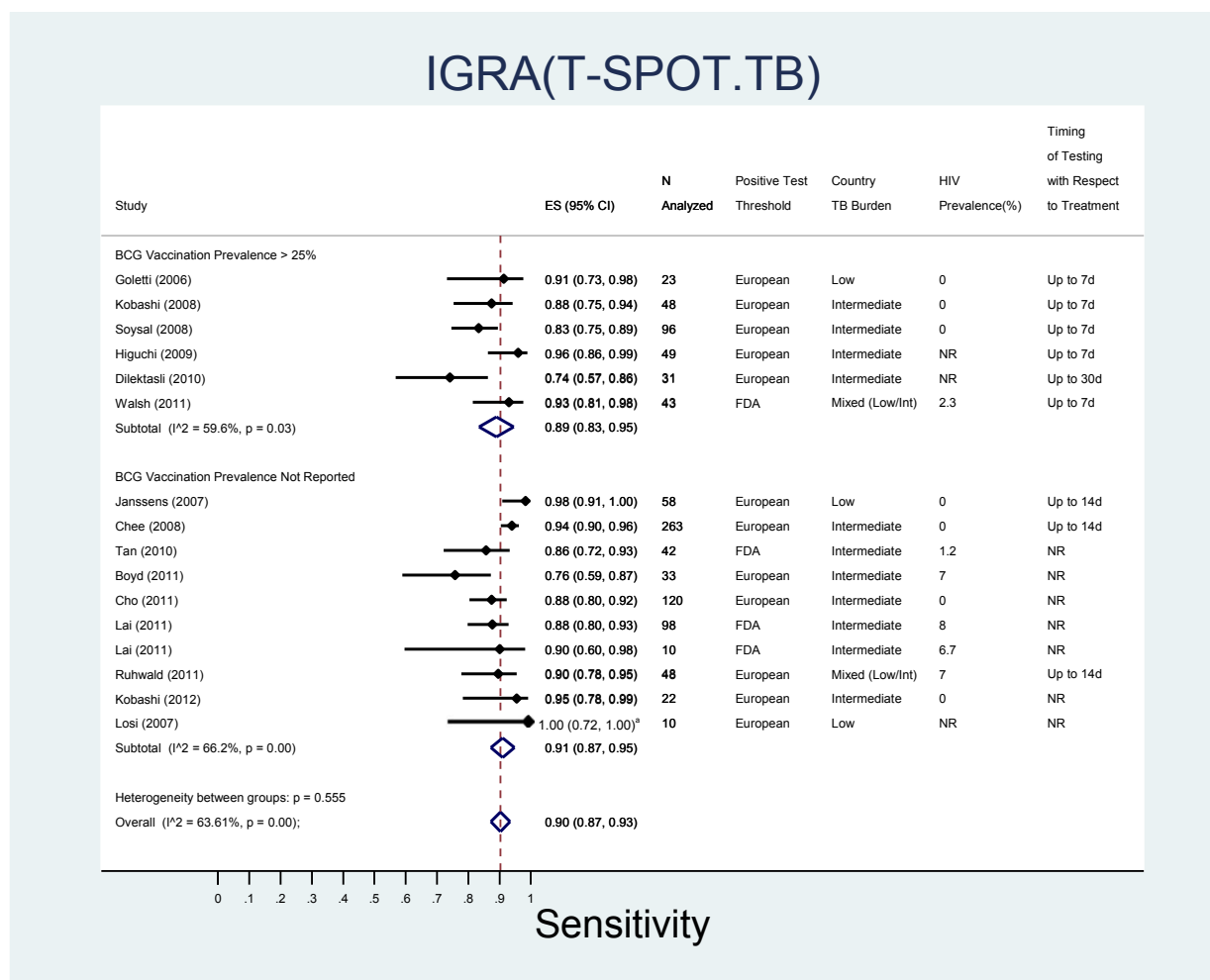
Appendix F Figure 22. Individual Study and Pooled Estimates (Where Applicable) of Sensitivity of T-SPOT.TB Test Stratified by Country TB Burden of the Study Setting



^a Excluded from pooled estimate due to point estimate of 1.0.

BCG = bacillus Calmette-Guerin; CI = confidence interval; d = day; ES = effect size; FDA = Food and Drug Administration; HIV = human immunodeficiency virus; IGRA = interferon gamma release assay; N = number; NR = not reported; TB = tuberculosis.

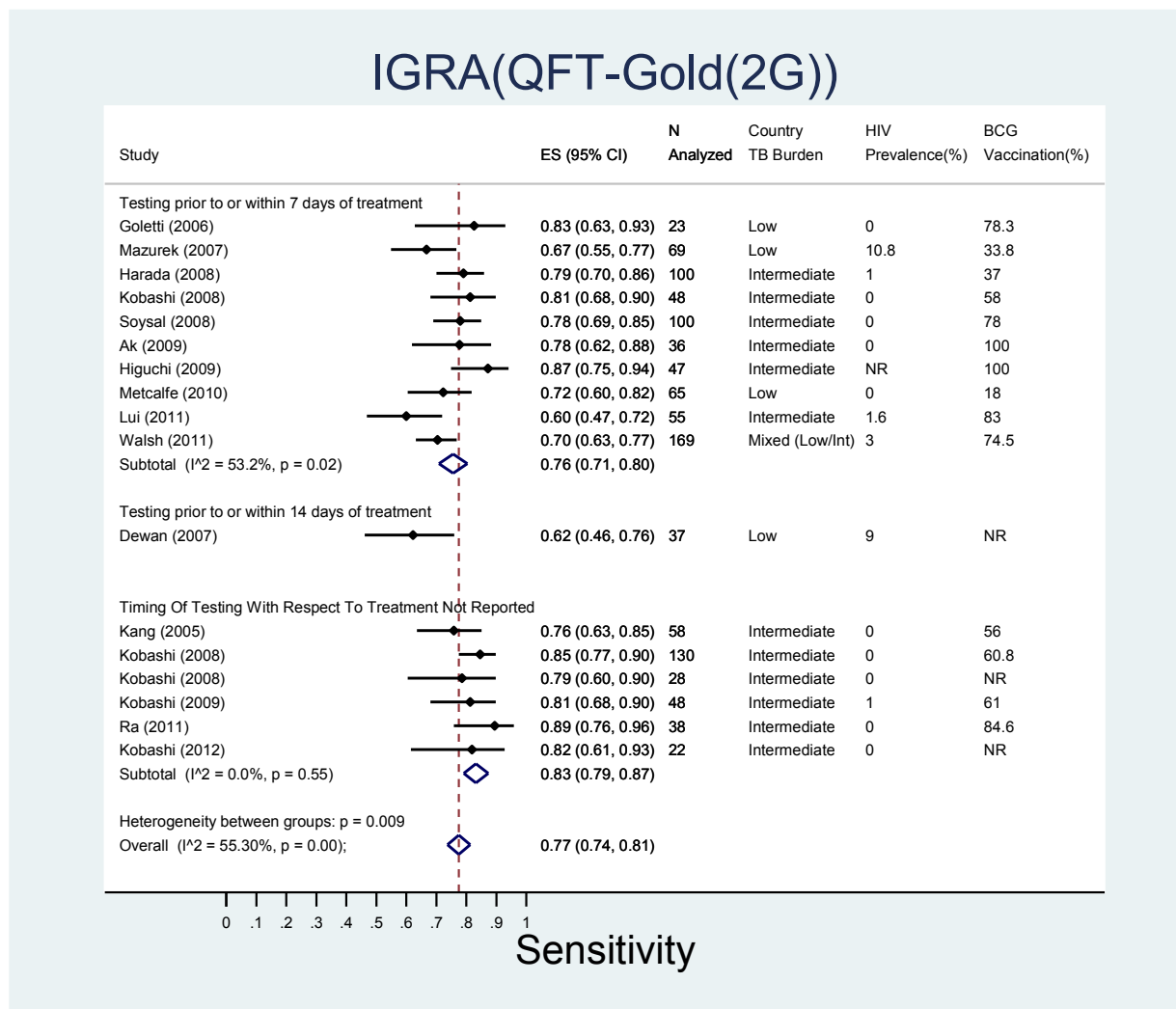
Appendix F Figure 23. Individual Study and Pooled Estimates (Where Applicable) of Sensitivity of T-SPOT.TB Test Stratified by Country BCG Vaccination Prevalence of the Study Population



^a Excluded from pooled estimate due to point estimate of 1.0.

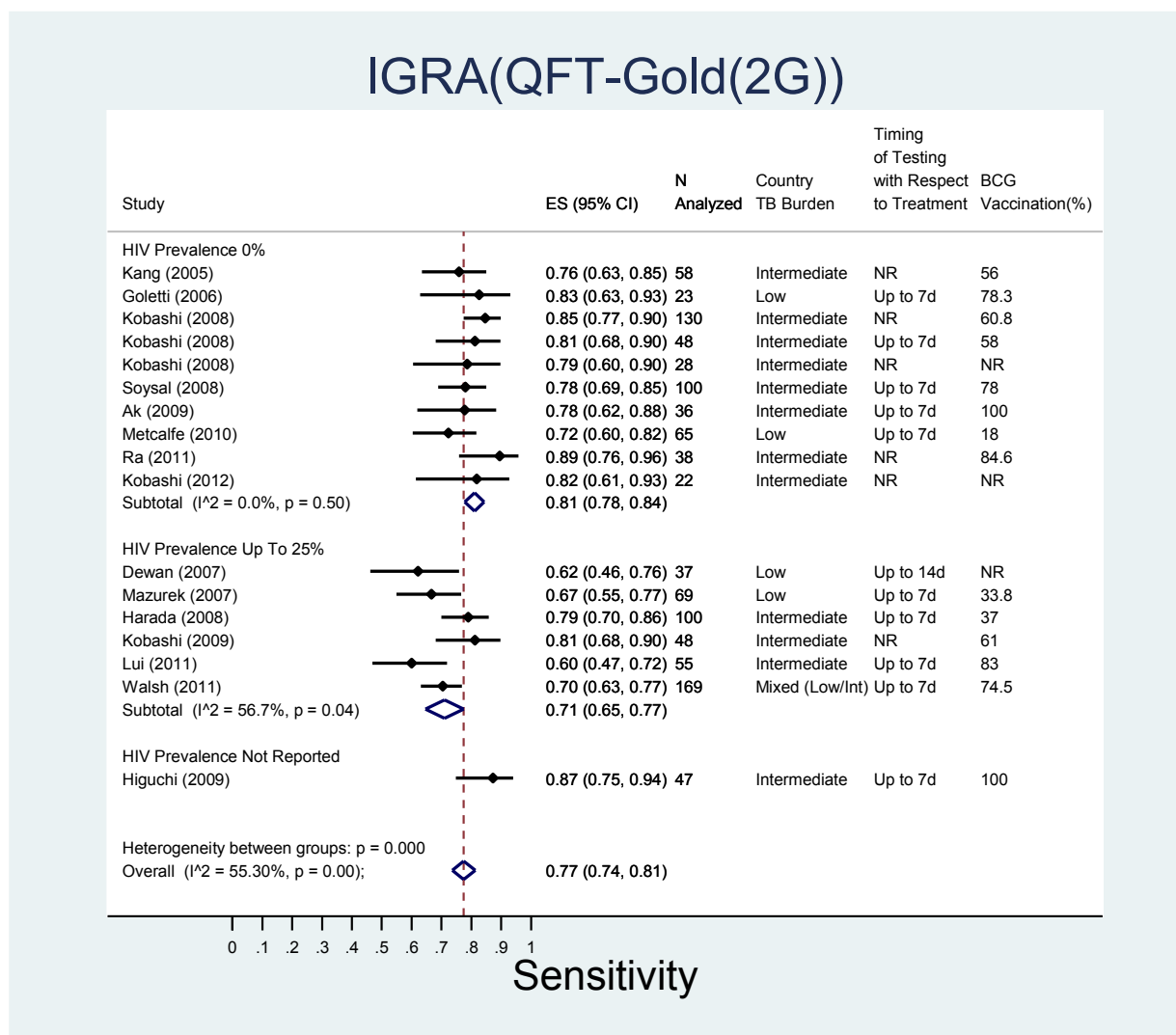
BCG = Bacillus Calmette-Guerin; CI = confidence interval; d = day; ES = effect size; FDA = Food and Drug Administration; HIV = human immunodeficiency virus; IGRA = interferon gamma release assay; N = number; NR = not reported; TB = tuberculosis.

Appendix F Figure 24. Individual Study and Pooled Estimates (Where Applicable) of Sensitivity of QFT-Gold (2nd Generation) Test Stratified by Timing of Testing With Respect to Antituberculosis Treatment



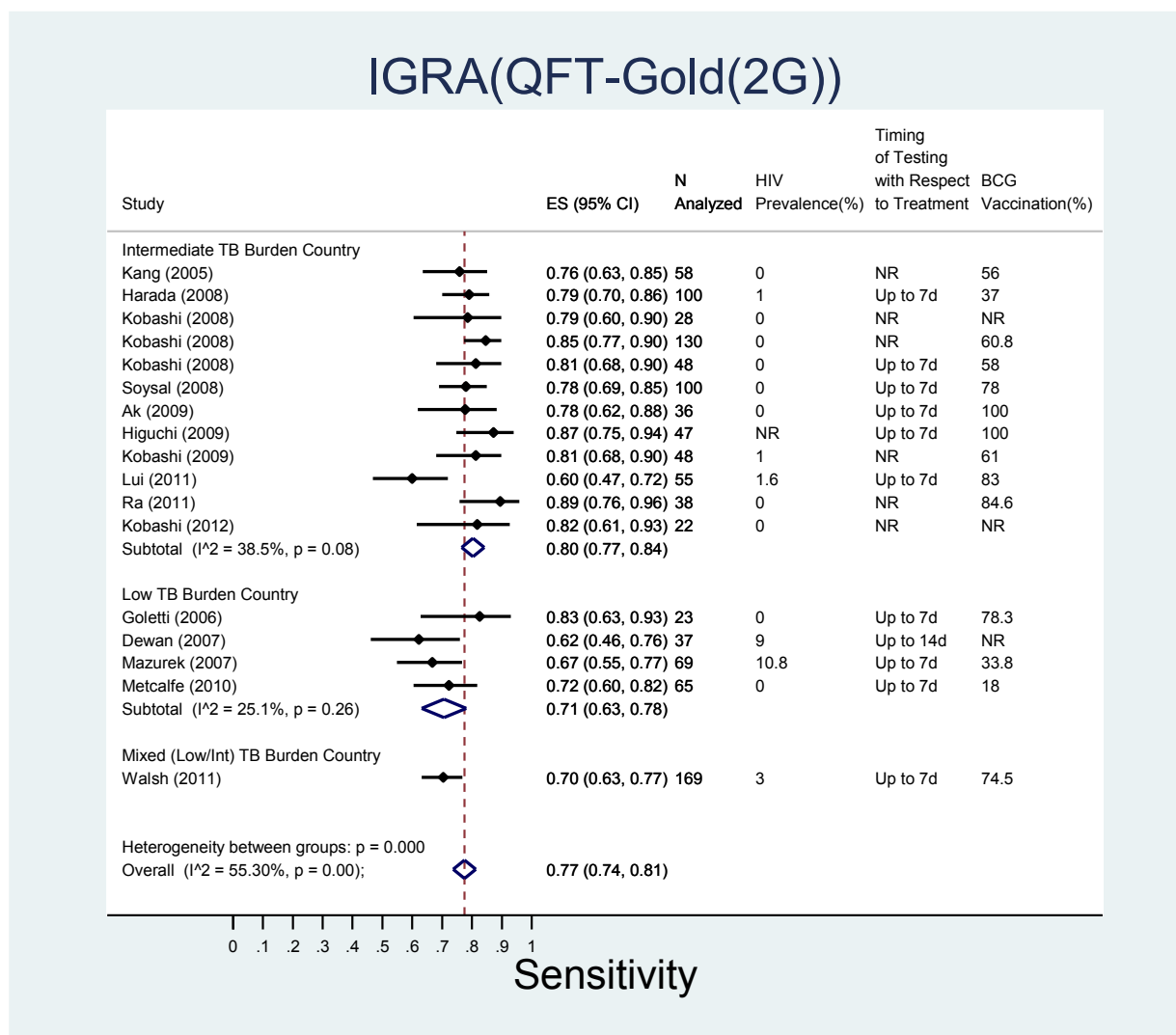
BCG = bacillus Calmette-Guerin; CI = confidence interval; ES = effect size; HIV = human immunodeficiency virus; IGRA = interferon gamma release assay; N = number; NR = not reported; TB = tuberculosis.

Appendix F Figure 25. Individual Study and Pooled Estimates (Where Applicable) of Sensitivity of QFT-Gold (2nd Generation) Test Stratified by HIV Prevalence of the Study Population



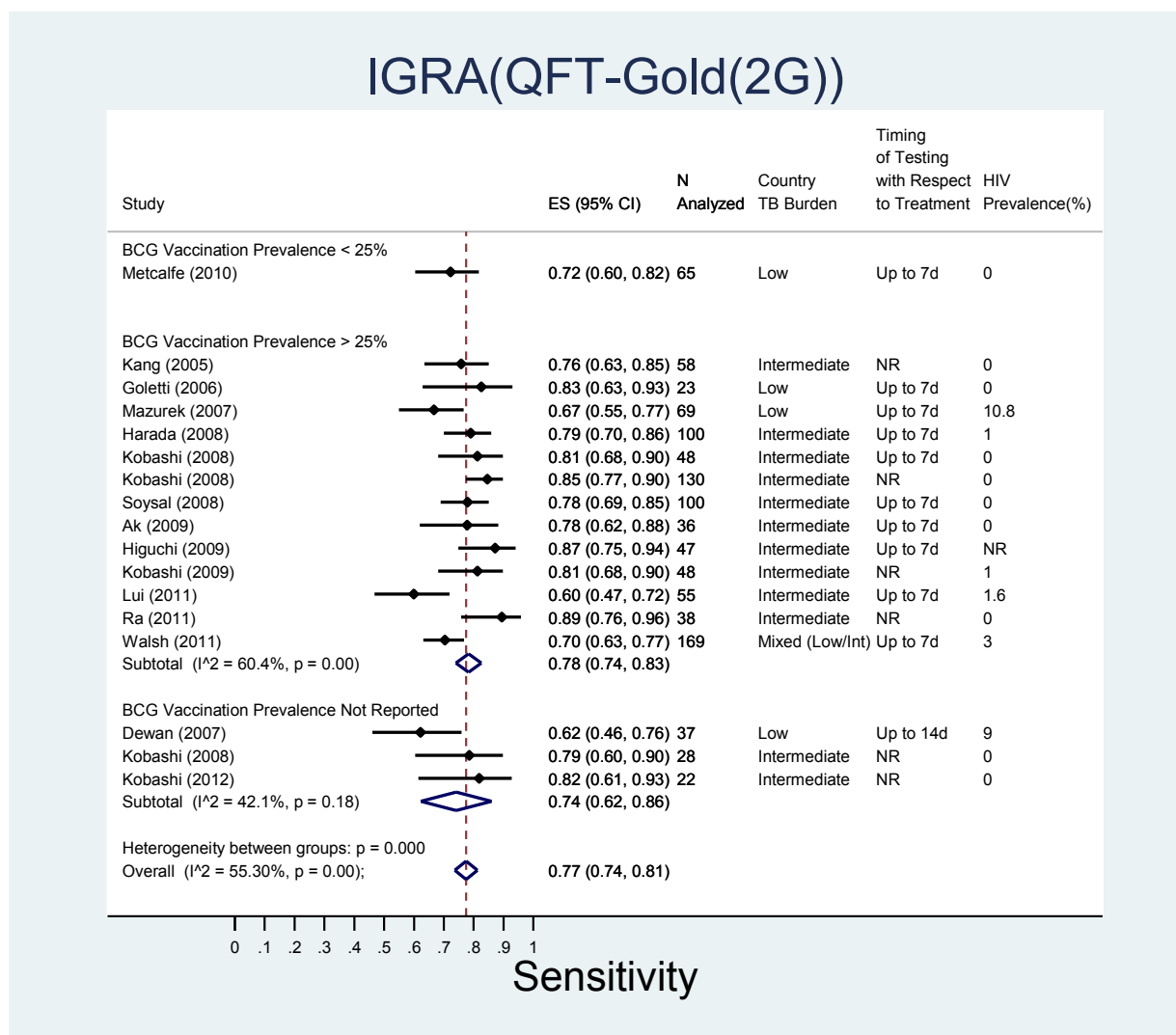
BCG = bacillus Calmette-Guerin; CI = confidence interval; d = day; ES = effect size; HIV = human immunodeficiency virus; IGRA = interferon gamma release assay; N = number; NR = not reported.

Appendix F Figure 26. Individual Study and Pooled Estimates (Where Applicable) of Sensitivity of QFT-Gold (2nd Generation) Test Stratified by Country TB Burden of the Study Setting



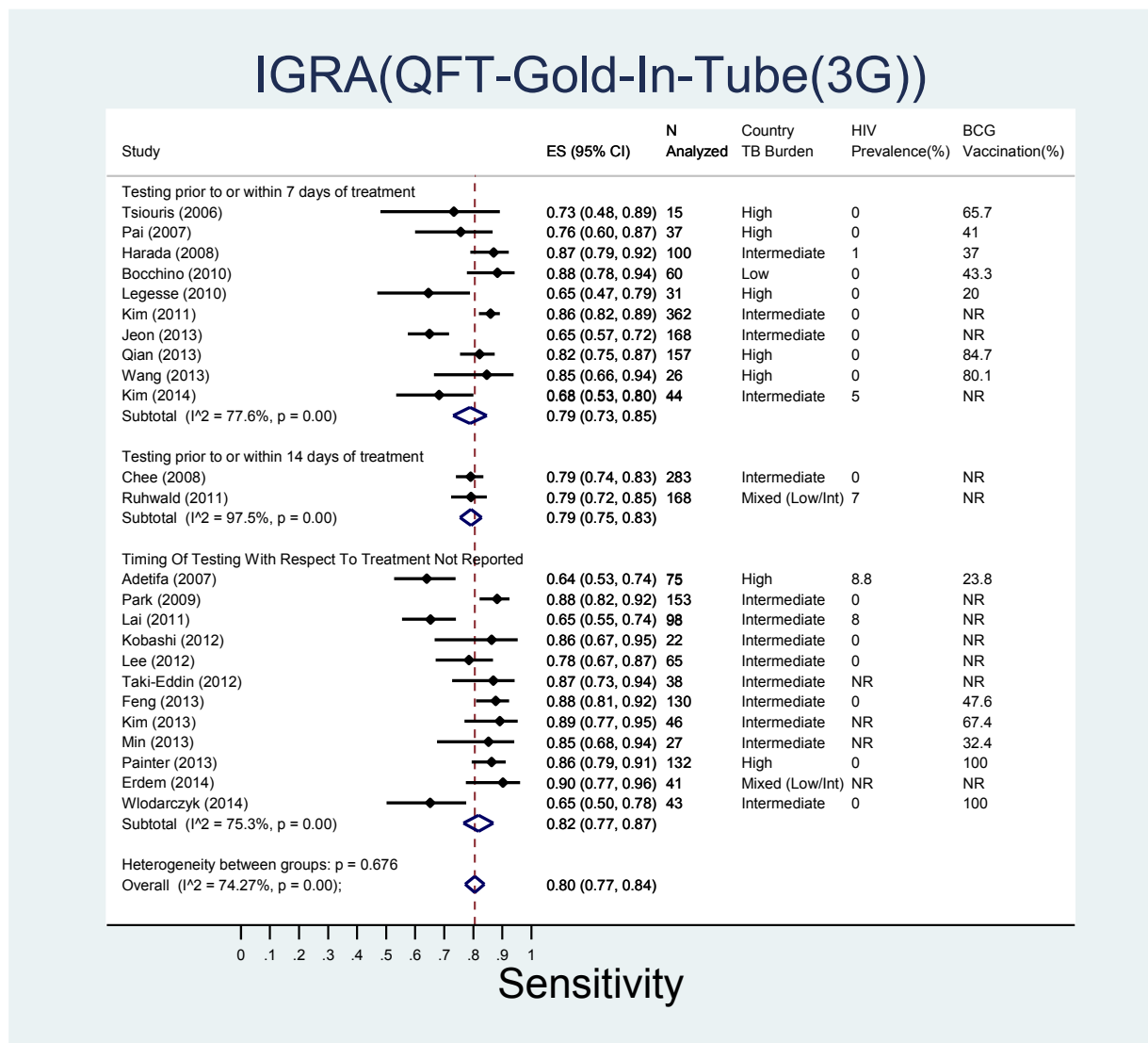
BCG = bacillus Calmette-Guerin; CI = confidence interval; d = day; ES = effect size; HIV = human immunodeficiency virus; IGRA = interferon gamma release assay; N = number; NR = not reported; TB = tuberculosis.

Appendix F Figure 27. Individual Study and Pooled Estimates (Where Applicable) of Sensitivity of QFT-Gold (2nd Generation) Test Stratified by Country BCG Vaccination Prevalence of the Study Population



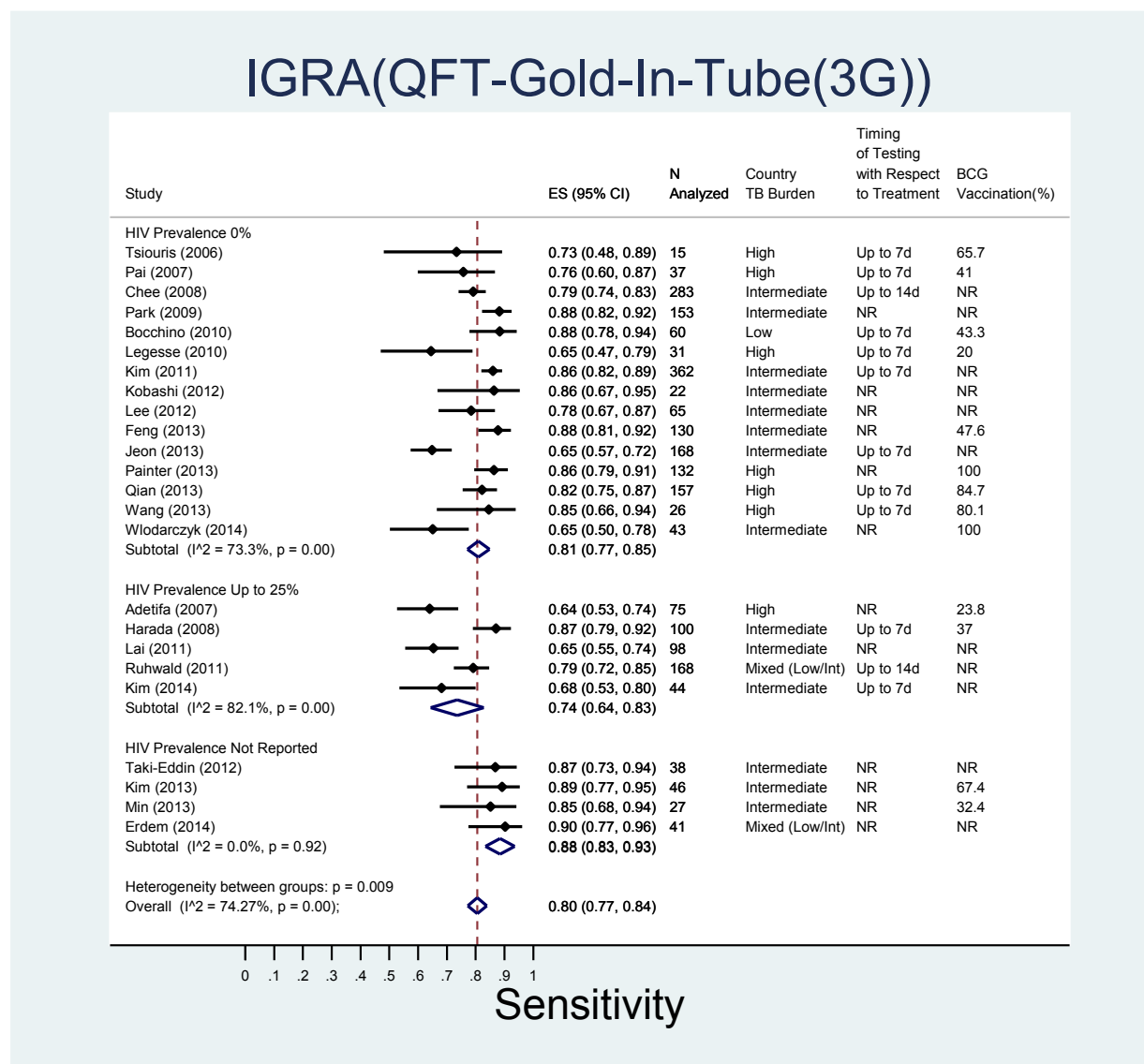
BCG = bacillus Calmette-Guerin; CI = confidence interval; d = day; ES = effect size; HIV = human immunodeficiency virus; IGRA = interferon gamma release assay; N = number; NR = not reported; TB = tuberculosis.

Appendix F Figure 28. Individual Study and Pooled Estimates (Where Applicable) of Sensitivity of QFT-Gold In-Tube (3rd Generation) Test Stratified by Timing of Testing With Respect to Antituberculosis Treatment



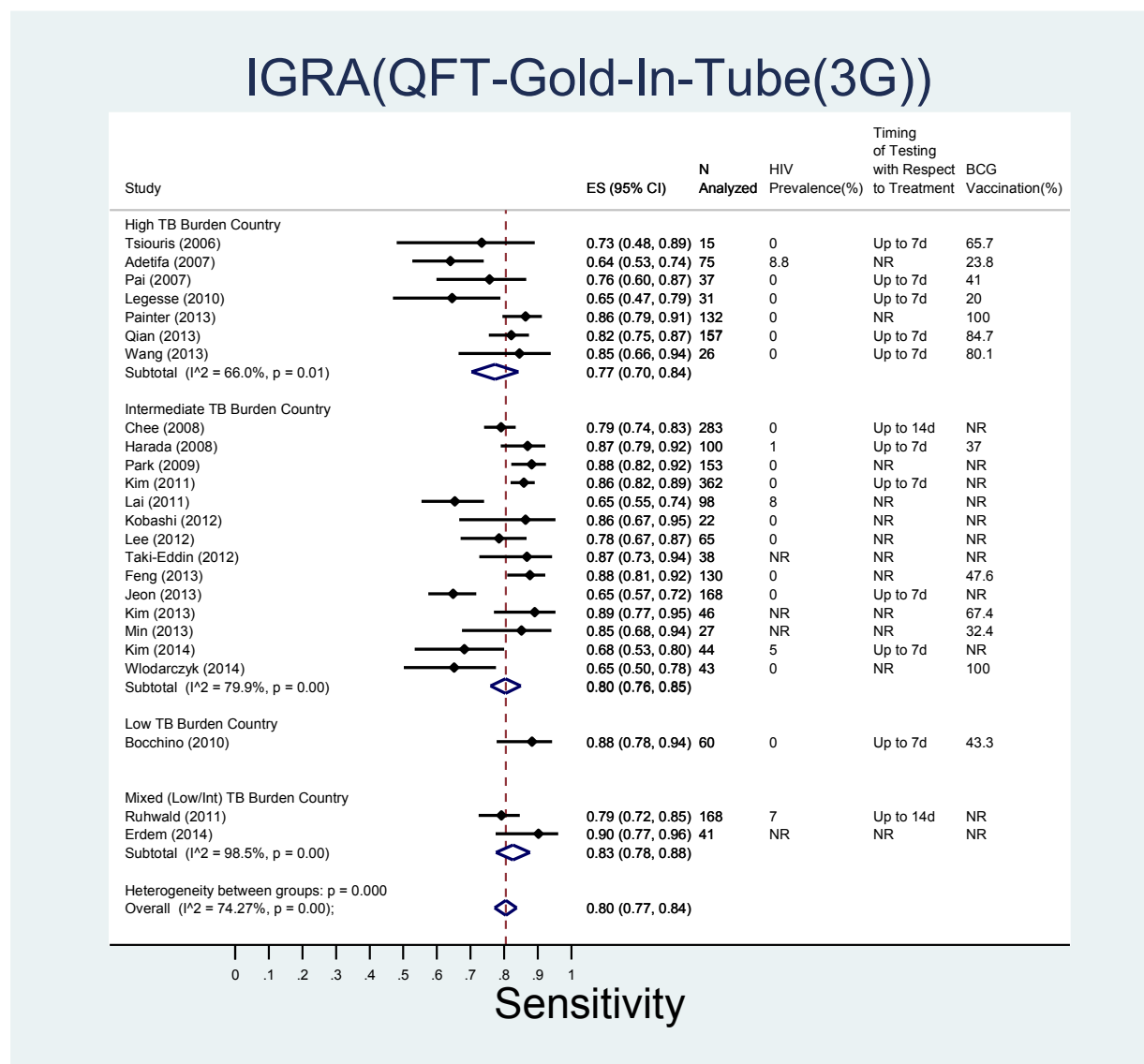
BCG = bacillus Calmette-Guerin; CI = confidence interval; ES = effect size; HIV = human immunodeficiency virus; IGRA = interferon gamma release assay; N = number; NR = not reported; TB = tuberculosis.

Appendix F Figure 29. Individual Study and Pooled Estimates (Where Applicable) of Sensitivity of QFT-Gold In-Tube (3rd Generation) Test Stratified by HIV Prevalence of the Study Population



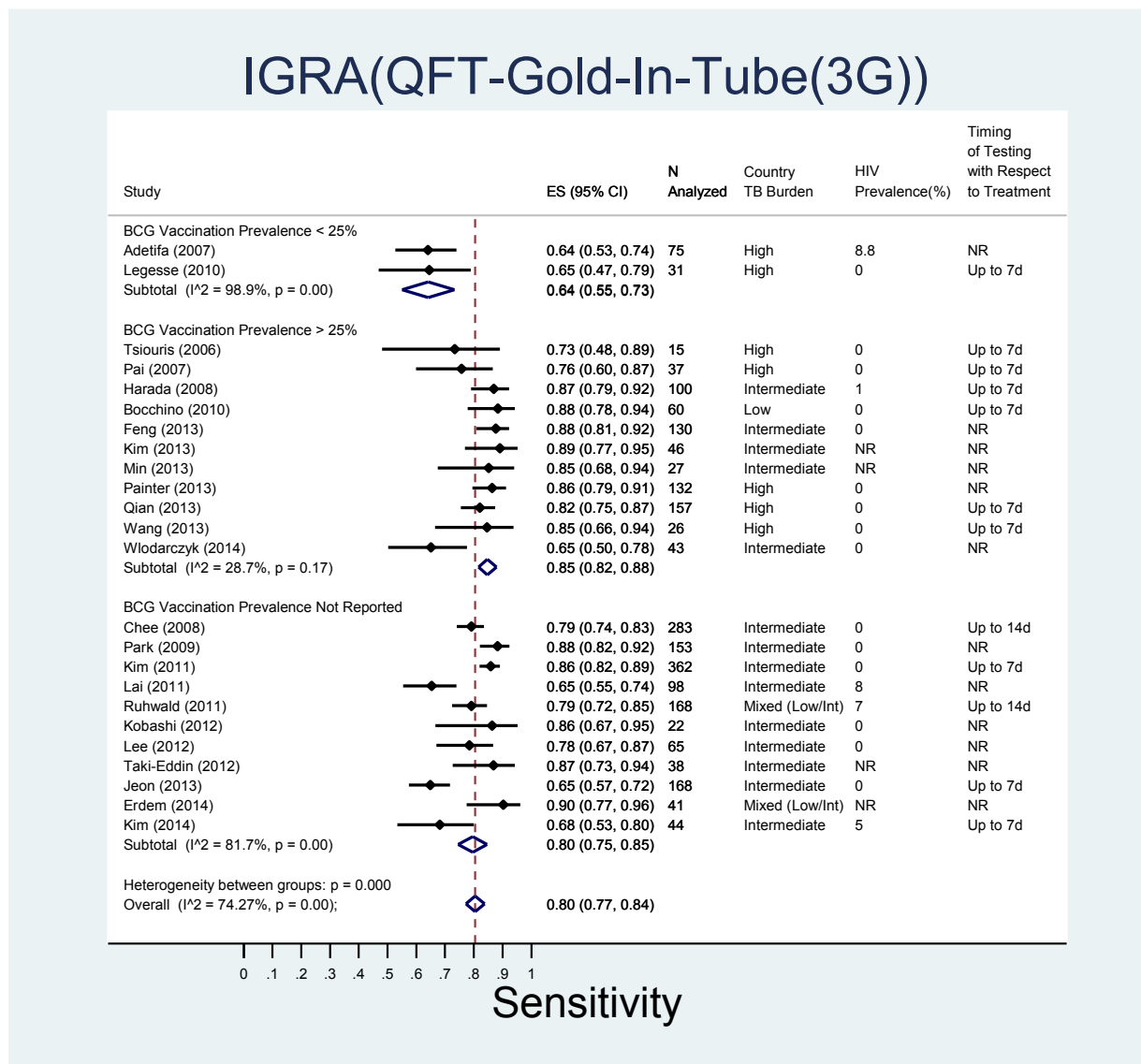
BCG = bacillus Calmette-Guerin; CI = confidence interval; d = day; ES = effect size; HIV = human immunodeficiency virus; IGRA = interferon gamma release assay; N = number; NR = not reported; TB = tuberculosis.

Appendix F Figure 30. Individual Study and Pooled Estimates (Where Applicable) of Sensitivity of QFT-Gold In-Tube (3rd Generation) Test Stratified by Country TB Burden of the Study Setting



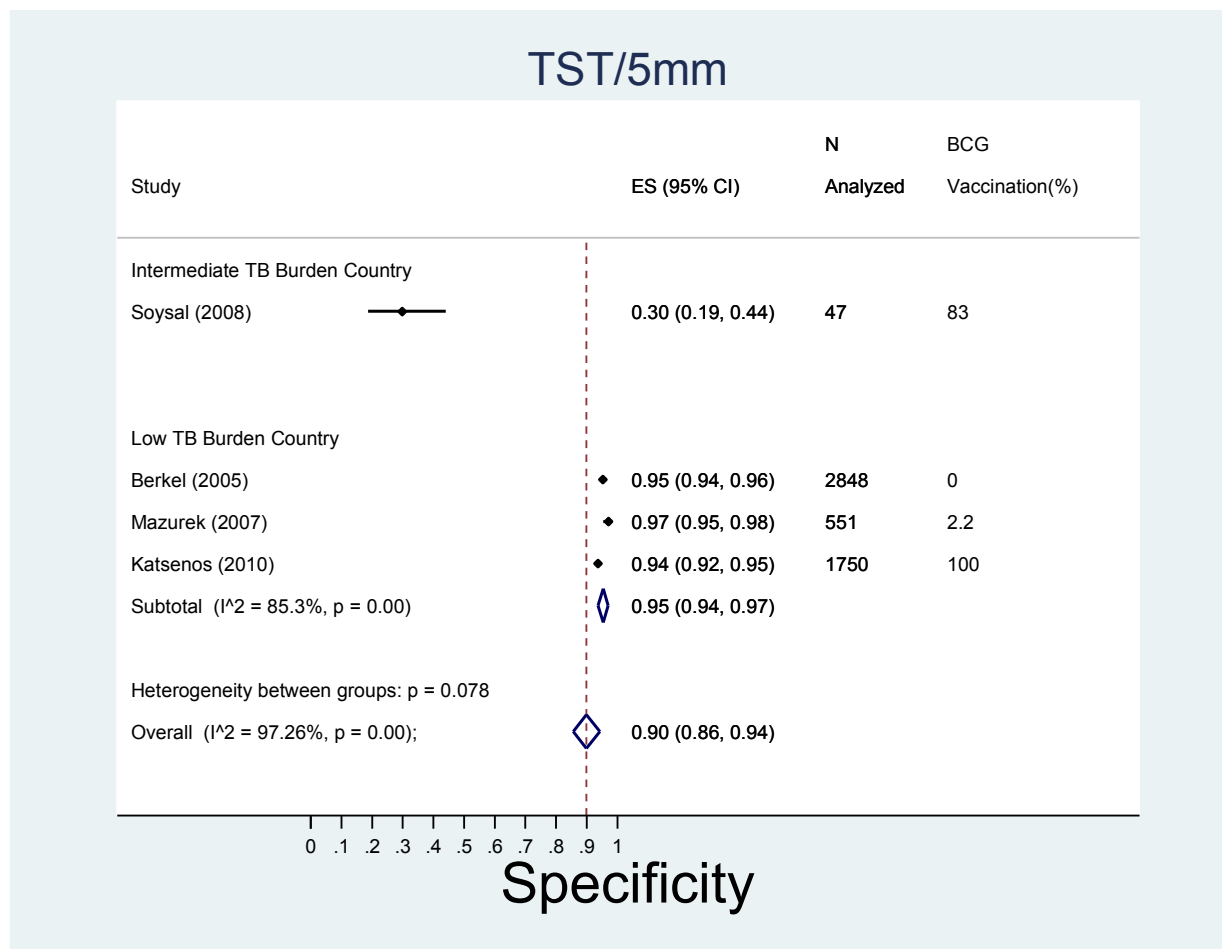
BCG = bacillus Calmette-Guerin; CI = confidence interval; d = day; ES = effect size; HIV = human immunodeficiency virus; IGRA = interferon gamma release assay; N = number; NR = not reported; TB = tuberculosis.

Appendix F Figure 31. Individual Study and Pooled Estimates (Where Applicable) of Sensitivity of QFT-Gold In-Tube (3rd Generation) Test Stratified by Country BCG Vaccination Prevalence of the Study Population



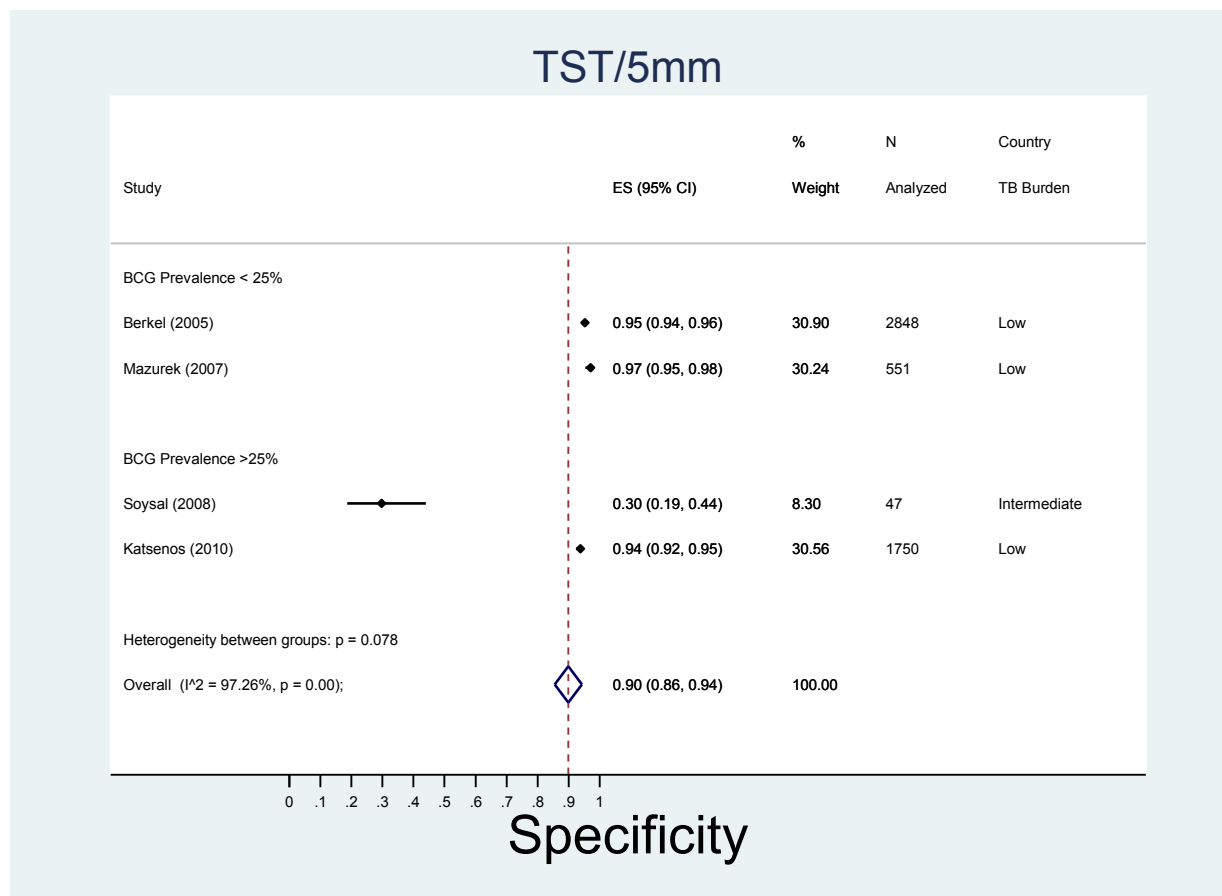
BCG = bacillus Calmette-Guerin; CI = confidence interval; d = day; ES = effect size; HIV = human immunodeficiency virus; IGRA = interferon gamma release assay; N = number; NR = not reported; TB = tuberculosis.

Appendix F Figure 32. Individual Study and Pooled Estimates (Where Applicable) of Specificity of TST at 5-mm Threshold Stratified by Country TB Burden of the Study Setting



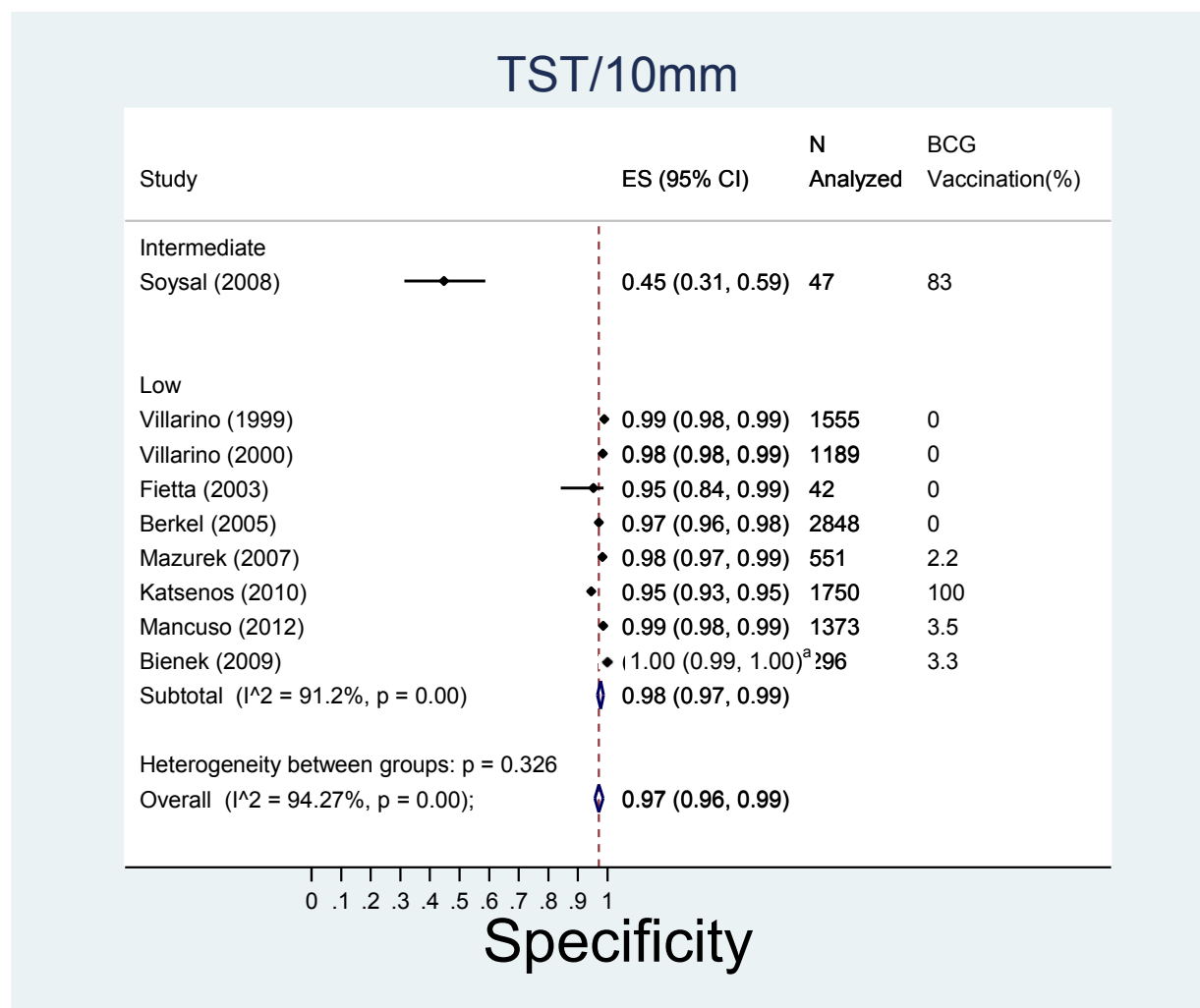
BCG = bacillus Calmette-Guerin; CI = confidence interval; ES = effect size; mm = millimeter; N = number; TB = tuberculosis; TST = tuberculin skin test.

Appendix F Figure 33. Individual Study and Pooled Estimates (Where Applicable) of Specificity of TST at 5-mm Threshold Stratified by Country BCG Vaccination Prevalence of the Study Population



BCG = bacillus Calmette-Guerin; CI = confidence interval; ES = effect size; mm = millimeter; N = number; TB = tuberculosis; TST = tuberculin skin test.

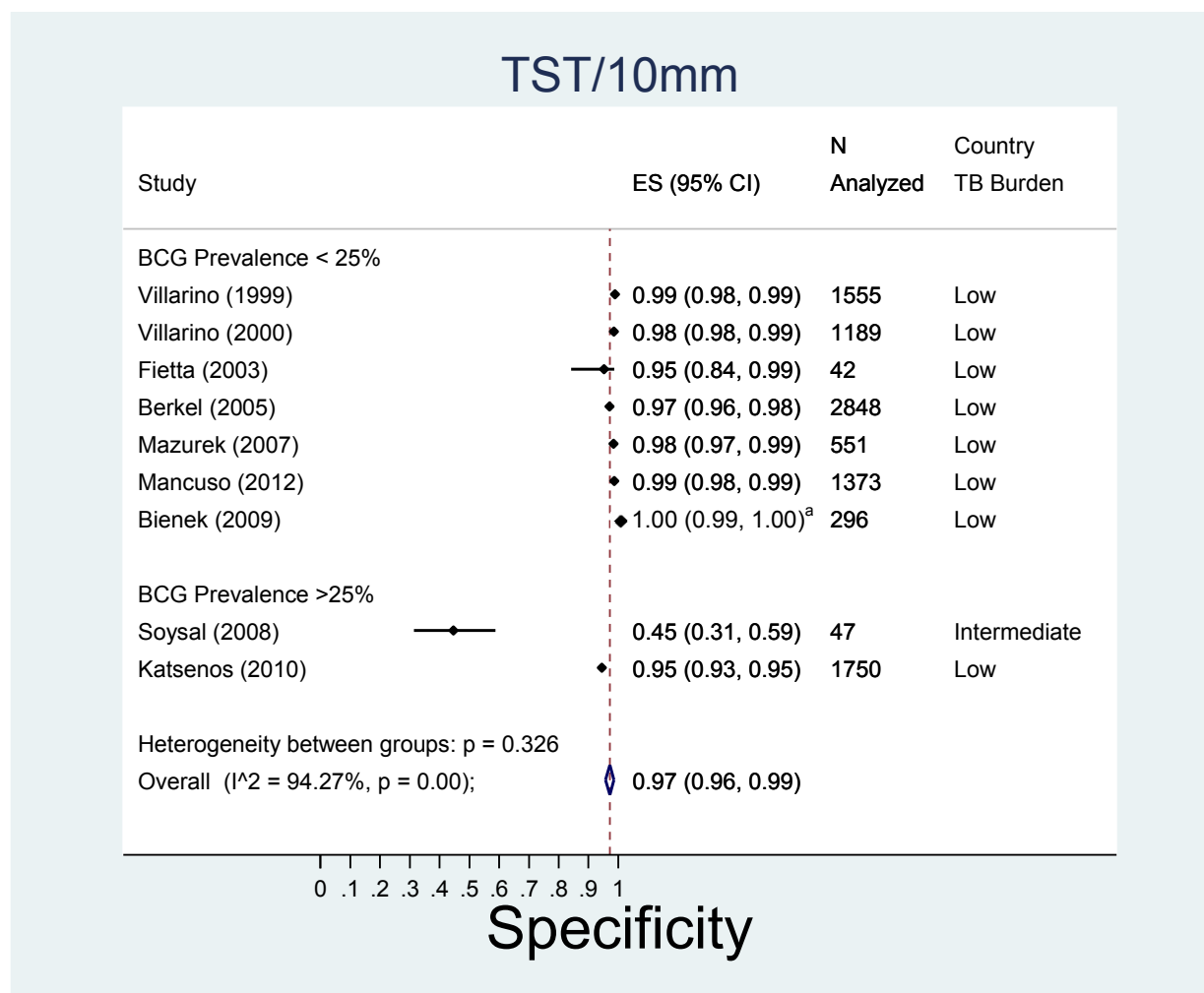
Appendix F Figure 34. Individual Study and Pooled Estimates (Where Applicable) of Specificity of TST at 10-mm Threshold Stratified by Country TB Burden of the Study Setting



^a Excluded from pooled estimate due to point estimate of 1.0.

BCG = bacillus Calmette-Guerin; CI = confidence interval; ES = effect size; mm = millimeter; N = number; TB = tuberculosis; TST = tuberculin skin test.

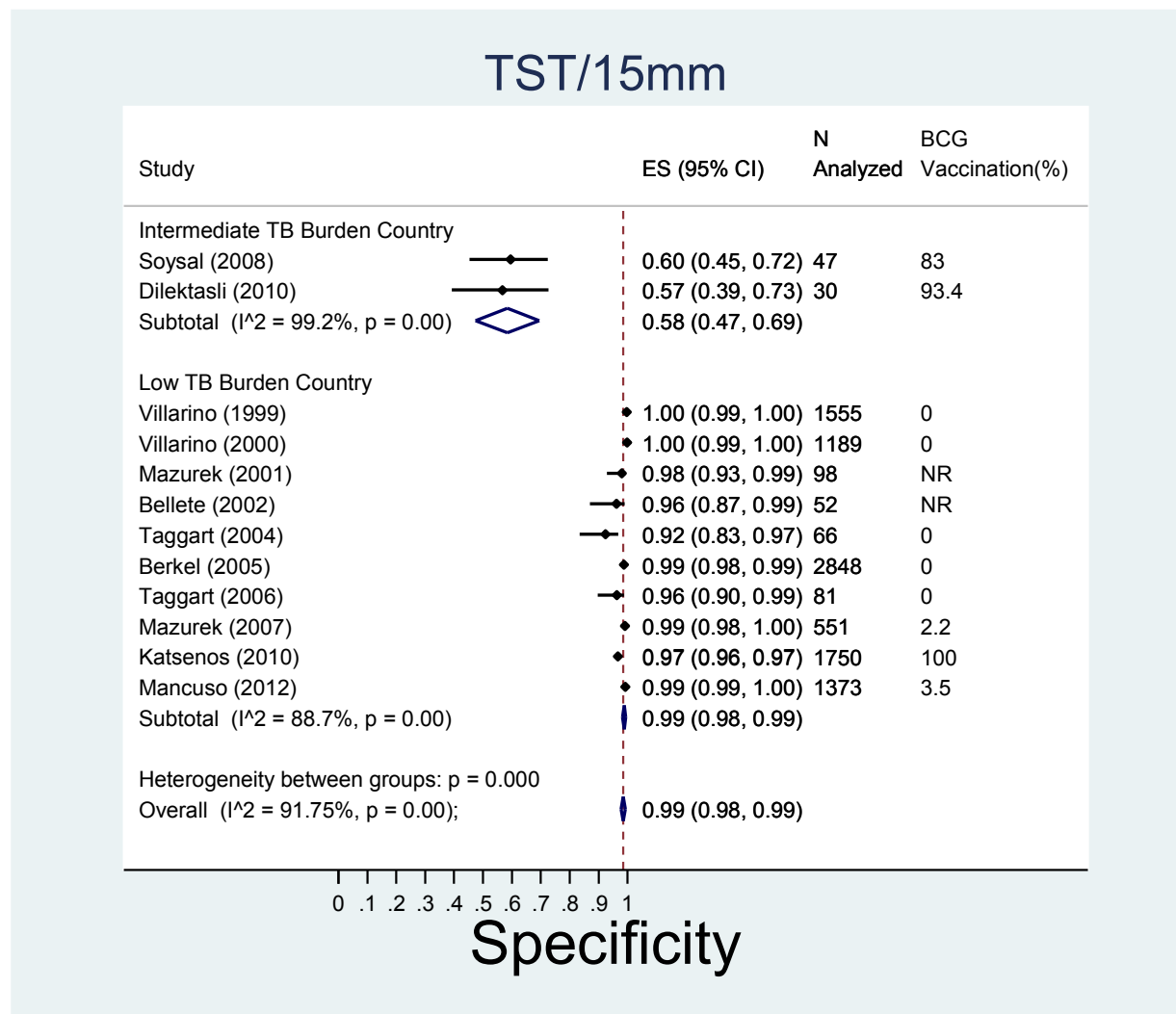
Appendix F Figure 35. Individual Study and Pooled Estimates (Where Applicable) of Specificity of TST at 10-mm Threshold Stratified by Country BCG Vaccination Prevalence of the Study Population



^a Excluded from pooled estimate due to point estimate of 1.0.

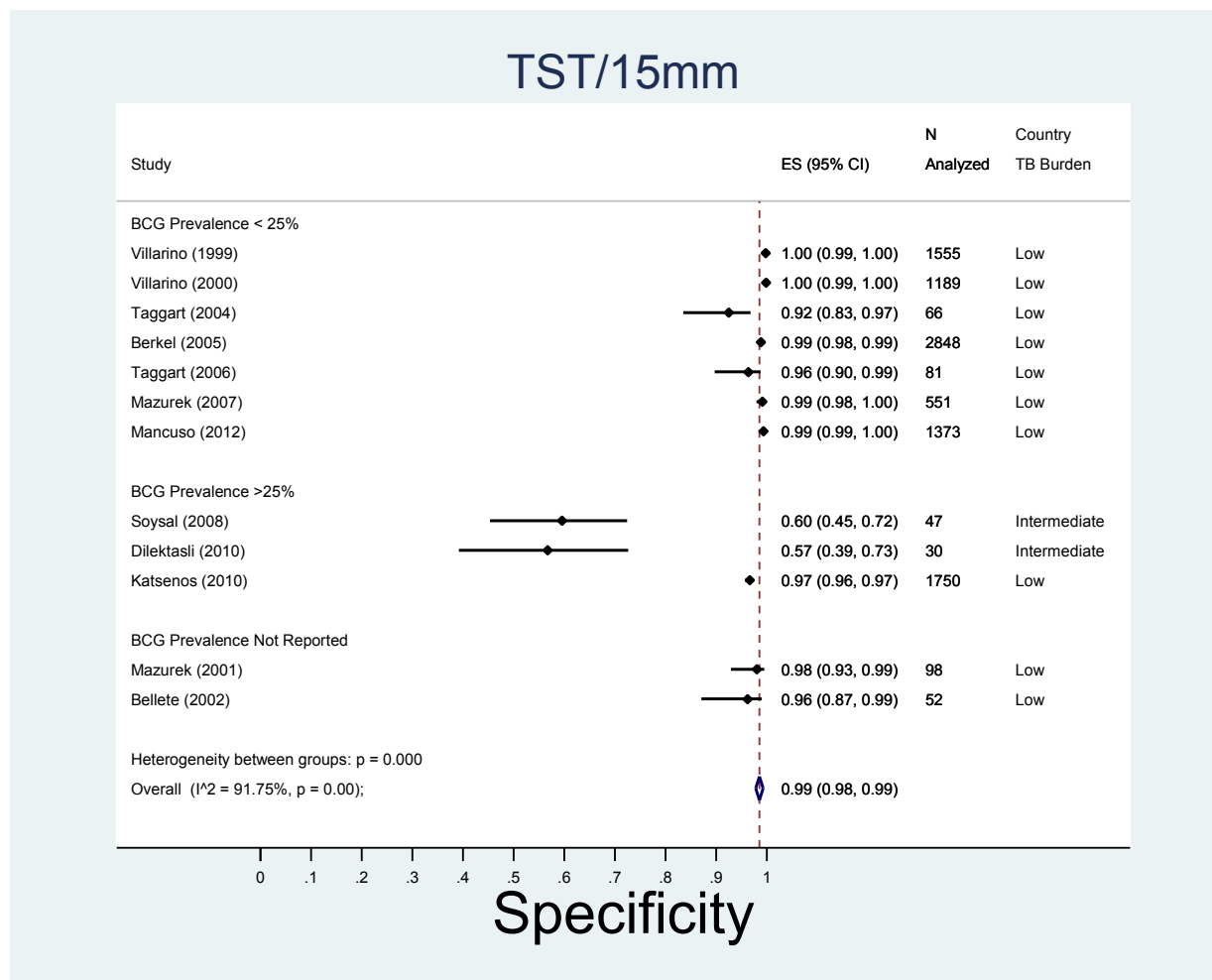
BCG = bacillus Calmette-Guerin; CI = confidence interval; ES = effect size; mm = millimeter; N = number; TB = tuberculosis; TST = tuberculin skin test.

Appendix F Figure 36. Individual Study and Pooled Estimates (Where Applicable) of Specificity of TST at 15-mm Threshold Stratified by Country TB Burden of the Study Setting



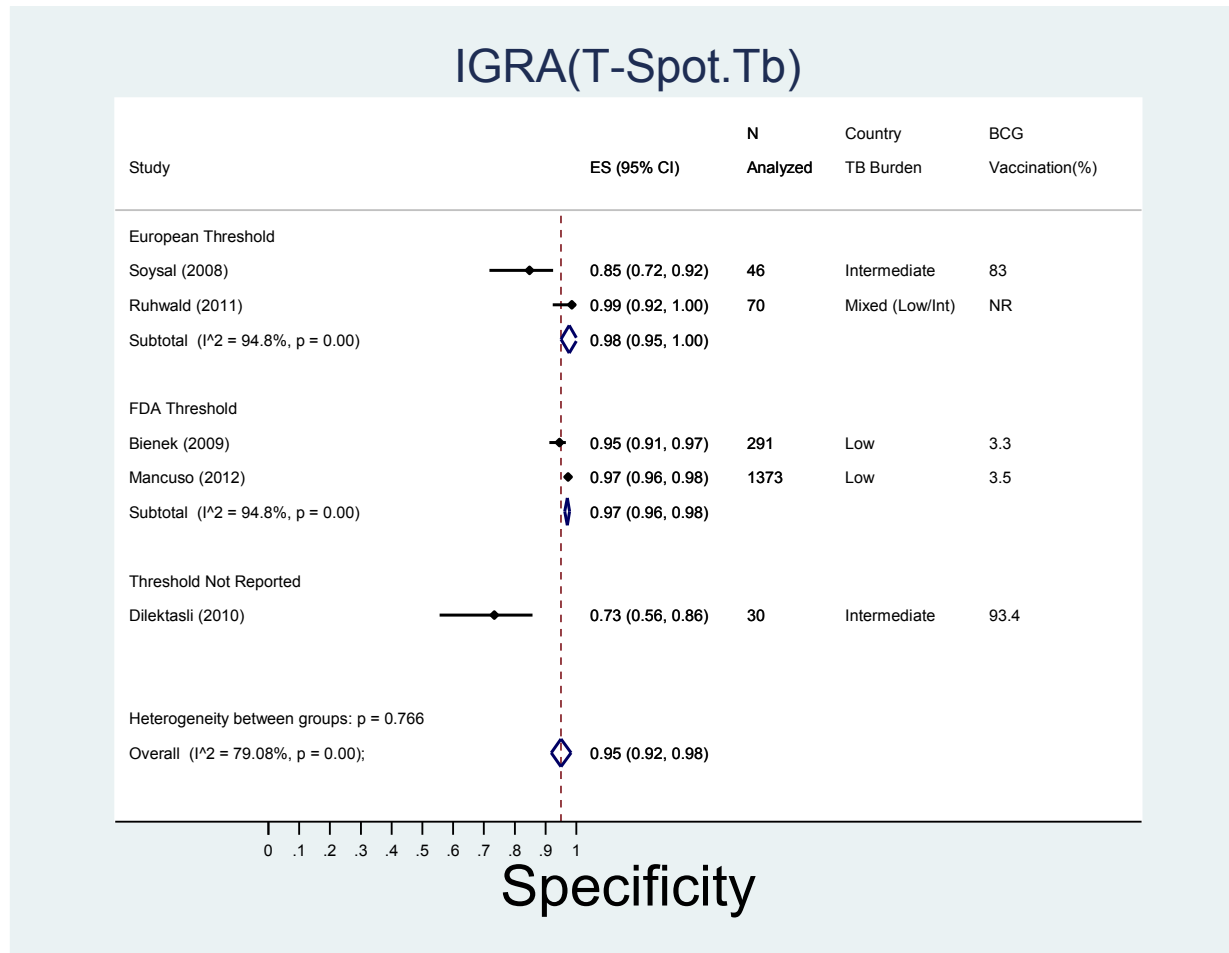
BCG = bacillus Calmette-Guerin; CI = confidence interval; ES = effect size; mm = millimeter; N = number; TB = tuberculosis; TST = tuberculin skin test.

Appendix F Figure 37. Individual Study and Pooled Estimates (Where Applicable) of Specificity of TST at 15-mm Threshold Stratified by Country BCG Vaccination Prevalence of the Study Population



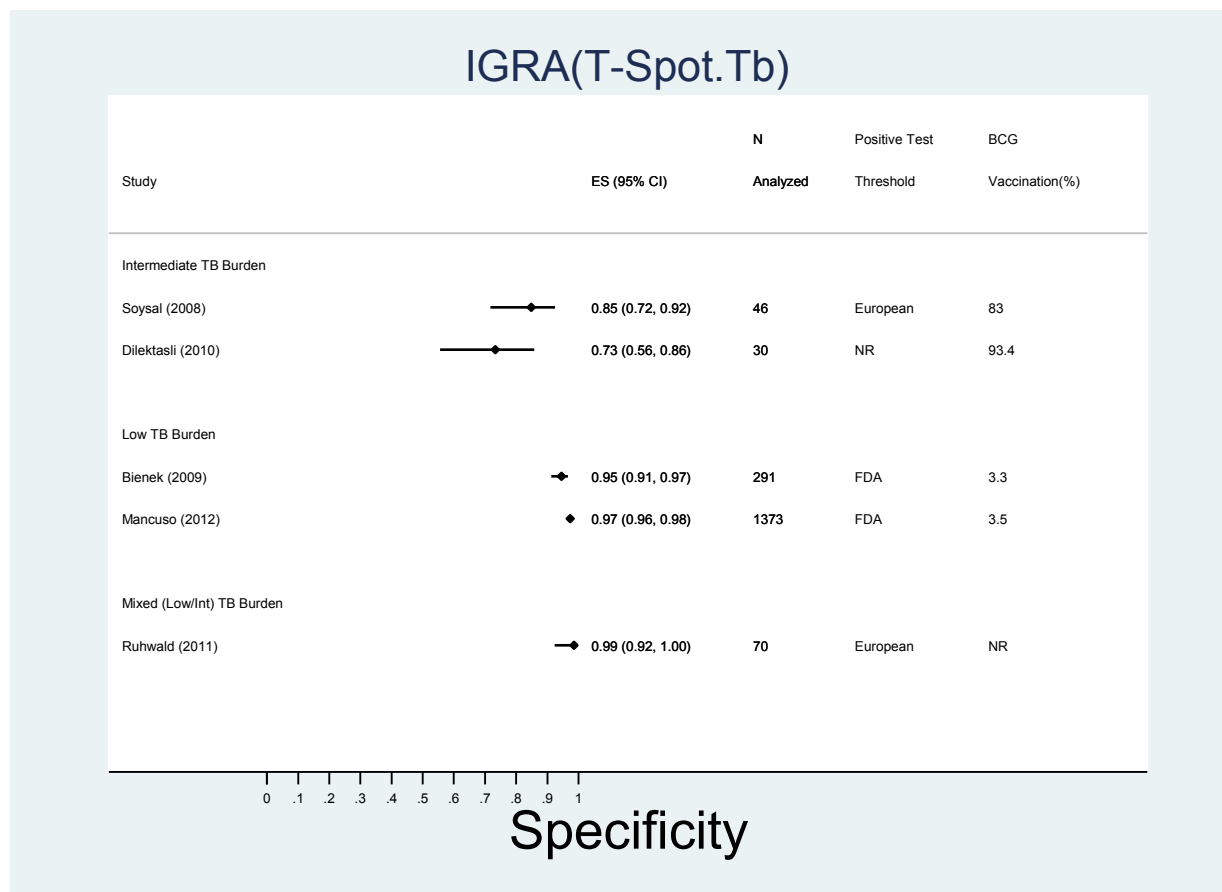
BCG = bacillus Calmette-Guerin; CI = confidence interval; ES = effect size; mm = millimeter; N = number; TB = tuberculosis; TST = tuberculin skin test.

Appendix F Figure 38. Individual Study and Pooled Estimates (Where Applicable) of Specificity of IGRA (T-SPOT.TB) Test Stratified by Country TB Burden of the Study Setting



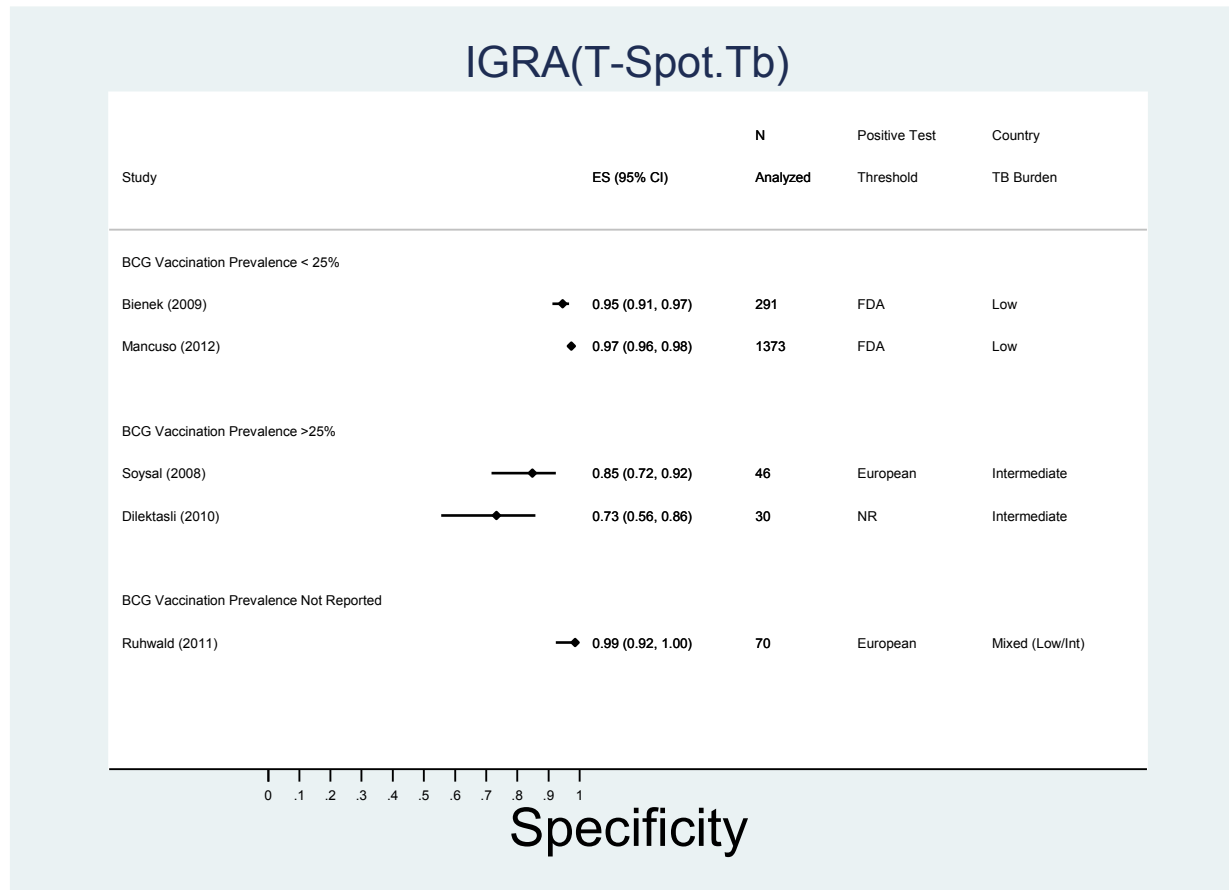
BCG = bacillus Calmette-Guerin; CI = confidence interval; ES = effect size; FDA = Food and Drug Administration; IGRA = interferon gamma release assay; N = number; TB = tuberculosis.

Appendix F Figure 39. Individual Study and Pooled Estimates (Where Applicable) of Specificity of IGRA (T-SPOT.TB) Test Stratified by Country TB Burden of the Study Setting



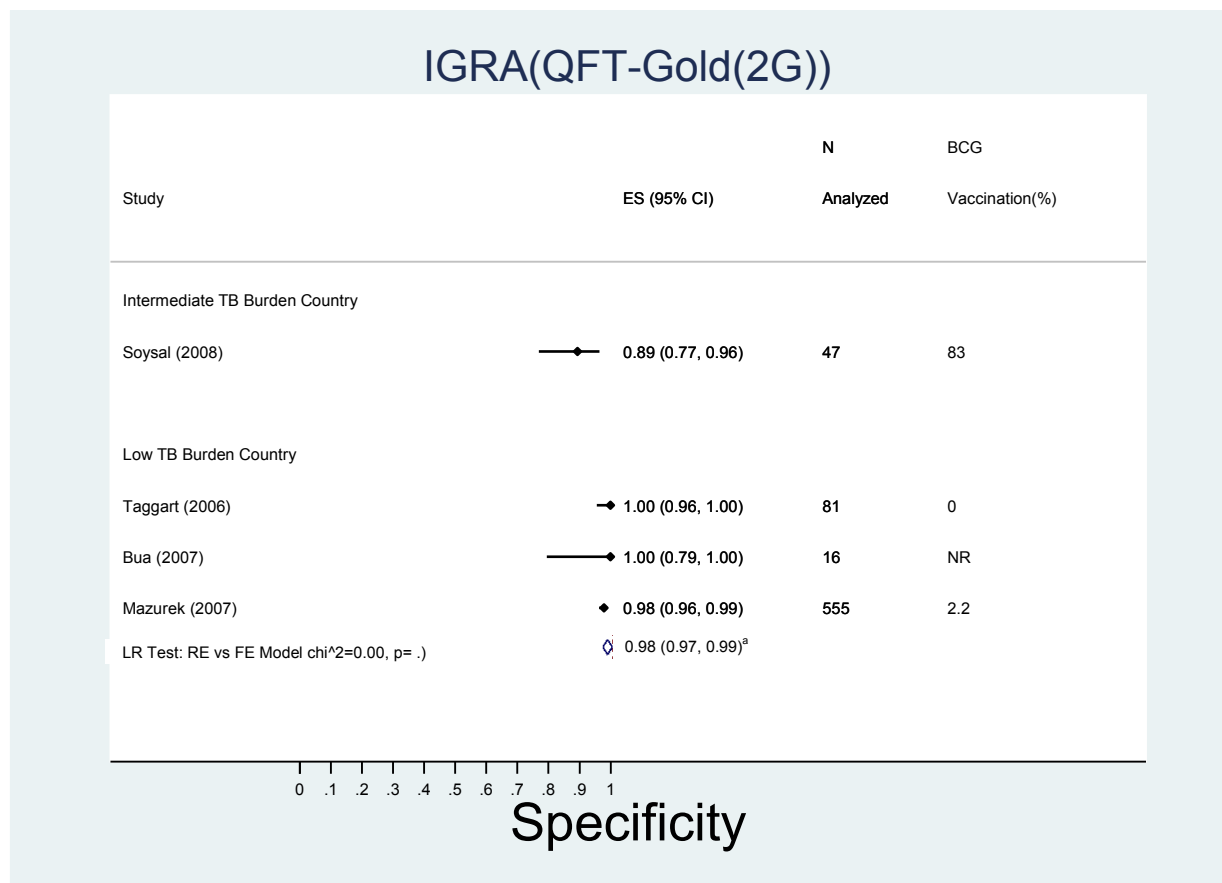
BCG = bacillus Calmette-Guerin; CI = confidence interval; ES = effect size; IGRA = interferon gamma release assay; N = number; TB = tuberculosis.

Appendix F Figure 40. Individual Study and Pooled Estimates (Where Applicable) of Specificity of IGRA (T-SPOT.TB) Test Stratified by Country BCG Vaccination Prevalence of the Study Population



BCG = bacillus Calmette-Guerin; CI = confidence interval; ES = effect size; FDA = Food and Drug Administration; IGRA = interferon gamma release assay; N = number; TB = tuberculosis.

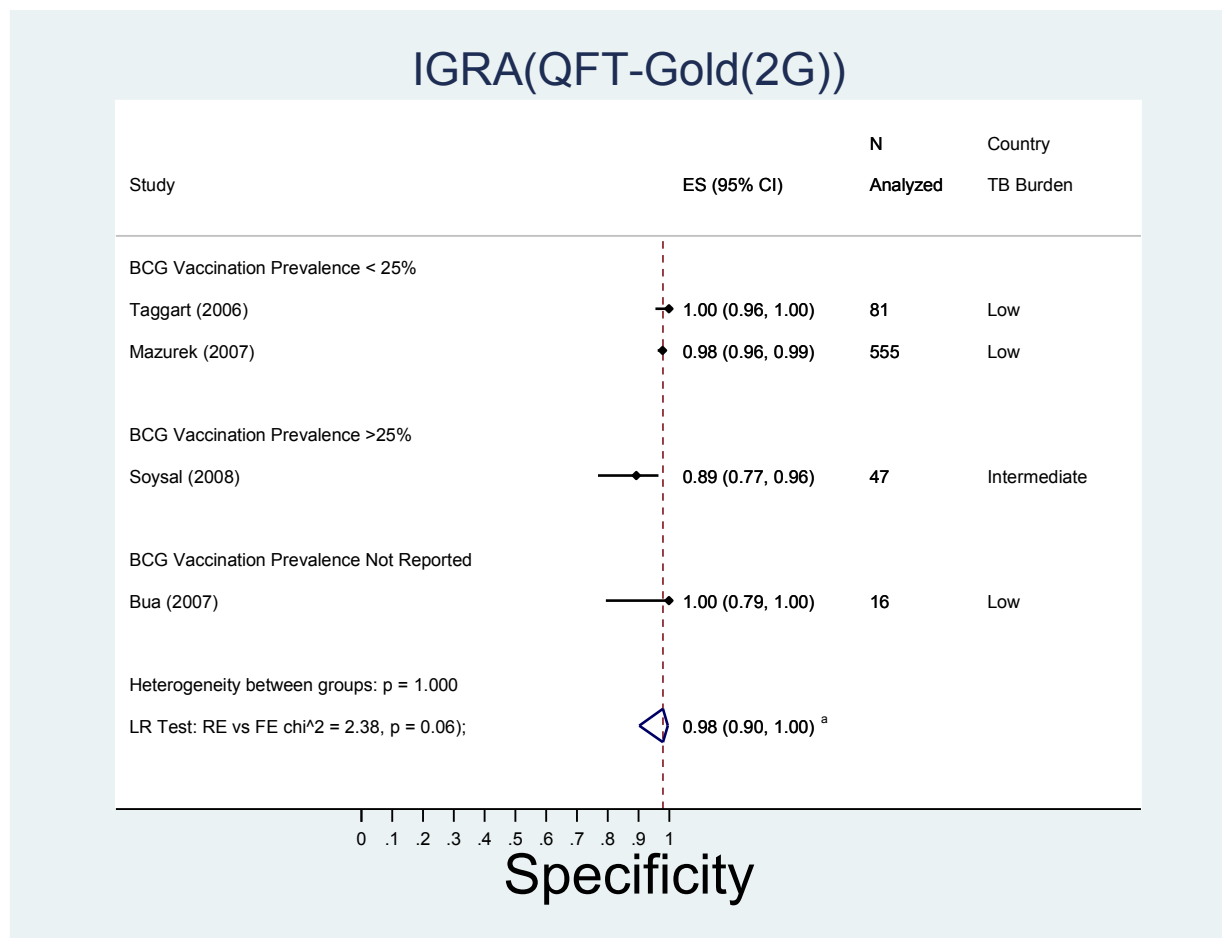
Appendix F Figure 41. Individual Study and Pooled Estimates (Where Applicable) of Specificity of QFT-Gold (2nd Generation) Stratified by Country TB Burden of the Study Setting



^a Subgroup pooled estimate using maximum likelihood estimator because of two point estimates of 1.0.

BCG = bacillus Calmette-Guerin; CI = confidence interval; ES = effect size; IGRA = interferon gamma release assay; N = number; NR = not reported; TB = tuberculosis.

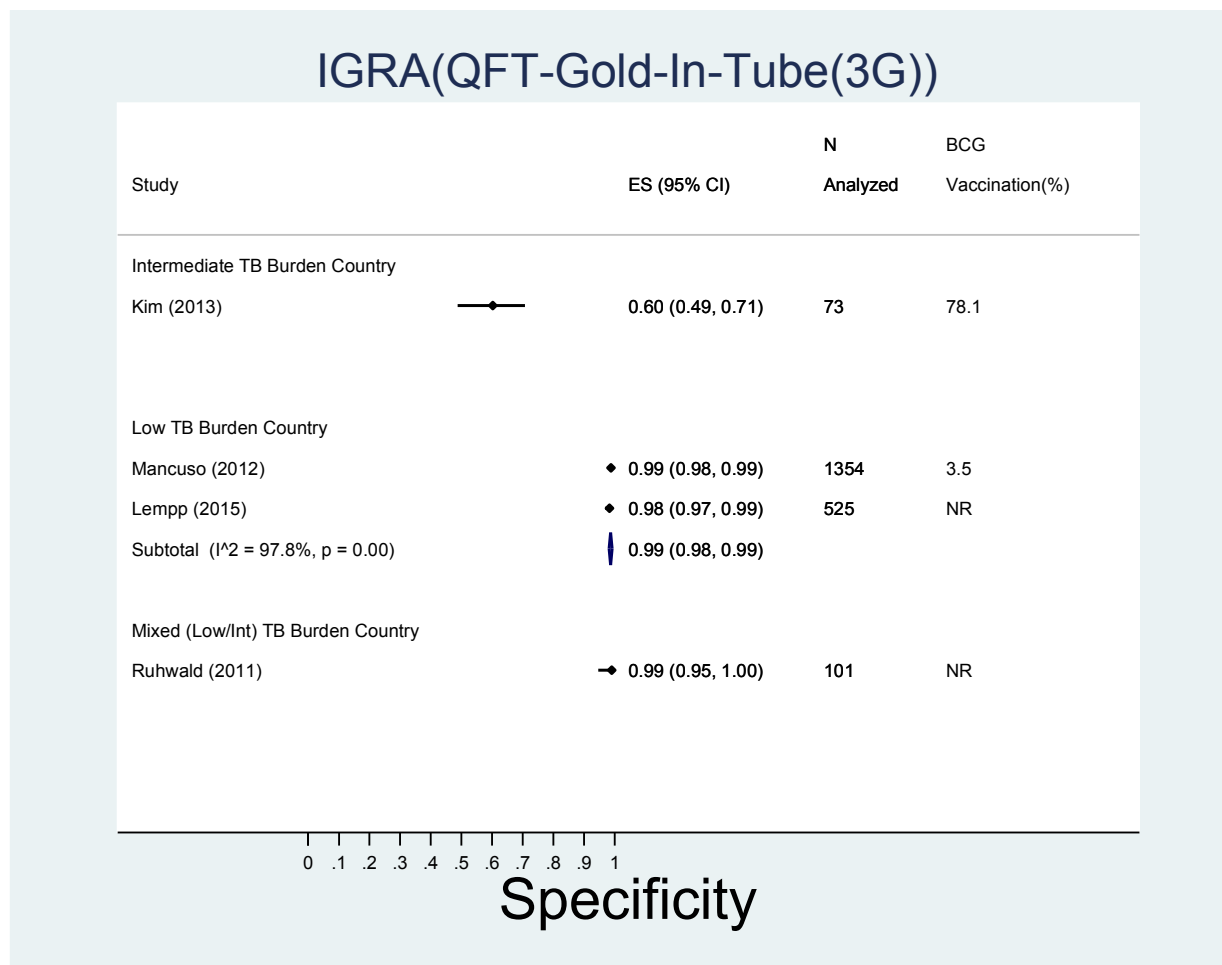
Appendix F Figure 42. Individual Study and Pooled Estimates (Where Applicable) of Specificity of QFT-Gold (2nd Generation) Stratified by Country BCG Vaccination Prevalence of the Study Population



^a Pooled estimate using maximum likelihood estimator because of two point estimates of 1.0.

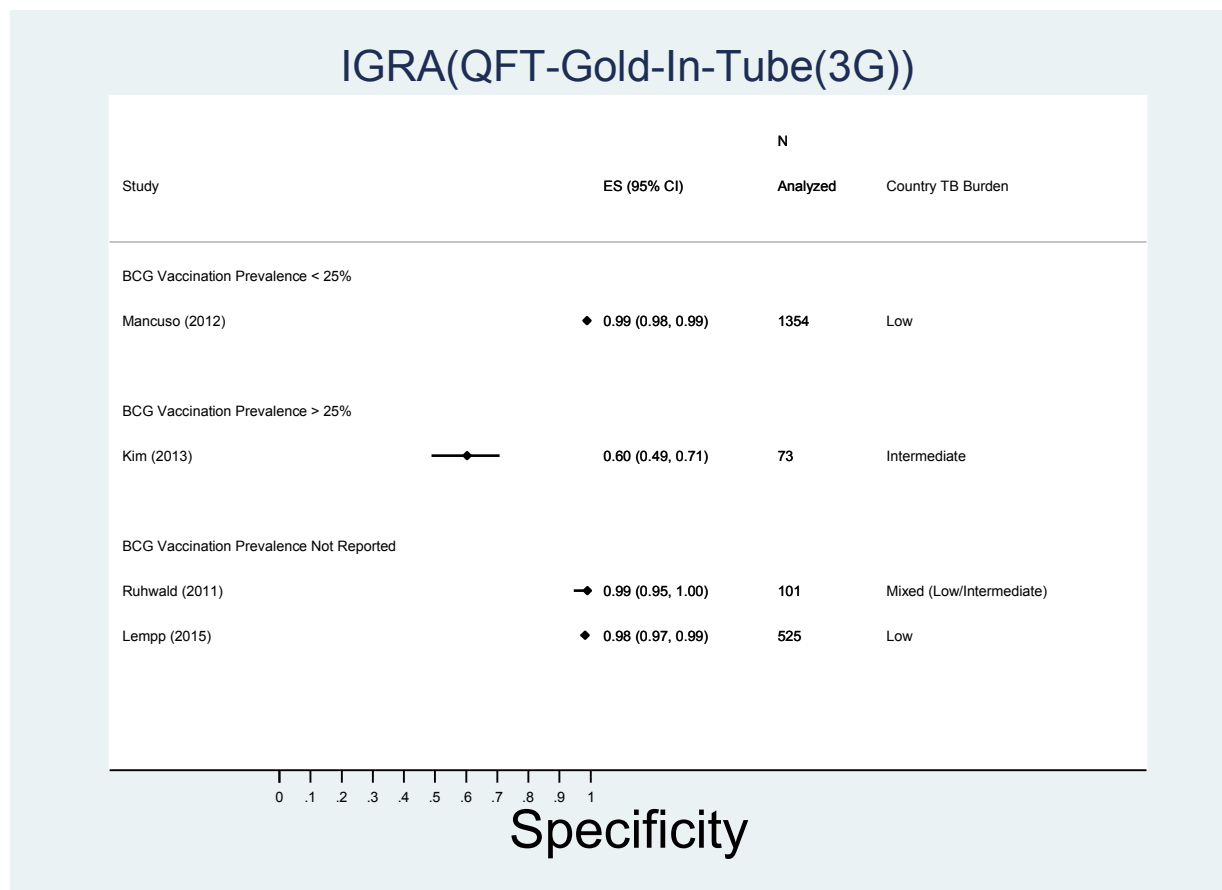
BCG = bacillus Calmette-Guerin; CI = confidence interval; ES = effect size; IGRA = interferon gamma release assay; N = number; TB = tuberculosis.

Appendix F Figure 43. Individual Study and Pooled Estimates (Where Applicable) of Specificity of QFT-Gold-In-Tube (3rd Generation) Stratified by Country TB Burden of the Study Setting



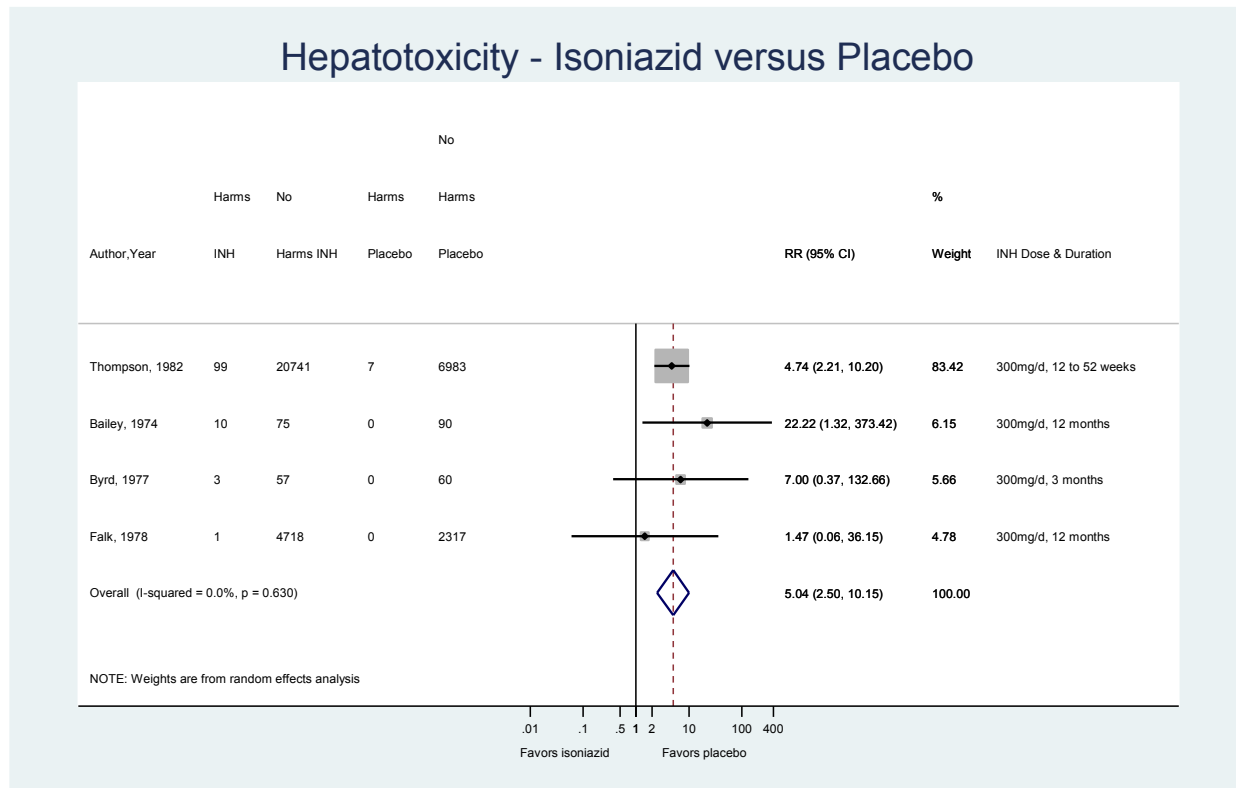
BCG = bacillus Calmette-Guerin; CI = confidence interval; ES = effect size; IGRA = interferon gamma release assay; N = number; NR = not reported; TB = tuberculosis.

Appendix F Figure 44. Individual Study and Pooled Estimates (Where Applicable) of Specificity of QFT-Gold-In-Tube (3rd Generation) Stratified by Country BCG Vaccination Prevalence of the Study Population



BCG = bacillus Calmette-Guerin; CI = confidence interval; ES = effect size; IGRA = interferon gamma release assay; N = number; TB = tuberculosis.

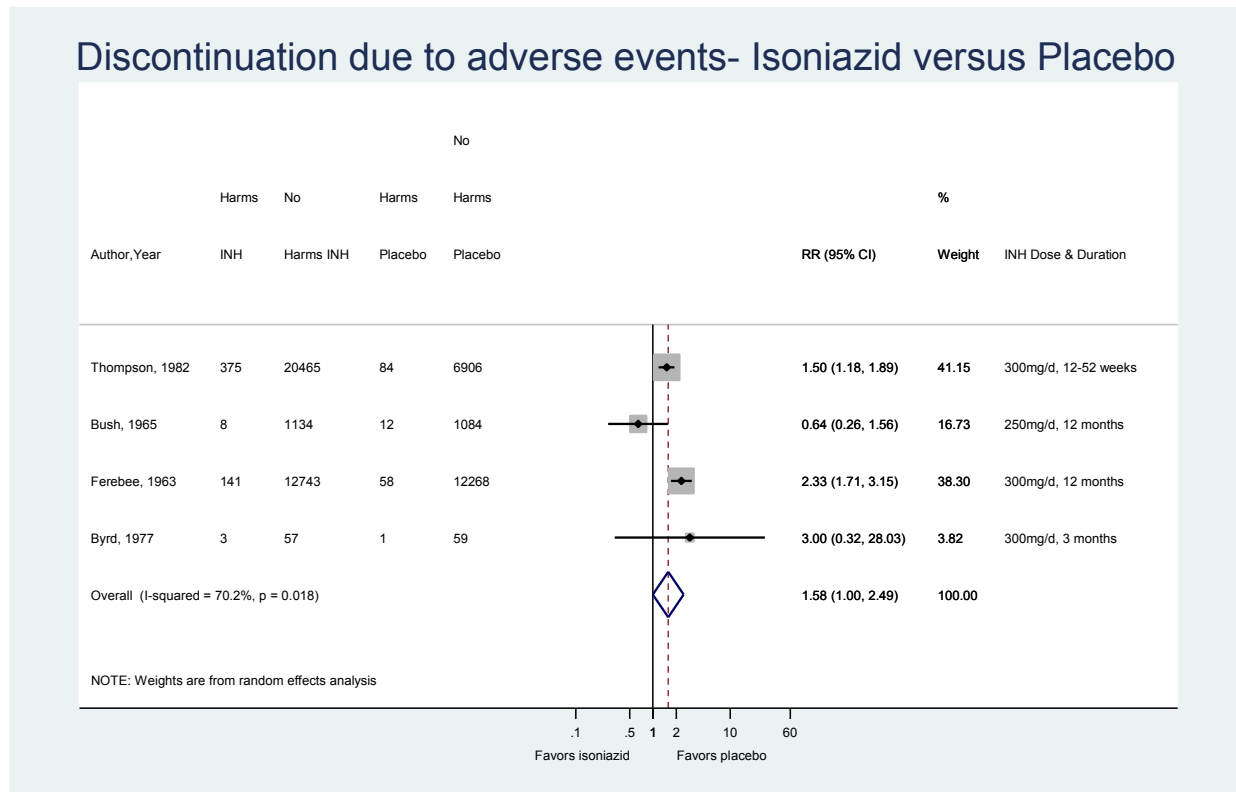
Appendix G Figure 1. Isoniazid Compared With Placebo, Relative Risk of Developing Hepatotoxicity: Sensitivity Analysis Including Data From Four Randomized, Controlled Trials



Notes: For Thompson, 1982 (IUAT trial), we included data from the 12-, 24-, and 52-week groups. A definition for hepatotoxicity (presented as “hepatitis” in this study) was not reported for this study. For Bailey, 1974, and Byrd, 1977, hepatotoxicity was defined as SGOT >100 mU/ml. For Falk, 1978, hepatotoxicity was defined only as “mild hepatitis.”

CI = confidence interval; INH= isoniazid; mg/d = milligrams per day; RR = relative risk.

Appendix G Figure 2. Isoniazid Compared With Placebo, Relative Risk of Treatment Discontinuation Due to Adverse Events: Sensitivity Analysis Including Data From Four Randomized, Controlled Trials



Notes: For Thompson, 1982 (IUAT trial), rates of discontinuation due to adverse events were reported only as a combined value across the three treatment duration groups (12-, 24-, and 52-week). For Bush, 1965, treatment discontinuation due to adverse events was categorized as gastrointestinal, rash, and other. For Byrd, 1977, treatment discontinuation was due to “symptomatology,” which included hepatotoxicity and mild nausea/abdominal cramps. For Ferebee, 1963, discontinuation due to adverse events corresponded to participants stopping medication due to being “sick” from pills.

CI = confidence interval; INH= isoniazid; mg/d = milligrams per day; RR = relative risk.